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| DOCUMENT NO: BSC/PATH/GDE/001 | REVISION NO: 21.6 |
| STANDARD REF: ISO 15189:2022, JCI – AOP .03 | DOCUMENT TYPE: POLICY & PROCEDURE |
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| APPROVED BY: DR. JUAN PINTO | PAGE 1 OF 203 |
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BON SECOURS HOSPITAL CORK

LABORATORY MANUAL

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1.0 INTRODUCTION

1.1 This manual is designed to give an overall view of the services available in the Pathology Department. It is intended as a quick reference guide for all Pathology users.

1.2 All Pathology services undergo continuous review through quality assurance and audit activities. The laboratory is committed to performing its activities in accordance with the requirements of the following regulations and standards:-

- The current version of the International Standard ISO 15189 titled “Medical Laboratories Particular Requirements for Quality and Competency”.
- The current version of the International Standard ISO 22870 titled "Point of Care Testing (POCT) – Requirements for Quality and Competence".
- Regulations and Statutory Instruments current versions including those relevant to Haemovigilance and Traceability, Infectious diseases, Safety and welfare at work, Data protection, Carriage of Dangerous goods and In vitro Diagnostic Medical devices. Current versions of regulations and statutory instruments are available in Q-Pulse”.

1.3 **This manual is intended for users of the Pathology Services both within the hospital, and those from outside agencies.**

1.4 Laboratory management are committed to:-

- ensuring staff are familiar with this policy and all other policies and procedures.
- seeking and acting on feedback from users of Laboratory Manual.
- staff recruitment, training, development and retention at all levels to provide a full and effective service to its users.
- the proper procurement and maintenance of such equipment and other resources as are needed for the provision of the service.
- the collection, transport and handling of all specimens in such a way as to ensure the correct performance of laboratory examinations.
- the use of accredited examination procedures and methods that will ensure the highest achievable quality of all tests performed.
- reporting results of examinations in ways which are timely, confidential, accurate and clinically useful.
- the assessment of user satisfaction, in addition to internal audit and external quality assessment, in order to produce continual quality improvement.
- the safe testing, distribution and transfusion of blood/ blood components.
- the operation and control of an effective point of care/ near patient testing service.
- having procedures in place to treat patients and samples with due care and with respect when samples are being processed within the laboratory.

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2.0 GUIDE TO USING THIS MANUAL

2.1 For internal users an **electronic version** of the manual is stored on the intranet. The document is stored in Adobe Acrobat format, which allows all computer users to read the document while preventing modification. Instructions on how to use the electronic version of the manual has been issued to Clinical Nurse Managers who have posted laminated instructions at workstations. Training programmes are in place to ensure all new clinical personnel are trained on the use of this manual. It is the responsibility of Clinical Nurse Managers to ensure that all staff are familiar with accessing the document. An e-mail is sent to all staff informing of an update to the manual and a list of changes. The manual is also accessible on the Bon Secours Website www.bonsecours.ie under Cork in the Pathology Section.

2.2 The laboratory tests and profiles you require information on can be located in the manual under sections 9 to 15 traceable to the department where the tests are performed.

3.0 GENERAL INFORMATION

3.1 Location of the Pathology Department

The Laboratory is located within the permanent facility of the Hospital grounds.



Pathology

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Please refer to the Bon Secours Health System website for parking information. [Please click here to access website.](#) If you need assistance, please contact security personnel who will assist you. Please note there are 2 designated parking spaces at the front of the Hospital for patients attending Phlebotomy. Please enter via the main door and follow signs for Blood Testing.

3.2 Pathology Department Opening Times

| Department/activity | Opening Hours |
|--|---|
| Pathology Reception | Monday to Friday 9.00am - 5.00pm |
| Phlebotomy Out-patient Service (By appointment only) | Monday to Thursday 8.00am - 4.15pm Friday 8.00am – 2.00pm |
| Phlebotomy In-patient Service | Monday to Friday from 7.00am - 4.30pm Saturday from 7.00am - 11.00am |
| Specimen Reception | Monday - Friday from 7.00am - 5.45pm Saturday 8.30am - 12.30pm |
| Routine Laboratory Blood (Biochemistry, Haematology, Blood Bank and Immunology) | Monday to Friday 7.00am – 6.00pm Saturday from 9.00am - 12.45pm |
| Routine Laboratory Microbiology | Monday to Friday 8.00am – 6.00pm Saturday from 9.15am - 12.45pm |
| Routine Laboratory Histopathology Service | Monday to Friday 8.00am – 6.00pm Saturday from 9.15am - 12.45pm |
| Emergency out of hours service (on call diagnostic service) | Monday to Friday, 6.00pm – 8.00am Saturday 12.45pm - Monday 8.00am Bank Holidays (24 Hours) |

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3.3 Availability of Clinical and Scientific Advice

3.3.1 Where **Medical and Scientific advice is required** on medical indications, appropriate selection of available testing procedures and queries with respect to Pathology test results, the Pathology Department welcomes your queries. For **telephone queries** use the provided listing.

| Department | Phone extension inside the Hospital | Phoning from outside the hospital |
|--|--|-----------------------------------|
| PATHOLOGY RECEPTION | 1717 | 021-4801717 |
| PHLEBOTOMY | 1733 | 021-4801733 |
| Head of Department – Natacha Rodrigues | | |
| PATHOLOGY OFFICE | 1720 | 021-4542005 |
| Head of Department – Amanda Long | 2197 | 021-4801720 021-4802197 |
| SPECIMEN RECEPTION | 1781 | 021-4801781 |
| Head of Department – Louise Linehan | 2261 | 021-4802261 |
| HAEMATOLOGY | 1722 | 021-4801722 |
| Chief Medical Scientist – Sarah O’Keeffe | 2264 | 021-4802264 |
| Consultant Haematologist – Dr. Eileen Kelleher | 1720 Pathology Office (Monday to Friday 9.15am – 5.30pm) | 021-4801720 |
| Consultant Haematologist – Dr. Susan O’Shea | | 021-4545899 (Cork Clinic) |
| Consultant Haematologist – Dr. Khalil Alnajjar | | |
| BLOOD BANK | 1721 | 021-4801721 |
| Chief Medical Scientist – Clare Stone | 1721 | |
| Consultant Haematologist – Dr. Eileen Kelleher | 1720 Pathology Office (Monday to Friday 9.15am – 5.30pm) | 021-4801720 |
| Consultant Haematologist – Dr. Susan O’Shea | | 021-4545899 (Cork Clinic) |
| Consultant Haematologist – Dr. Khalil Alnajjar | | |
| HAEMOVIGILANCE | 1659 | 021-4801659 085-2540151 |
| Haemovigilance Officers – Mary O’Sullivan and Fiona A. Murphy | | |
| HISTOPATHOLOGY including Diagnostic Cytology | 1727 | 021-4801727 |
| Chief Medical Scientist – Sandra Murphy | 1665 | 021-4801665 |
| Consultant Pathologist – Dr. Paul Ryan | 1967 | 021-4941967 086-6777487 |
| Consultant Pathologist – Dr. Aoife McCarthy | 1723 | 021-4801723 |

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| Department | Phone extension inside the Hospital | Phoning from outside the hospital |
|--|-------------------------------------|-----------------------------------|
| Consultant Pathologist – Dr. Triona Hayes | 1724 | 021-4801724 |
| Consultant Pathologist – Dr. Adeline Chelliah | 1931 | 021-4941931 |
| Consultant Pathologist – Dr. Juan Pinto | 1152 | 021-4861152 |
| Consultant Pathologist – Dr. Adeyemi Idowu | 1147 | 021-4861147 |
| IMMUNOLOGY | | |
| Senior Medical Scientist – Danny Brazil | 1997 | 021-4941997 |
| Consultant Immunologist – Prof. Conleth Feighery | N/A | 087-9969041 |
| BIOCHEMISTRY | | |
| Chief Medical Scientist – Emma Herbert | 1725 | 021-4801725 |
| | 1726 | 021-4801726 |
| Chemical Pathologist – Dr. Mike Louw | N/A | 086-8254528 |
| MICROBIOLOGY | | |
| | 1730/ 1731 | 021-4801730 021-4801731 |
| Chief Medical Scientist – Theresa Cunningham | 1948 | 021-4941948 021-4941948 |
| Consultant Microbiologist – Dr. Olive Murphy | 1759 | 021-4801759 086-0121136 |
| Consultant Microbiologist – Dr. Marianne Fraher | 1759 | 021-4801759 086-0121136 |
| Consultant Microbiologist – Dr. Deirdre O’Brien | 1950 | 021-4801950 086-0121136 |
| LABORATORY SERVICES MANAGER – Mary Kelly | 1748 | 021-4801748 |
| LABORATORY INFORMATION SYSTEM MANAGER – Miriam O'Donovan, Janet Sharkey | 1729 | 021-4801729 |
| QUALITY ASSURANCE – Berna Murray | 1960 | 021-4941960 |
| POINT OF CARE/ NEAR PATIENT MANAGER – Nicola Goulding | 2240 | 021-4542807 |

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3.3.2 List of Contacts for Out of Hours Service

The first point of contact out of hours is Hospital Reception 021 4801602.

| CONTACT | TELEPHONE NUMBER |
|--|--|
| Laboratory Consultant Pathologists | Number available at Hospital Reception |
| Laboratory Services Manager | Number available at Hospital Reception |
| Chief Medical Scientists | Number available at Hospital Reception |
| Hospital Scientific Staff | Number available at Hospital Reception |
| Irish Blood Transfusion Service Scientific Staff (24 hours) | 021 4807400 |
| Irish Blood Transfusion Service Crossmatch | 021 4807418 |
| Irish Blood Transfusion Service Despatch | 021 4807419 |
| Irish Blood Transfusion Service Medical Officers | 021 4807400 |
| Irish Blood Transfusion Service (Dr. Sorcha Ní Loingsigh) Consultant Haematologist | 021 4807400 |

3.3.3 Contacting Internal Bon Secours Laboratory Staff with an Urgent Request for Blood Out of Hours

3.3.3.1 Contact main reception and inform them that you require to speak with the on-call Blood Bank Laboratory staff. You must state that this request is urgent.

3.3.3.2 When the Laboratory staff makes contact with the nurse, the following details must be given:-

- Ward/ Location making the request
- The urgency of the request
- The patients details i.e. patient's name
- Proposed movement of the patient i.e. if the patient is to be relocated to perhaps Theatre or CCU
- The blood components/ tests requested
- State, if known, whether a blood sample for group and screen has already been reserved

3.3.3.3 “Emergency Stock” of group O Rh Negative Blood is always available in the Blood Bank refrigerator. It is the Consultant's decision to use the emergency stock or if it is safe to wait for the arrival of the Laboratory staff and the processing of the samples.

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3.4 **Bon Secours Hospital Website and Telephone Number**

Website: www.bonsecours.ie
Phone No.: 021- 4542807 (General Hospital Number)

3.5 **Laboratory Fees**

A list of Pathology charges is available on request from the Laboratory Services Manager.

3.6 **Staffing**

The Pathology department team consists of:-

- Laboratory Services Manager
- Clinical Directorate
- Consultant Pathologists
- Consultant Chemical Pathologist
- Consultant Microbiologists
- Consultant Haematologists
- Consultant Immunologist
- Scientific/ Non Scientific Heads of Department
- Laboratory Scientific Staff
- Support Services
 - Cleaning Contractors
 - Information Technology
 - Phlebotomy
 - Secretarial
 - Lab Quality Assurance
 - Facilities Management
 - Specimen Reception
 - Validation Technician
 - Haemovigilance Officer
 - Point of Care Nurse Specialists
 - Point of Care/ Near Patient Manager
 - Quality, Safety and Risk Dept.

3.7 **Accreditation Status**

- a. The Laboratory is accredited as a medical testing Laboratory to the International Standard ISO 15189, registration number 153MT (with the exception of Anti-Neutrophil Cytoplasmic Antibody (ANCA), Anti-Nuclear Antibody (ANA), Centromere, DNA Antibodies, Endomysial Antibodies IgA, Endomysial Antibodies IgG, Gastric Parietal Cell Antibodies, LKM Antibodies (Liver Kidney Microsomal Antibody), Mitochondrial Antibodies, Smooth Muscle Antibodies, Carba R Assay for CRE, ESBL, VRE, Syphilis RPR, Syphilis TPPA, all Chemical Pathology and Infectious Serology tests run and reported from the Abbott Alinity platforms.
Please refer to www.inab.ie.
- b. The Laboratory services are accredited by JCI (Joint Commission International) as part of the overall Hospital accreditation process.
- c. The Laboratory has been accredited as a training Laboratory by the Joint Committee for Biomedical Sciences for the in-service year for Medical Laboratory Scientist students.

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4.0 LABORATORY REQUEST FORMS, SPECIMEN BOTTLES AND CONTAINERS

4.1 General Information

This section deals with the **information** that is required to be documented on the laboratory **request form** and the **specimen bottle** or container, prior to the analyses of samples.

The laboratory has a number of different request forms. These are used for different Pathology analyses as outlined below. It is **important** that the correct form is supplied for a particular test.

1. **Bon Secours Hospital General Laboratory Request Form (PL001)** is used for most Biochemistry, Haematology and Immunology blood tests
2. **Bon Secours Hospital Microbiology Form (PL002)** is used for Urines, Swabs, Sputum, Blood Cultures and Faeces
3. **Bon Secours Hospital Blood Transfusion Form (PL003)** is used for transfusion requests
4. **Bon Secours Hospital Allergy Request Form (PL006)**
5. **Bon Secours Hospital Histopathology Request Form – Breast Biopsy (PL007)**
6. **Bon Secours Hospital Virology Request Form (PL009)**
7. **Bon Secours Hospital Histopathology Request Form** is used for Histopathology specimens (**Form no. 425**)
8. **Bon Secours Hospital Mantoux Form** (reference BSC/PHLE/SOP/009 Att. 7.1)
9. **Bon Secours Hospital CSF Request Form (PL012)**
10. **Inpatient Respiratory Investigations Request Form (PL015)**
12. **Bon Secours Hospital Test Request Form for UPMC Hillman Cancer Centre (PL017)**
13. **Bon Secours Hospital Pleural Fluid Request Form (PL019)**
14. **Bon Secours Hospital Haematology – Bone Marrow Clinic Request Form (PL021)**

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Note: Specific request forms are required for specialised testing processes that we do not perform in-house e.g. Chromosome Analysis/ DNA Genetic Screening. Please print off these forms as requested using the links provided in this manual, to accompany specimen(s) to external referral Laboratories.

4.2 Completing the Request Form

4.2.1 The following **essential** information must be documented in a **legible** manner on the request form including the **duplicate copy** where provided:-

1. Patient's **Hospital Number** (in-patients)
2. Patient's **Full Name** (Surname, Forename)
3. Patient's **Full Home Address**
4. Patient's **Date of Birth**
5. Patient's **Location** (Hospital Ward or room number). Where the requesting Physician is at an external location to that of the Bon Secours Hospital, Cork the postal address of the location should be included.
NOTE: The Cork Clinic is not considered an external location.
6. Patient's **gender**
7. The name of the **requesting Clinician**
8. **Specimen type** and **anatomical site** where appropriate (specifically Histopathology specimens and Microbiology swabs)
9. **Examination(s)** required
10. **Date and time of specimen collection in 24 hr format.** Note the Laboratory records the date and time specimens are received.
11. **Clinical information** appropriate to the test(s) requested must be supplied e.g. history of administration of drugs, blood transfusion history diagnosis etc.
12. Specific requirements for the Blood Transfusion Laboratory, reference PL003:-
 - If specific blood products are required i.e. CMV negative, irradiated, this should be requested.
 - The specific type and date of surgery and reason for a transfusion request should be documented on the transfusion form.
 - Blood transfusion specimens must have the **signature** of the person collecting and labelling the sample and also the signature of the person completing the request form i.e. two signatures, even if they are the same person.

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- The request form must contain the following minimum identification:-
 - Surname, first name
 - Hospital number
 - Date of birth
 - Signature of person taking the sample

Unlabelled samples or samples labelled with addressograph labels will be rejected.

13. Prioritization of Test Requests

- The Laboratory operates where practical a "first in, first out" policy with respect to the receipt, registration and processing of Haematology, Blood bank, Serology, Immunology, Biochemistry and Microbiology specimens with the exception of blood cultures, CSF and fluids. However, specific and clearly designated clinical areas (e.g. DOSA, CATH Lab, St Veronicas Day Ward, Oncology) may use a process that identifies to Specimen Reception personnel via the use of a coloured sticker that these identified specimens are deemed priority and are to be processed immediately upon receipt in Specimen Reception.
- The Laboratory operates an urgent/ routine process with respect to the receipt, registration and processing of Histopathology specimens. The urgent status applies to:-
 - Specimens for frozen section
 - Specimens marked urgent
 - Specimens of breast core biopsies
 - Specimens of muscle and nerve biopsies
 - Specimens of liver biopsies
 - Specimens of temporal arteries
 - Specimens of renal biopsies
 - Intrathecal specimens

14. The **signature** of the person **completing** the form (including Blood Bank form)

Note: Where addressograph labels are used to identify the above details, a check must be undertaken by the person completing the form to ensure the data is correct i.e. location or primary Physician.

4.2.2 Allergy Test Request Form Requirements (PL006)

Request for specific allergen testing will only be accepted for processing on the allergy request form PL006. A maximum of five allergens may be requested as per the listing on the back of the request form and should be based on clinical history. A referral letter on its own is not sufficient. Failure to do so will result in delays as the Laboratory cannot proceed without the correct request form.

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4.2.3 GP and External Requirements for Completing Request Forms/Referral Letter for Pathology Services

This section refers to GP requests and patients presenting to Laboratory outpatients. A request form/referral letter must accompany all requests for Pathology services. Failure to do so may result in patient delay as the Laboratory cannot proceed without test request documentation (request form/referral letter). Contact details (phone number) of the referring doctor must be provided on the request form. This is to ensure that the requesting doctor is contactable in the event of abnormal findings. Be advised that requests for specific allergen testing will only be accepted for processing on the allergy request form PL006. As the referring doctor, if you have any queries concerning the completion of the request form/referral letter, please do not hesitate to contact the Laboratory. For contact details, please refer to section 3.3 of this document.

4.3 Labelling the Specimen Container

4.3.1 The following **essential** information should be documented in a **legible** manner on the specimen container:-

1. Patient's full name
2. Date of birth
3. Hospital number (Blood Transfusion only)
3. Date and time of specimen collection
4. The **initials** of the person **collecting** the specimen (blood specimens only)
5. **Specimen type and anatomical site specifically for Histopathology, Non Gynaecological Cytology specimens and Microbiology swabs.**

Note 1: At a minimum two core identifiers must be used Name and Date of Birth.

Note 2: When requesting tests on **twins, patients** with the same surname in the same location or specimens requesting **blood grouping/ compatibility** testing services the following information is mandatory:-

1. Patient's full name
2. Date of birth
3. Hospital number (Blood Transfusion only)
4. Date and time of specimen collection

Blood transfusion samples must be handwritten and labelled, dated and signed by the person taking the sample. Unlabelled blood transfusion samples or samples labelled with addressograph labels will be rejected.

Note: All patient samples must be labelled at the bedside. (This is to prevent misidentification and labelling errors).

Sample labelling is performed in accordance with Nursing Policy ORG0001 titled "Patient Identification".

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Specimens from selected Hospital locations may be labelled with labels printed at the patient's bedside using the Blood Track TX software. Labels will have the following information:-

- | | | |
|--|---|---|
| 1. Patient's full name | } | Obtained from 2D barcode on patient's wristband |
| 2. Hospital number | | |
| 3. Date of birth | | |
| 4. Sex | | |
| 5. Collectors/ staff members name (obtained from staff ID badge) | | |
| 6. Location | | |
| 7. Date and time of collection | | |

4.3.2 Instructions for 24 Hour Urine Collection

Instructions for Ward Collection

- Please obtain a bottle for this collection from the Laboratory. This will contain appropriate preservatives if required.
Caution: These may be acid or base depending on the test, so care should be taken when handling.

Instructions for Collection Provided to Patients

- The container may contain a preservative. This must not be washed out and care should be taken when filling the container as the preservative will be either acid or alkali.*
- At a convenient time, e.g. 8am, void the bladder. This specimen is discarded. From this time until the same time the following day, **all** urine passed must be collected in the container.*
- Once the collection is complete, it must be brought to the Laboratory and left in Specimen Reception during normal working hours or in the fridge if outside normal working hours.*
- If any of the urine is discarded during collection, the collection must be restarted. A new container will have to be obtained from the Biochemistry department.*

A copy of this information is provided on each 24 hour container.

4.4 Addressograph Labels on Specimen Bottles

4.4.1 Patient details must be handwritten on blood specimen bottles except as defined in section 4.3.1.

Addressograph labels are not permitted on any blood specimen bottles.

Addressograph labels are permitted on all Microbiology, Histopathology and Cytopathology specimens.

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4.5 Quality of Blood Specimens

4.5.1 Laboratory personnel must inspect prior to testing each blood specimen received for:-

- Evidence of Haemolysis
- Gross Lipaemia
- Presence of clots in all specimens requesting full blood count and coagulation tests
- For coagulation tests, specimens that are under filled or over filled cannot be analysed.

For other specimens, the integrity of the specimen is inspected i.e. broken cytology slides) etc.

In such instances, a **second specimen** may be requested or the **issued report** will have an appended comment noting the presence of haemolysis, lipaemia or clots as appropriate.

4.6 Non-Conforming Specimen Bottles, Forms or Specimen Quality Issues

4.6.1 Where the requirements with respect to labelling the request form and specimen container or specimen quality issues are not met the following will apply.

| SPECIMEN ISSUES | REQUIRED ACTION |
|--|--|
| <ul style="list-style-type: none"> • No specimen received. | <ul style="list-style-type: none"> • Specimen Reception inform the clinical area and request a 2nd specimen |
| <ul style="list-style-type: none"> • Specimen site not identified on form or specimen • Specimen collected at incorrect time. | <ul style="list-style-type: none"> • Clinical area identifies the correct site on specimen • Ward contacted by Specimen Reception to provide correct time |
| <ul style="list-style-type: none"> • Two of the three mandatory unique identifiers are not correct or absent from the specimen (Full name, DOB, hospital no). | <ul style="list-style-type: none"> • Request second specimen or the ward staff will accept responsibility for same in emergency cases or where the specimen cannot be replaced. • If tested the report will show the non-conforming event. |

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| SPECIMEN ISSUES | REQUIRED ACTION |
|---|---|
| <ul style="list-style-type: none"> Addressograph label on Blood Bank tube(s) or unlabelled Blood Bank tube(s). | <ul style="list-style-type: none"> Blood transfusion samples which are unlabelled or labelled with an addressograph label will be rejected and a new sample requested. |
| Addressograph labels on blood specimen other than Blood Transfusion with the exception of selected Hospital locations that use the Blood Track TX software for the generation of bedside printed labels | <ul style="list-style-type: none"> Clinical staff may correct the error by removing the addressograph label and handwriting the patient details on the blood specimen or a 2nd specimen is collected. The clinical staff member rectifying the issue will be identified on the test report. |
| <ul style="list-style-type: none"> Specimens unlabeled (other than Blood Bank). | <ul style="list-style-type: none"> The specimen should be repeated where possible. Specimen Reception staff will phone for a repeat. If the specimen is irreplaceable and a repeat cannot be obtained, ward staff will take responsibility by relabelling the specimen. The clinical staff member rectifying the issue will be identified on the test report. |
| FORM ISSUES | REQUIRED ACTION |
| <ul style="list-style-type: none"> No request form provided with specimen. | <ul style="list-style-type: none"> Clinical areas/ ward to provide form |
| <ul style="list-style-type: none"> Demographic details on the form and specimen do not match. | <ul style="list-style-type: none"> Clinical area/ ward updates the form with the correct information |
| <ul style="list-style-type: none"> Required times missing from the form e.g. sample collect time | <ul style="list-style-type: none"> Clinical area/ ward updates the form with the correct information |
| QUALITY ISSUES | REQUIRED ACTION |
| <ul style="list-style-type: none"> Specimen grossly haemolysed Specimen gross Lipaemic Specimen too old | <ul style="list-style-type: none"> A second specimen is to be collected from the patient |

Please note the Laboratory staff document all non conforming events electronically.

- 4.6.2** It is important to note the Laboratory cannot process specimens with non conforming issues until they are resolved by clinical staff. Undue delay in correcting issues may result in a requirement to take a repeat specimen. The Pneumatic Tube System may be used to transport non conforming specimens and/ or forms to and from the clinical area for the speedy resolution of issues.

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4.7 **Further Additional Testing**

If on sending a specimen for testing and **further additional testing** is required, please contact Specimen Reception to investigate the feasibility of using the initial specimen for analysis, as age of specimen may impact on the validity of test results. **A request form or equivalent must accompany such requests as the Pathology department cannot process additional test requests until the request form is provided.** Refer to section 6.2 of the current version of the procedure BSC/SR/SOP/002 titled "Procedure for the Receipt, Checking, Computer Registration, Secondary Processing Including Labelling and Distribution of Pathological Specimens".

4.8 **Non-Conforming Issues and Credit**

It is the policy of the Pathology department to credit the patient's account where non-conforming issues lead to the non testing of specimens.

5.0 **DELIVERY, PACKING, TRANSPORT AND POSTAL REQUIREMENTS FOR DIAGNOSTIC AND INFECTIOUS (OR SUSPECTED INFECTIOUS) SPECIMENS**

5.1 **General Information**

It is the policy of the Pathology Department to treat all specimens and samples as potentially infectious or high risk. Therefore, we advise you to take universal precautions in the collection, packaging and the delivery of specimens being sent to the Pathology Department for analysis.

5.1.1 Note routine specimens (where testing will be performed the following day) collected and delivered to the Laboratory during the out of hours period will result in an increase in the turnaround time for the test.

5.2 **Specimen Delivery From Within the Hospital**

- During the **routine** Pathology opening hours **blood specimens** will be **taken** by the **Phlebotomy** team.
- **Outside routine Pathology opening hours blood specimens** will be taken by either medical doctors or nurses on the ward.
During routine hours, the Pneumatic Tube System is routinely used for the transport of specimens from all ward/ outpatient locations to the Laboratory. Instructions on the correct use of the Pneumatic Tube System are posted on each station, reference BSC/SR/SOP/007.
- Specimens being sent to the laboratory via the Pneumatic Tube System should be placed in a plastic sample bag. The sample bag may or may not be attached to the form. This depends on the form type. Specimens that are permitted for transport within the Pneumatic Tube System are placed in a red carrier and transported to Specimen Reception as per procedure BSC/SR/SOP/007.

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- It is the responsibility of Specimen Reception staff to clock in the date and time of the specimen using the date and time clock located in Specimen Reception for specimens that are delivered via the Pneumatic Tube System.

5.2.1 Procedure for the Out of Hours Delivery and Storage of Specimens to Pathology

5.2.1.1 Emergency out of hours specimens are delivered by the Pneumatic Tube System.

5.2.1.2 It is the responsibility of the Clinical Nurse Managers to ensure non-urgent specimens are delivered to Pathology immediately following collection. **Non-emergency** specimens received out of hours are to be stored as follows:-

1. **24 hour urines** are stored on the bottom shelf of the fridge opposite the Specimen Reception door.
2. In **fridge** labelled “**Non-Emergency Specimens Over Night Storage**” the following specimens are stored in designated labelled trays:-
 - urines (universal specimen) and 24 hour urine specimens
 - faeces specimens
 - swab specimens
 - sputum specimens
 - blood specimens (but not blood cultures)
 - diagnostic cytology
 - miscellaneous

This fridge is located opposite the Specimen Reception door.

3. In press labelled “**Histopathology Specimens Only**”, Histopathology specimens are stored in appropriate containers with fixative. This press is beside the Specimen Reception door.
4. It is the responsibility of the **person who delivers** routine specimens out of hours to the Pathology department to clock the date and time of the specimen using the time and date clock located in the Pathology foyer. **Under no circumstances can the Pneumatic Tube System be used to deliver routine specimens to the Laboratory during the out of hours period as the Specimen Reception department is not staffed.**

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5.3 **Specimen Delivery from Outside of the Hospital**

The requirements stated below apply to all specimens or samples directed to the Pathology Department. These will be required to be packed and transported in accordance with the European Agreement concerning the International Carriage of Dangerous Goods by Road (UNADR). Posted samples are not suitable for urea and electrolytes, coagulation, phosphate, AST, LDH, glucose (unless in a fluoride tube), magnesium, hormone assays and tumour marker assays.

It is the policy of the Pathology department to provide our customers with specimen transport packaging materials. Please do not hesitate to ask us for your supply.

5.3.1 **Packing Procedure for the Transport of Diagnostic Specimens (Non Infectious)**

1. Specimen to be sent should be stored in a secure (preferably plastic) primary container.
2. Place primary sample into secondary receptacle (hard white container supplied by BSH) with absorbent material.
3. This will be placed in a 95kPa specimen transport bag.
4. Place the specimen bag with the sample in a padded (jiffy bag) envelope or a Surepath Solution Specibox.
5. Label the envelope with a hazard warning label, "Diagnostic Specimen, Category B, UN 3373".
6. Place the name, address and contact number of the destination laboratory and the sending Laboratory on the outside of the envelope/ specibox.
7. The specimen can be transported as appropriate.

There is no requirement for a licensed courier to transport non infectious diagnostic samples.

5.3.2 **Procedure for the Transport of Infectious or Suspected Infectious Specimens**

When sending specimens or samples suspected or known to contain infectious or suspected infectious substances, the Microbiology and Infection Control departments should be contacted for advice regarding transport.

5.4 **Disposal of Waste Material Used in Specimen Collection**

All materials used in specimen collection should be treated as potentially hazardous and discarded using sharps containers and other appropriate colour coded bags. Do not hesitate to contact the Laboratory for appropriate disposal advice.

5.5 **Storage of Examined Specimens for Archive and Look Back Purposes**

Specimens post examinations are retained for defined periods depending on the specimen type. If you require further details, please contact the Pathology department. Contact details are provided in section 3.3 of this document titled "Availability of Clinical and Scientific Advice".

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6.0 EXTERNAL THIRD PARTY ASSESSMENT PROGRAMME

- 6.1 The Pathology Department **participates** in relevant available **external third party assessment schemes**. We use these independent third party programmes to verify the accuracy of the test procedures used. Results of these assessments are reviewed regularly by clinical Pathology personnel.

7.0 PROVISION OF SERVICES

| SERVICE | DESCRIPTION |
|---|---|
| Provision of Diagnostic Services | There is a wide range of Pathology tests available. For the full list of diagnostic services refer to chapters 9 to 16 of this document. |
| Biochemistry | The automated chemistry service provides analysis of samples for renal, liver, cardiac, lipid, iron studies and specific protein assays. The Immunology section performs Autoimmune, Serum Protein Electrophoresis (plus Immunofixation if necessary based on review of Electrophoresis), Urine Immunoelectrophoresis, allergy and associated assays while the Immunoassay section performs endocrine, tumour marker, troponin and therapeutic drug monitoring (TDM) assays. |
| Haematology | A diagnostic Haematology service is provided which includes blood counts and blood film examination. Routine coagulation screening includes PT-INR, APTT and Fibrinogen tests. |
| Blood Bank | The Hospital Blood Bank provides routine and emergency Blood Grouping, Antibody Screening and compatibility testing. A supply of blood products is also available, refer to the Hospital Transfusion Handbook for details or contact the Blood Transfusion department. |
| Serology | A comprehensive Serology service is provided including immunity status testing. |
| Histopathology | The Histopathology department provides a varied range of services including Tissue Pathology (including frozen sections for rapid diagnosis) and Diagnostic Cytology . For outpatient specimens collected outside the confines of the Bon Secours Hospital, Cork, a signed Service Level Agreement must be in place between the Pathology Department and the referring Doctor/Dentist/Service. |
| Immunology | The Immunology department provides a diagnostic testing service, specifically in relation to facilitating the differential diagnosis of autoimmune diseases. |

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| SERVICE | DESCRIPTION |
|---|--|
| Microbiology | The Laboratory examines a diverse range of specimens for bacterial, fungal, viral and parasitic infections and determines the sensitivity of bacteria to antibiotics. The department provides a clinical service which ensures that patients are treated in a timely and effective manner. The department works closely with the Hospital Infection Prevention and Control Department. |
| Related Diagnostic Services | Glucose tolerance test (refer to section 9.5) and mantoux testing. Mantoux testing is performed on the ward and ordered by the doctor using form BSC/PHLE/SOP/009 att. 7.1 which is available from the postroom. All necessary information is available and recorded here. |
| Phlebotomy Service | The Phlebotomy department on a routine basis 7.00am - 4.30pm Monday to Friday and Saturday from 7.00am - 11.00am take blood samples for diagnostic testing from in-patients. Outside these hours, blood samples are taken by trained Hospital personnel. Outpatient Phlebotomy is from 8.00am – 4.15pm Monday to Thursday 8.00am – 2:00pm Friday. |
| Consultant Advisory Services | Consultant Pathology advisory services are available in the following specialities, Histopathology, Immunology, clinical Microbiology, Haematology, Blood Transfusion and Biochemistry. For further details, refer to section 3.2 of this manual. |
| Warfarin Clinic | An outpatient Warfarin clinic service is available on Tuesdays between 10.00 am and 12.00 pm in the Phlebotomy department. This testing service should be pre booked by contacting 021-4801720 or 4802197 as it is by appointment only. |
| Therapeutic Phlebotomy | The therapeutic Phlebotomy procedure is available for the treatment of certain clinical conditions such as Haemochromatosis and Myeloproliferative Disorders (polycythaemia). This process is performed by Phlebotomy staff under the clinical guidance of a Consultant Gastroenterologist (with special interest in Liver Disorders) for Haemochromatosis and Consultant Haematologists for Myeloproliferative Disorders (polycythaemia). This testing service should be pre booked by contacting 021-4801720 or 4802197 for Myeloproliferative Disorders (polycythaemia) or 021-4344714 (Consultant Gastroenterologist Clinic) for Haemochromatosis as services are by appointment only. |
| Health Check and Wellness Centre Screening | To meet customer requirements a full range of pathology test packages are available as part of health screening. These packages are designed to cover any combination of pathology tests as required by the customer. |

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| SERVICE | DESCRIPTION |
|--|--|
| Haemovigilance Service | All Haemovigilance reactions and events are documented and reported to the National Haemovigilance Office as per the requirements of the Hospital Transfusion Booklet. The Bon Secours Pathology Department is committed in conjunction with the Haemovigilance Officer, Haemovigilance to providing a reporting mechanism that assists the Quality Management Review Process. A Hospital Transfusion Committee is in operation and its membership includes Medical, Scientific and Nursing staff and also includes the Regional Director of the Irish Blood Transfusion Service or nominee. The committee meets regularly and discusses and advises on transfusion policies, inventory management, quality issues, haemovigilance and traceability. |
| Point of Care/ Near Patient Testing Service | A point of care/ near patient testing service is in place that meets clinical requirements as defined by the POC Committee. The POC Committee reports to Hospital/ Laboratory Management for point of care testing services/ issues. This POC Committee meets on a quarterly basis and if clinically required. The POC meeting is chaired by the Point of Care/ Near Patient Manager. |

8.0 LABORATORY TESTS/ PROFILES AVAILABLE

8.1 This section outlines the tests that are available in the different Pathology Laboratories. These tests will be described under the following **disciplines/ sections:-**

- **Biochemistry**
- **Blood Transfusion**
- **Haematology**
- **Histopathology/ Cytology**
- **Immunology**
- **Microbiology** including Serology and Virology
- **Chromosome Analysis**

8.2 Laboratory Test/ Profile Description

Each laboratory test will be described under the following headings:-

- **Test Name**
- **Specimen type/ site**
Where the **specimen is blood** and the required additive is stated as none, the requirement should be interpreted as a **clotted sample**.
- **Specimen requirements** including additive, required specimen volume and container type.

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- **Special Requirements**

The special requirements column defines for each diagnostic test if (applicable) the following:-

- Patient preparation, e.g. fasting
- Consent form
- Special timing for collection of samples e.g. pre and post drug administration
- Any special handling needs between time of collection and time received by the laboratory (transport requirements, refrigeration, warming, immediate delivery etc.)

- **Turnaround Time**

Turnaround time is defined as the time **from receipt of specimen** in the Pathology department to the time **authorized results are electronically available**. In this document, turnaround time is defined as hours or working days. Working days denotes Monday to Friday and does not include out of hour periods including weekends and bank holidays.

Occasionally, the Laboratory may be unable to meet the defined turnaround time as specified in this document for tests that are routinely performed in-house, e.g. or where a second opinion is required to confirm a Histopathology final diagnosis. Where delays in reporting results are encountered and which may compromise patient care, the Laboratory will communicate such delays to appropriate clinical personnel.

8.3 **Repeat Examination due to Analytical Failure or Further Examination of the Primary Specimen**

8.3.1 **Repeat Examination due to Analytical Failure**

It is the policy of the Pathology department in the event of an analytical failure to:-

- Repeat the test using a back-up instrument.
- or
- Store the specimens in appropriate conditions until the cause of the analytical failure is identified and corrected and then repeat the test. The urgency of the outstanding specimens is reviewed by the relevant laboratory Consultant Pathologist or nominee.

8.3.2 **Further Examination of the Primary Specimen**

Where further testing is relevant to the investigation or diagnosis of the condition or symptoms which gave rise to the original test request then it is the policy of the Pathology department to pursue a diagnosis by performance of additional tests using the primary specimen.

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8.4 **Tests Not Listed**

If you require a diagnostic test that is not listed, please contact the Pathology department who will endeavour to **outsource** as appropriate your test requirement. The Pathology Department (Scientific/QA/Consultant staff) evaluate and select referral Laboratories and Consultants who provide second opinions based on objective criteria set out in Laboratory Policy. There are no outside sources of Laboratory services owned or recommended by referring physicians.

8.5 **External Laboratory Testing**

Some specimen/ samples are referred to external laboratories for testing. These will be recognised by the presence of an **asterisk *** after the test name.

8.6 **Emergency Out of Hours Service**

Tests provided out of hours in this service will be recognised by the presence of this symbol ♦ in the turnaround time column. A master list of tests performed out of hours is defined in section 25.0 of this document. If any other test is required, the person requesting the test should contact the relevant Laboratory Medical Consultant.

8.7 **Instructions on the Collection of Specimens**







Instructions on the collection of specimens are detailed in the special requirements section applicable to each test as detailed in Chapters 9 to 16 and or in the following Hospital policies and Laboratory procedures:-

| Specimen Type | Document Name | Document Number |
|----------------------|--|------------------------|
| Blood | Procedure for Venepuncture of In-Patients | BSC/PHLE/SOP/001 |
| Blood | Procedure for Venepuncture of Out-Patients by Phlebotomy Staff | BSC/PHLE/SOP/003 |
| Urine | Sterile Urine Samples from Infants and Toddlers | CW0010 |
| Urine | Collection of Mid Stream Urine for Culture and Sensitivity | NUR0179 |
| Urine | Collection of Urine from an Ileal Conduit | STOMA0001 |
| Urine | Performing Routine Urinalysis on Inpatients | NUR0052 |

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9.0 BIOCHEMISTRY TESTS

9.1 Endocrinology












| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|---|--------------------------|--|---------------------------|-----------------------|---|-----------------|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| 11 Deoxycortisol* | Blood | None  | 4.9 | Blood Tube | None | 15 working days |
| 17 Hydroxy Progesterone* | Blood | None  | 4.9 | Blood Tube | None | 15 working days |
| ACTH*† Adrenocorticotrophic Hormone | Plasma | Potassium EDTA and aprotinin which is added to tube by Specimen Reception before blood is taken  | 7.5 | Blood Tube x 2 | Freeze plasma immediately after specimen collection. Obtain tubes from lab before taking blood. | 15 working days |
| ADH*† Anti Diuretic Hormone | Blood | Potassium EDTA and aprotinin which is added to tube by Specimen Reception before blood is taken  | 7.5 | Blood Tube | Freeze plasma within 1 hr of specimen collection | 20 working days |
| Aldosterone* | 24 hour urine collection | None | 100 | 5L Urine container | Refer to section 4.3.2 | 15 working days |
| Aldosterone*† | Plasma | Potassium EDTA  | 1 | Blood Tube | Freeze serum within 4 hrs of specimen collection. State whether patient is standing or lying (after 8 hrs rest) | 15 working days |
| Androstenedione* | Blood | None  | 4.9 | Blood Tube | None | 20 working days |

* These specimens/ samples are referred to external laboratories for testing.

† These tests require special patient preparation or conditions. For clarification purposes, please contact Biochemistry department before taking sample.

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BIOCHEMISTRY TESTS - Endocrinology.....cont'd

| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|--|------------------------|---|---------------------------|-----------------------|--|----------------------------|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| Anti-Mullerian Hormone (AMH)* | Blood | None  | 4.9 | Blood Tube | Freeze serum within 4 hrs of specimen collection. Do not use haemolysed specimen | 20 working days |
| Calcitonin*† | Blood | None  | 4.9 | Blood Tube | Freeze serum within 4 hrs of specimen collection | 15 working days |
| Cortisol (30 mins post synacthen) | Blood | None  | 4.9 | Blood Tube | Take exactly 30 min post synacthen | 1 working day Mon - Fri |
| Cortisol (60 mins post synacthen) | Blood | None  | 4.9 | Blood Tube | Take exactly 60 min post synacthen | 1 working day Mon - Fri |
| Cortisol (am) | Blood | None  | 4.9 | Blood Tube | Patient must be resting, take between 7 & 9 am | 1 working day Mon - Fri |
| Cortisol (midnight) | Blood | None  | 4.9 | Blood Tube | Take at midnight | 1 working day Mon - Fri |
| Cortisol (pm) | Blood | None  | 4.9 | Blood Tube | Take between 3 and 5pm | 1 working day Mon - Fri |
| Cortisol (pre synacthen) | Blood | None  | 4.9 | Blood Tube | Patient must be resting | 1 working day Mon - Fri |
| Cortisol (random) | Blood | None  | 4.9 | Blood Tube | None | 1 working day Mon - Fri |
| Cortisol Urinary* | 24 hr urine collection | None | 50 | 5L Urine container | Refer to section 4.3.2 | 15 working days |
| Dehydro Epiandrosterone DHEA* | Blood | None  | 4.9 | Blood Tube | None | 15 working days |
| Dehydro Epiandrosterone DHEA (Urinary)* | Urine | None | 100 | 5L Urine container | Refer to section 4.3.2 | 15 working days |
| Dehydro Epiandrosterone Sulphate DHEAS* | Blood | None  | 4.9 | Blood Tube | None | 15 working days |











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BIOCHEMISTRY TESTS - Endocrinology.....cont'd







| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|--|---------------|--|---------------------------|-----------------------|---|----------------------------|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| Dihydrotestosterone DHT* | Blood | None  | 4.9 | Blood Tube | None | 15 working days |
| Erythropoietin* | Blood | None  | 4.9 | Blood Tube | None | 15 working days |
| Ferritin | Blood | None  | 4.9 | Blood Tube | None | 1 working day Mon - Fri |
| Folate (Folic Acid) | Blood | None  | 4.9 | Blood Tube | None | 1 working day Mon – Fri |
| Follicle Stimulating Hormone (FSH) | Blood | None  | 4.9 | Blood Tube | None | 1 working day Mon - Fri |
| Free T₃ (Triiodothyronine) | Blood | None  | 4.9 | Blood Tube | None | 1 working day Mon - Fri |
| Reverse T₃* | Blood | None  | 4.9 | Blood Tube | Separate and freeze within 4 hours | 25 working days |
| Free T₄ (Thyroxine) | Blood | None  | 4.9 | Blood Tube | None | 1 working day Mon - Fri |
| Gastrin*† | Blood | None  | 4.9 | Blood Tube | Freeze serum within 1 hr of specimen collection. Patient should be fasting a minimum of 10 hrs prior to collection. | 25 working days |
| Glucagon*† | Blood | Potassium EDTA  and aprotinin which is added to tube by Specimen Reception before blood is taken | 7.5 | Blood Tube x 2 | Freeze plasma within 1 hr of specimen collection. Obtain tubes from lab before taking blood. | 30 working days |

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† These tests require special patient preparation or conditions. For clarification purposes, please contact Biochemistry department before taking sample.

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BIOCHEMISTRY TESTS - Endocrinology.....cont'd

| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|---|---------------|---|---------------------------|-----------------------|--|----------------------------|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| Growth Hormone*† | Blood | None  | 4.9 | Blood Tube | Freeze serum within 4 hrs of specimen collection. | 15 working days |
| hCG Serum | Blood | None  | 4.9 | Blood Tube | Refer to section 17.0 for recommended use of hCG in Pregnancy testing. | ♦ 120 mins ϕ |
| Insulin*† | Blood | None  | 4.9 | Blood Tube | Freeze serum within 30 mins of specimen collection. State whether patient is fasting or post prandial. | 15 working days |
| Insulin C-peptide* | Blood | None  | 4.9 | Blood Tube | Separated and frozen in <1 hour. State whether patient is fasting or post prandial. | 15 working days |
| LH (Luteinising Hormone) | Blood | None  | 4.9 | Blood Tube | None | 1 working day Mon – Fri |
| Monomeric Prolactin (Macroprolactin) | Blood | None  | 4.9 | Blood Tube | Contact Biochemistry for information | 1 working day |

- * These specimens/ samples are referred to external laboratories for testing.
- † These tests require special patient preparation or conditions. For clarification purposes, please contact Biochemistry department before taking sample.
- ♦ Tests provided in the emergency out of hours service.
- ϕ Note: 95% of results reported for this test will achieve the turnaround time as stated.








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| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|--------------|---------------|---|---------------------------|-----------------------|---|-------------------------|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| Oestradiol | Blood | None  | 4.9 | Blood Tube | This assay should NOT be used to assess Oestradiol levels for patients undergoing Fulvestrant or Mifepristone treatment. Structural and functional analogues of steroid hormones, including the Oestradiol molecule, have the potential to cause interference/cross reactivity with the Alinity i Oestradiol assay. Samples from patients administered medications which inhibit tumour cell proliferation (e.g. CDK 4/6 inhibitors) may be subject to interference/cross reactivity with the Alinity i Oestradiol assay. In addition, drugs which interfere with or activate production of steroid hormones (e.g. Aromatase inhibitors) may also interfere or cross react with the Alinity i Oestradiol assay. In such cases, an alternate method such as chromatography should be used. | 1 working day Mon - Fri |

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| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|-------------------------------------|--------------------------|--|---------------------------|-----------------------|--|-------------------------|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| Pancreatic Polypeptide PPP*† | Blood | None  | 4.9 | Blood Tube | Freeze serum within 1 hr of specimen collection | 30 working days |
| Parathyroid Hormone (PTH) | Blood | Potassium EDTA  | 2.7 | Blood Tube | Freeze plasma if not analysed within 8 hrs. | 1 working day Mon - Fri |
| Pregnanetriol * | 24 hour urine collection | None | 700 | 5L Urine container | Refer to section 4.3.2 | 15 working days |
| Progesterone | Blood | None  | 4.9 | Blood Tube | None | 1 working day Mon - Fri |
| Proinsulin* | Blood | None  | 4.9 | Blood Tube | Bring to lab immediately, must be frozen in <4 hrs | 15 working days |
| Prolactin | Blood | None  | 4.9 | Blood Tube | None | 1 working day Mon - Fri |
| Renin*† | Blood | Potassium EDTA  | 2.7 | Blood Tube | EDTA plasma frozen <4 hrs. State whether patient is standing or lying (after 8 hrs rest) | 15 working days |
| Serotonin*† | Blood | Lithium Heparin  | 9 | Blood Tube | For 48 hrs before collection, the patient should not drink or eat: bananas, chocolate, tomatoes, grapefruit, nuts, avocado, pineapple, plums, citrus fruit, tea and coffee. Freeze whole blood within 1 hr of specimen collection | 15 working days |

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| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|--|---------------|---|---------------------------|-----------------------|---|-------------------------|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| Sex Hormone Binding Globulin SHBG* | Blood | None  | 4.9 | Blood Tube | None | 15 working days |
| Somatostatin* | Blood | Potassium EDTA and Aprotinin  | 2.7 x 2 | Blood Tube x 2 | Bring to lab immediately, must be frozen in <1 hr | 25 working days |
| Testosterone with SHBG* | Blood | None  | 4.9 | Blood Tube | None | 15 working days |
| Testosterone (Free) | Blood | None  | 4.9 | Blood Tube | None | 20 working days |
| Testosterone (Urinary)* | Urine | None | 100 | 5L Urine container | Refer to section 4.3.2 | 15 working days |
| Thyroid Stimulating Hormone | Blood | None  | 4.9 | Blood Tube | None | 1 working day Mon - Fri |
| Vasoactive Intestinal Peptide (VIP)*† | Blood | Potassium EDTA to which Aprotinin has been added to tube by Spec. Reception before blood is taken.  | 7.5 | Blood Tube | Freeze plasma within 1 hr of specimen collection | 25 working days |












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† These tests require special patient preparation or conditions. For clarification purposes, please contact Biochemistry department before taking sample.

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9.2 BIOCHEMISTRY TESTS - Tumour Markers

| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|--|---------------|---|---------------------------|-----------------------|--|----------------------------|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| Alkaline Phosphatase (Placental)* | Blood | None  | 4.9 | Blood Tube x 2 | None | 15 working days |
| Alpha Feto Protein (AFP) | Blood | None  | 4.9 | Blood Tube | None | 1 working day Mon - Fri |
| CA15-3 | Blood | None  | 4.9 | Blood Tube | None | 1 working day Mon - Fri |
| CA19-9 | Blood | None  | 4.9 | Blood Tube | None | 5 working days |
| CA-50* | Blood | None  | 4.9 | Blood Tube | None | 15 working days |
| CA72-4* | Blood | None  | 4.9 | Blood Tube | None | 15 working days |
| CA125 | Blood | None  | 4.9 | Blood Tube | None | 1 working day Mon - Fri |
| CYFRA 21-1* | Blood | None  | 4.9 | Blood Tube | None | 15 working days |
| Carcinoembryonic Antigen (CEA) | Blood | None  | 4.9 | Blood Tube | None | 1 working day Mon – Fri |
| hCG Total (Serum Quantitative as Tumour Marker) | Blood | None  | 4.9 | Blood Tube | Please provide spot urine sample in addition to blood sample when hCG is being used as tumour marker | 1 working day Mon - Fri |
| Inhibin B*† | Blood | None  | 4.9 | Blood Tube | Freeze serum within 1 hr of specimen collection | 20 working days |
| Neuroblastoma screen*† | Spot Urine | None | 20 | Sterile Universal | Freeze urine within 1 hr of specimen collection | 15 working days |

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9.2 BIOCHEMISTRY TESTS - Tumour Markerscont'd

| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|---|---------------|---|---------------------------|-----------------------|--|-----------------------------|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| Prostate Specific Antigen (PSA) | Blood | None  | 4.9 | Blood Tube | Serum to be separated within 4 hrs of collection | 1 working day Mon - Fri |
| Prostate Specific Antigen (PSA Free) † | Blood | None  | 4.9 | Blood Tube | Serum to be separated within 3 hrs of collection | 4 working days Mon - Fri |
| Thyroglobulin* | Blood | None  | 4.9 | Blood Tube | None | 15 working days |










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† These tests require special patient preparation or conditions. For clarification purposes, please contact Biochemistry department before taking sample.

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9.3 **BIOCHEMISTRY TESTS - Therapeutic Drug Monitoring**

9.3.1 All drugs must be requested **using generic, not trade names** to ensure clarity. Please refer to the British National Formulary (BNF) which can be accessed by clicking on the following link www.medicinescomplete.com/mc/bnf/current/index.htm.









| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|--------------------------------------|---------------|---|---------------------------|-----------------------|---|--|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| Carbamazepine (Tegretol)* | Blood | None  | 4.9 | Blood Tube | Take before next dose | 1 working day Mon - Fri |
| Digoxin* | Blood | None  | 4.9 | Blood Tube | Usual sampling time is >6 hrs after dose. Assumed to be oral administration. | 1 working day Mon - Fri Urgent 3 hrs and verbal report |
| Flecainide* | Blood | None  | 4.9 | Blood Tube | Bring to lab immediately, must be frozen in <4 hrs | 15 working days |
| Gentamicin Once Daily Dosing | Blood | None  | 4.9 Adult | Blood Tube | Pre Level: Refer to Gentamicin Once Daily Guideline. | ♦ 120 mins φ |
| Gentamicin Multi Daily Dosing | Blood | None  | 4.9 Adult | Blood Tube | Pre and Post Levels: Refer to Endocarditis Infection Guideline. | ♦ 120 mins φ |
| Levetiracetam (Keppra)* | Blood | None  | 4.9 | Blood Tube | Take before next dose | 15 working days |
| Lithium* | Blood | None  | 4.9 | Blood Tube | Take 12 hrs post dose | 2 working days |
| Methotrexate* | Blood | None  | 4.9 | Blood Tube | Contact Biochemistry before test. Wrap tube in tin foil in transit. Whole blood, do not centrifuge. | 1 working day |
| Mitotane* | Blood | Potassium EDTA  | 2.7 | Blood Tube | None | 15 working days |

- * These specimens/ samples are referred to external laboratories for testing.
♦ Tests provided in the emergency out of hours service.
φ Note: 95% of results reported for this test will achieve the turnaround time as stated.

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BIOCHEMISTRY TESTS - Therapeutic Drug Monitoring.....cont'd

| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|--|---------------|---|---------------------------|-----------------------|---|---|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| Paracetamol* | Blood | None  | 4.9 | Blood Tube | Take at least 4 hrs post suspected overdose. | Routine therapeutic <2 days Urgent/ query OD Verbal in < 3 hours from CUH. |
| Perampanel (Fycompa)* | Blood | None  | 4.9 | Blood Tube | Take before next dose | 15 working days |
| Phenobarbital* (Phenobarbitone) | Blood | None  | 4.9 | Blood Tube | Take before next dose | 2 working days |
| Phenytoin* (Epanutin) | Blood | None  | 4.9 | Blood Tube | Take before next dose | 1 working day |
| Quinidine* | Blood | None  | 4.9 | Blood Tube | Before next dose. Bring to lab immediately, serum to be frozen within 4 hrs of collection | 15 working days |
| Salicylate* | Blood | None  | 4.9 | Blood Tube | None | Routine therapeutic <2 days Urgent/ query OD Verbal in < 3 hours from CUH. |
| Tacrolimus (Prograf)* | Blood | Potassium EDTA  | 4 | Blood Tube | Take as trough sample | 7 working days |
| Theophylline* | Blood | None  | 4.9 | Blood Tube | Sample time 2-4 hrs post oral administration of drug | 5 working days |

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BIOCHEMISTRY TESTS - Therapeutic Drug Monitoring.....cont'd

| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|--------------------------------|---------------|---|---------------------------|-----------------------|---|-----------------|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| Tobramycin | Blood | None  | 4.9 | Blood Tube | Refer to Tobramycin Once Daily Guideline. Please specify on the request form whether sample is peak (post) or trough (pre) and indicate the time since last administration of the drug | ◆ 120 mins ϕ |
| Toxicology Screen* | Blood | None  | 4.9 | Blood Tube | Please state medications | 15 working days |
| Toxicology Screen* | Spot urine | None | 20 | Sterile Universal | Please state medications | 15 working days |
| Valproic Acid (Epilim)* | Blood | None  | 4.9 | Blood Tube | Take before next dose | 1 working day |
| Vancomycin | Blood | None  | 4.9 Adult | Blood Tube | Refer to Vancomycin Dosing & Monitoring Guideline | ◆ 120 mins ϕ |

- * These specimens/ samples are referred to external laboratories for testing.
- ◆ Tests provided in the emergency out of hours service.
- ϕ Note: 95% of results reported for this test will achieve the turnaround time as stated.

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9.4 BIOCHEMISTRY TESTS – Allergy Testing

| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|---|---------------|---|---------------------------|-----------------------|--|-----------------|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| Penicillin Allergen Panel Penicillin V and G | Blood | None  | 4.9 | Blood Tube | None Please click to access Allergy Request Form PL006 | 10 working days |
| Single Allergen Test (Specify required Allergen(s) from list on back of form PL006) | Blood | None  | 4.9 | Blood Tube | None Refer to section 9.4.1 for details of available Allergens Please click to access Allergy Request Form PL006 | 10 working days |
| Referred Single Allergen Test (Specify Allergen on right hand side of form PL006)* | Blood | None  | 4.9 | Blood Tube | None Refer to section 9.4.1 for details of available Allergens Please click to access Allergy Request Form PL006 | 15 working days |
| IgE Immunoglobulin E (Please request on form PL006) | Blood | None  | 4.9 | Blood Tube | None Please click to access Allergy Request Form PL006 | 10 working days |

* These specimens/ samples are referred to external laboratories for testing.

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9.4.1 **BIOCHEMISTRY TESTS – Allergen Testing**

| |
|---|
| SAMPLE REQUIREMENTS: ALLERGY REQUEST |
| 4.9mL blood tube containing no additive |
| TURNAROUND TIME: |
| 10 working days for in-house tests |

We have changed our testing protocols to provide a focused allergen testing service instead of Allergy Screens in accordance with the recommendations of the all-Ireland Consultant Immunologist Group.






1. Allergen requests will only be processed if the appropriate Allergen request form (PL006) is used. Clinical details and symptoms as well as any previous allergies should be stated on the form. GPs should download this form and complete as we cannot accept GP referral letters for allergy testing.
2. Requests for **Screens** or for “**RAST**” will not be processed and this comment will be issued on the report.
3. A maximum of five allergens may be requested and should be based on clinical history.

The following table shows the available Allergens.

| Inhalant Allergens: | Foods Allergens: | Foods Allergens: |
|--------------------------------|------------------------------|-------------------------|
| House dust mite (d1) | Egg white (f1) | Soybean (f14) |
| Cat epithelium and dander (e1) | Milk (f2) | White bean (f15) |
| Horse dander (e3) | Fish (cod) (f3) | Pea (f12) |
| Dog dander (e5) | Wheat (f4) | Chick pea (f309) |
| Rabbit epithelium (e82) | Egg yolk (f75) | |
| | Shrimp (f24) | |
| Timothy grass (g6) | Kiwi (f84) | |
| Meadow Fescue (g4) | Peanut (f13) | |
| Rye Grass (g5) | Brazil Nut (f18) | |
| English plantain (w9) | Almond (f20) | |
| Common silver birch (t3) | Cashew Nut (f202) | |
| | Pistachio (f203) | |
| Penicillium mould (m1) | Walnut (f256) | |
| Cladosporium mould (m2) | Sesame (f10) | |
| Aspergillus mould (m3) | Hazel nut (f17) | |
| Alternaria mould (m6) | Pecan nut (f201) | |
| Occupational Allergens | Penicillin Allergens: | |
| Latex (k82) | Penicillin G (c1) and V (c2) | |
| Chlorhexidine (c8) | | |
| | | |
| Total IgE | Order IgE using PL006 | |

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9.5 BIOCHEMISTRY TESTS – General

| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|--|--------------------------|---|---------------------------|-----------------------|--|-----------------|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| 5-HIAA* 5-Hydroxyindoleacetic Acid | 24 Hour Urine Collection | 50mL 2M HCl | 100 | 5L Urine container | Refer to section 4.3.2. No pineapple, nuts, bananas or kiwi fruit are to be eaten immediately before or during collection | 15 working days |
| Abatacept* (Orencia) | Blood | None  | 4.9 | Blood Tube | Please right click link to access Request Form choose 'Copy link address' from dropdown menu and paste the link into the Chrome browser, press return. Review request form for Abatacept levels and send with the sample. Sample must be taken before the next dose – trough level. Antibody analysis is currently not available | 15 working days |
| ACE* Angiotensin Converting Enzyme | Blood | None  | 4.9 | Blood Tube | None | 10 working days |
| Acetylcholine Receptor Antibodies* | Blood | None  | 4.9 | Blood Tube | Freeze 1mL serum <1 hr of specimen collection | 15 working days |
| Acyl Carnitine Profile* | Blood | None  | Fill Circles | Guthrie Card | Click here to access Metabolic Investigations Request Form and send with the sample | 25 working days |
| Adalimumab* (Humira) | Blood | None  | 4.9 | Blood Tube | Please right click link to access Request Form , choose 'Copy link address' from dropdown menu and paste the link into the Chrome browser, press return. Review request form for Adalimumab antibodies and levels and send with the sample. Sample must be taken before the next dose – trough level. | 15 working days |

* These specimens/ samples are referred to external laboratories for testing.

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BIOCHEMISTRY TESTS – Generalcont'd

| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|--|--------------------------|---|---------------------------|-----------------------|--|-----------------|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| Albumin | Blood | None  | 4.9 | Blood Tube | None | ♦ 120 mins ϕ |
| Albumin Creatinine Ratio | Spot Urine | None | 2 | Sterile Universal | Please note that the drugs Flucytosine and Eltrombopage can cause interference with the Creatinine method. Please inform the Laboratory if the patient is on either of these medicines so that Creatinine can be measured by a different method. | 120 mins ϕ |
| Albumin (Fluid) | Fluid (pleural etc) | None | 2 | Sterile Universal | None | ♦ 120 mins ϕ |
| Albumin (Urinary) Microalbumin | 24 hour urine collection | None | 10 | 5L Urine container | Refer to section 4.3.2 | 120 mins ϕ |
| Alcohol* (Testing not for legal purposes, please contact the Lab if request is in relation to legal or road traffic act) | Blood | Fluoride  | 2.7 | Blood Tube | Do not use alcohol swab on arm. | 10 working days |
| Aldolase* | Blood | None  | 4.9 | Blood Tube | 1 mL serum post 30 mins rest | 15 working days |
| Alkaline Phosphatase isoenzymes* | Blood | None  | 4.9 | Blood Tube | None | 20 working days |







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♦ Tests provided in the emergency out of hours service.

ϕ Note: 95% of results reported for this test will achieve the turnaround time as stated.

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BIOCHEMISTRY TESTS – Generalcont'd

| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|--|---------------|--|---------------------------|-----------------------|---|-------------------------|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| Alkaline Phosphatase | Blood | None  | 4.9 | Blood Tube | None | ♦ 120 mins ϕ |
| Alkaline Phosphatase (Fluid) | Fluid | None | 2 | Sterile Universal | None | ♦ 120 mins ϕ |
| Alpha-Amino adipic Semialdehyde (AASA)* | Urine | None | 2 | Sterile Universal | Freeze sample within 30 mins | 25 working days |
| Alpha-1 Antitrypsin | Blood | None  | 4.9 | Blood Tube | None | 1 working day Mon - Fri |
| Alpha-1 Antitrypsin in faeces* | Faeces | None | 20 | Sterile Universal | None | 15 working days |
| Alpha-1 Antitrypsin Phenotype* | Blood | None  | 4.9 | Blood Tube | Note Specimen Reception 2 x 1mL aliquot required | 25 working days |
| ALT Alanine Amino Transferase | Blood | None  | 4.9 | Blood Tube | None | ♦ 120 mins ϕ |
| Amino Acids* | Blood | Lithium Heparin  | 4.9 Adult 2 x 1.2 Paed | Blood Tube | Click here to access Metabolic Investigations Request Form and send with the sample | 15 working days |
| Amino Acids* | Spot Urine | None  | 10 | Sterile Universal | Must be frozen. Send to lab immediately. Check pH in lab and if > 8.5 then a repeat sample must be obtained. Click here to access Metabolic Investigations Request Form and send with the sample | 15 working days |

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BIOCHEMISTRY TESTS – General.....cont'd

| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|----------------------------------|---------------------|--|--------------------------------|-----------------------------|--|-----------------|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| Ammonia | Blood | Potassium EDTA  | 2.7 Adult 1.3 Paed | Blood Tube | Transport immediately on ice. Separate and freeze if delayed | ♦ 120 mins ϕ |
| Amylase | Blood | None  | 4.9 | Blood Tube | None | ♦ 120 mins ϕ |
| Amylase (Fluid) | Fluid (pleural etc) | None | 2 | Sterile Universal | None | ♦ 120 mins ϕ |
| Amyloid A Protein* | Blood | None  | 4.9 | Blood Tube | Bring to lab immediately, must be frozen in <1 hr | 15 working days |
| Anti-GAD Antibodies* | Blood | None  | 4.9 | Blood Tube | Freeze serum within 1 hr of specimen collection | 20 working days |
| Anti-GAD Antibodies* | CSF | None | Refer to form PL012 for volume | Sterile Universal | Bring to lab immediately | 20 working days |
| Anti-tyrosine Antibodies* | Blood | None  | 4.9 | Blood Tube | Freeze serum within 4 hrs of specimen collection | 15 working days |
| Apolipoprotein A* | Blood | None  | 4.9 | Blood Tube | 12 hrs fasting prior to collection | 15 working days |
| Apolipoprotein B* | Blood | None  | 4.9 | Blood Tube | 12 hrs fasting prior to collection | 15 working days |
| Apolipoprotein E Apo E* | Blood | Serum  | 4.9 | Blood Tube | None | 15 working days |
| Arsenic* | Blood | Lithium Heparin  | 7.5 | Trace metal-free tube (Bio) | Urine arsenic is the recommended sample | 15 working days |









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


BIOCHEMISTRY TESTS – General.....cont'd

| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|---|-----------------|--|---------------------------|-----------------------|--|-----------------|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| Arsenic* | Spot Urine | None | 25 | Sterile Universal | None | 15 working days |
| AST Aspartate Amino Transferase | Blood | None  | 4.9 | Blood Tube | None | ♦ 120 mins φ |
| Beta-2 Transferrin* | Fluid and Blood | None  | 2 | Sterile Universal | None | 30 working days |
| | | None  | 5 | Blood Tube | | |
| Bicarbonate | Blood | Lithium Heparin  | 4.9 | Blood Tube | None | 1 working day |
| Bilirubin Direct | Blood | None  | 4.9 Adult 1.3 Paed | Blood Tube | Please note that the drug Eltrombopage can interfere with this method. Please inform the laboratory if the patient is on this medicine. | ♦ 120 mins φ |
| Bilirubin Total | Blood | None  | 4.9 Adult 1.3 Paed | Blood Tube | Please note that the drug Eltrombopage can interfere with this method. Please inform the laboratory if the patient is on this medicine. | ♦ 120 mins φ |
| Biotinidase* | Blood | Lithium Heparin  | 1.3 | Blood Tube | Separate and freeze plasma within 3 hrs | 25 working days |
| Bisoprolol* | Blood | None  | 4.9 | Blood Tube | None | 15 working days |
| Blood Gases - Venous - Arterial - Angiography | Blood | Heparinised syringe | 1 | Blood Gas Syringe | Do not transport by the Pneumatic Tube System. To be walked immediately to Lab. Sample must be assayed within 60 mins. If lactate required, sample must be received and analysed within 10 mins. | ♦ <60 mins |

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BIOCHEMISTRY TESTS – General.....cont'd

| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|--|--------------------------|--|---------------------------|-----------------------------|--|-----------------|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| BNP (B-Type Natriuretic Peptide) | Blood | Potassium EDTA  | 2.7 | Blood Tube | Separate EDTA sample is required for this test. | ♦ 120 mins ϕ |
| Breath Test for Helicobacter pylori | Breath | Follow Helicobacter Diagnostic Test Kit Protocol | | | Samples of breath are taken in OPD. Before taking specimen, patient:- <ul style="list-style-type: none"> • Should be off antibiotics for at least 4 weeks • Should be off antacids for at least 2 weeks • Should not eat for at least 6 hrs (may drink water) Refer to policy GASTRO008 and GASTRO009. | 15 working days |
| Cadmium* | Blood | Lithium Heparin  | 7.5 | Trace metal-free tube (Bio) | None | 15 working days |
| Cadmium* | Spot Urine | None | 100 | Sterile Universal | None | 15 working days |
| Caffeine* | Blood | None  | 1.3 | Blood Tube | None | 5 working days |
| Calcium | Blood | None  | 4.9 | Blood Tube | None | ♦ 120 mins ϕ |
| Calcium (Urinary) | 24 hour urine collection | 50 mL 2M HCl | 10 | 5L Urine Container | Refer to section 4.3.2 | 1 working day |
| Calcium* (Ionised) | Blood (Arterial/Venous) | Balanced Heparin Syringe | 1 | Blood Gas Syringe | Arrange test in advance with Biochemistry lab as assay must be arranged with Mercy Hospital. Transport immediately to lab. | 2 hours |

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BIOCHEMISTRY TESTS – General.....cont'd

| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|--|--------------------------|--|---------------------------|-----------------------|--|----------------------|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| Calcium Creatinine Ratio in Urine | Urine | None | 10 | Sterile Universal | Please note that the drugs Flucytosine and Eltrombopage can cause interference with the Creatinine method. Please inform the Laboratory if the patient is on either of these medicines so that Creatinine can be measured by a different method. | 120 mins ϕ |
| EGFR (Calculation for Chemotherapy Only) | Blood | None  | 4.9 | Blood Tube | Provide patients weight and height | ◆ 120 mins ϕ |
| Calculi Analyses* - Kidney Stone - Gall Stone | Calculi (Stone) | None | Not applicable | Sterile Universal | None | 20 working days |
| Carbohydrate Deficient Transferrin* | Blood | None  | 4.9 | Blood Tube | None | 15 working days |
| Carnitine (Total)* | Blood | Lithium Heparin  | 9 | Blood Tube | Centrifuge & freeze plasma immediately after specimen collection | 15 working days |
| Carnitine (Urine)* | Spot Urine | None | 10 | Sterile Universal | Please ensure blood sample is taken at the same time. | 15 working days |
| Catecholamines and Metanephrines* | 24 hour urine collection | 50mL, 2M HCl | 100 | 5L Urine Container | Refer to section 4.3.2 | 15 working days |
| Catecholamines * | Blood | Potassium EDTA  | 9 | Blood Tube | Diet: Within 48 hrs of assay do not eat chocolate, bananas or citrus fruit, and consume tea and coffee in moderation. Beta-blocker treatment may interfere with assay. Freeze plasma within 1 hr of specimen collection. | 15 working days |

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






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BIOCHEMISTRY TESTS – General.....cont'd










| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|---|---------------|--|---------------------------|-----------------------|--|-----------------|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| Ceruloplasmin* | Blood | None  | 4.9 | Blood Tube | None | 15 working days |
| Chloride | Blood | None  | 4.9 | Blood Tube | None | ♦ 120 mins ϕ |
| Cholesterol | Blood | None  | 4.9 | Blood Tube | None | ♦ 120 mins ϕ |
| Cholesterol – HDL | Blood | None  | 4.9 | Blood Tube | None | ♦ 120 mins ϕ |
| Cholesterol – LDL | Blood | None  | 4.9 | Blood Tube | None | ♦ 120 mins ϕ |
| Cholinesterase* | Blood | None  | 4.9 x 2 | Blood Tube | None | 15 working days |
| Chromium* | Blood | Lithium Heparin  | 7.5 | Trace Metal-Free Tube | None | 15 working days |
| CK Isoenzymes* | Blood | None  | 4.9 | Blood Tube | None | 20 working days |
| CKMB* | Blood | None  | 4.9 | Blood Tube | None | 15 working days |
| CK-total | Blood | None  | 4.9 | Blood Tube | None | ♦ 120 mins ϕ |
| Cobalt* | Blood | Lithium Heparin  | 7.5 | Trace Metal Free Tube | None | 15 working days |
| Complement C1 Inhibitor Quantitation and Function* | Blood x 2 | None Sodium Citrate | 4.9 5 | Blood Tube x 2 | Quantitation will only be performed if Lab receives only serum sample. | 20 working days |

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BIOCHEMISTRY TESTS – General.....cont'd

| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|----------------------------------|--------------------------|--|---------------------------|-----------------------|--|----------------------------|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| Complement Function Test* | Blood | None  | 4.9 | Blood Tube | None | 15 working days |
| Complement C₂* | Blood | None  | 4.9 | Blood Tube | None | 15 working days |
| Complement C₃ | Blood | None  | 4.9 | Blood Tube | None | 1 working day Mon - Fri |
| Complement C₄ | Blood | None  | 4.9 | Blood Tube | None | 1 working day Mon - Fri |
| Complement C₅* | Blood | None  | 4.9 | Blood Tube | None | 15 working days |
| Complement Total (CH50)* | Blood | None  | 4.9 | Blood Tube | Freeze serum within 1hr of specimen collection | 20 working days |
| Copper* | Blood | Lithium Heparin  | 7.5 | Trace Metal Free Tube | None | 15 working days |
| Copper* | 24 hour urine collection | None | 200 | 5L Urine container | Refer to section 4.3.2 | 15 working days |
| C-peptide* | Blood | None  | 4.9 | Blood Tube | Freeze serum within 4hrs of specimen collection | 7 working days |
| Creatinine | Blood | None  | 4.9 | Blood Tube | Please note that the drugs Flucytosine and Eltrombopage can cause interference with the Creatinine method. Please inform the Laboratory if the patient is on either of these medicines so that Creatinine can be measured by a different method. | ♦ 120 mins φ |

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BIOCHEMISTRY TESTS – General.....cont'd

| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|---|--------------------------|---|---------------------------|-----------------------|--|-------------------|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| Creatinine Clearance | 24 Hour urine collection | None | 10 | 5L Urine container | Refer to section 4.3.2 4.9 mL clotted blood to be taken during 24 hrs collection | 120 mins ϕ |
| | Blood | None  | 4.9 | Blood Tube | Please note that the drugs Flucytosine and Eltrombopage can cause interference with the Creatinine method. Please inform the Laboratory if the patient is on either of these medicines so that Creatinine can be measured by a different method. | 120 mins ϕ |
| Creatinine Clearance Calculated GFR for Chemotherapy | Blood | None  | 4.9 | Blood Tube | Please provide patient's weight and height. Please note that the drugs Flucytosine and Eltrombopage can cause interference with the Creatinine method. Please inform the Laboratory if the patient is on either of these medicines so that Creatinine can be measured by a different method. | ◆ 120 mins ϕ |
| Creatinine Fluid | Fluid (pleural etc) | None | 2 | Sterile Universal | Please note that the drugs Flucytosine and Eltrombopage can cause interference with the Creatinine method. Please inform the Laboratory if the patient is on either of these medicines so that Creatinine can be measured by a different method. | ◆ 120 mins ϕ |

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BIOCHEMISTRY TESTS – General.....cont'd

| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|---|---------------|---|---------------------------|---|--|-----------------|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| C – Reactive Protein (CRP) | Blood | None  | 4.9 | Blood Tube | None | ♦ 120 mins φ |
| Cryoglobulins | Blood | None  | 4.9 | Pre-warmed Blood Tube | Keep at 37°C Bring to Biochemistry immediately | 7 working days |
| CSF Amino Acids* | CSF | None | 1 | Sterile Universal | Please right click link to access Request Form. choose 'Copy link address' from dropdown menu and paste the link into the Chrome browser, press return. Send request form with the sample. | 28 working days |
| CSF Neurotransmitters * | CSF | Special Tubes from Bio | 2 | Special Tubes from Bio | Must contact Biochemistry one week in advance of collecting sample. | 30 working days |
| CSF Phospho-Tau Protein* | CSF | None | 0.25 | Sterile Universal | Sample to be frozen on day of collection. Request this test using CSF form PL012. | 30 working days |
| CSF Spectrophotometry for Xanthochromia* | CSF | Special tube from Bio. Protect from light. | 1 | Special tube from Bio and completed special request form, including CT scan result. | Please contact Bio lab before taking sample. Sample to be taken minimum 12 hours and up to 7 days maximum post event. Special request form to be completed. | 5 working days |
| CSF Tau A-Beta Protein* | CSF | None | 0.25 | Sterile Universal | Sample to be frozen at - 80°C on day of collection. Request this test using CSF form PL012. | 30 working days |

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BIOCHEMISTRY TESTS – General.....cont'd

| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|--|---------------|---|---------------------------|-----------------------|--|--|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| CTX (C-Telopeptide of Type 1 procollagen)* | Serum | None  | 4.9 | Blood Tube | Fasting sample required. Non haemolysed serum to be Transported Frozen: Note: Possible interference in patients treated with Biotin (Vitamin B7, B8 or H) or taking food supplements containing Biotin. Patient should not take these materials for 8 days prior to sample collection. | 10 working days |
| Cystine* | Spot Urine | None | 10 | Sterile Universal | 10 mL early morning urine (fasting). Frozen <1 hr. | 15 working days |
| Elastase* | Faeces | None | 2g | Sterile Universal | None | 15 working days |
| Electrophoresis of Serum (Serum Protein Electrophoresis SPE) | Blood | None  | 4.9 | Blood Tube | Do not request an electrophoresis of serum if it is less than one month since the previous order for this test. In exceptional circumstances, please contact the Biochemistry Lab if a sample needs to be processed in a stated timeframe. | 5 working days Note: If immunofixation is required based on the Electrophoresis result, the turnaround time will be increased by 6 working days |

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BIOCHEMISTRY TESTS – General.....cont'd

| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|--|---------------------|--|---------------------------|-----------------------|---|--|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| Electrophoresis of Urine (Urine Immunoelectrophoresis) | Early Morning Urine | None | 20 | Sterile Universal | Refrigerate if delay in bringing to Lab. Do not request an electrophoresis of urine if it is less than one month since the previous order for this test. In exceptional circumstances, please contact the Biochemistry Lab if a sample needs to be processed in a stated timeframe. | 5 working days Note: If additional immunofixation is required based on the original result, the turnaround will be increased by 6 working days. |
| Ethosuximide* | Blood | None  | 4.9 | Blood Tube | Trough sample, give details of dosage including length of time patient is on this medication | 10 working days |
| Faecal Calprotectin* | Faeces | None | 20g minimum | Sterile Universal | Store at 4°C | 15 working days |
| Fibroblast Growth Factor 23* | Blood | Potassium EDTA  | 7.5 | Blood Tube | None | 25 working days |
| Free Fatty Acids* | Blood | None  | 4.9 | Blood Tube | Bring to lab immediately, must be frozen in <1 hr. Patient must be fasting | 15 working days |
| Very Long Chain Fatty Acids (VLCFA)* | Blood | Lithium Heparin  | 4.9 | Blood Tube | Bring to lab immediately | 25 working days |
| Fructosamine* | Blood | None  | 4.9 | Blood Tube | None | 20 working days |
| Gamma GT | Blood | None  | 4.9 | Blood Tube | None | ♦ 120 mins φ |









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BIOCHEMISTRY TESTS – General.....cont'd

| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|--|---------------------|---|--------------------------------------|-----------------------|---|-----------------|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| Globulin | Blood | None  | 4.9 | Blood Tube | None | ♦ 120 mins ϕ |
| Globulin (CSF) | CSF | None | Refer to form PL012 for volume | Sterile Universal | None | ♦ 120 mins ϕ |
| Glucose (Fasting) | Blood | Fluoride  | 2.7 Adult 1.3 Paed | Blood Tube | 12 hrs fasting prior to collection | ♦ 120 mins ϕ |
| Glucose (Non-Fasting) | Blood | Fluoride  | 2.7 Adult 1.3 Paed | Blood Tube | None | ♦ 120 mins ϕ |
| Glucose 2 hours post (Prandial) | Blood | Fluoride  | 2.7 Adult 1.3 Paed | Blood Tube | 2 hrs post meal | ♦ 120 mins ϕ |
| Glucose (CSF) | CSF | None | Refer to form PL012 for volume | Sterile Universal | Ensure concurrent blood glucose measurement | ♦ 120 mins ϕ |
| Glucose (Fluid) | Fluid (pleural etc) | None | 2 | Sterile Universal | None | ♦ 120 mins ϕ |
| Glucose Tolerance Test | Blood | Fluoride  | 2.7 Adult 1.3 Paed | Blood Tube x 3 | Contact Pathology dept. to arrange | 5 hours |
| Glycosamino-glycans (Mucopolysaccharides)* | Spot Urine | Merthiolate (Added in Biochemistry) | 30 | Sterile Universal | 2 nd specimen of day | 50 working days |
| Guthrie Card (New Born Screen)* | Blood | None  | Fill circles on Guthrie Card and dry | N/A | Take > 7 days after birth | 25 working days |
| Haptoglobin* | Blood | None  | 4.9 | Blood Tube | None | 15 working days |
| HbA1c Glycosylated Haemoglobin | Blood | Potassium EDTA  | 2.7 | Blood Tube | Please take separate sample for HbA1c analysis. Whole blood, do not centrifuge. | 3 working days |
| Histamine (Urine)* | Urine | None | 20 | Sterile Universal | None | 15 working days |



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BIOCHEMISTRY TESTS – General.....cont'd













| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|-----------------------------|--------------------------|---|---------------------------|-----------------------|---|-----------------------|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| Histamine (Plasma)* | Blood | Potassium EDTA  | 2.7 x 2 | Blood Tube x 2 | None | 15 working days |
| Homocysteine* | Blood | Sodium Citrate  | 4.9 Adult | Blood Tube | Centrifuge & freeze immediately after specimen collection | 15 working days |
| | | | 1.5 Paed | Blood Tube | Paediatric Samples: Centrifuge & freeze immediately after specimen collection. Click here to access Metabolic Investigations Request Form and send with the sample | 15 Working days |
| Homogentistic Acid* | Early morning urine | None | 20 | Sterile Universal | Bring to lab immediately, must be frozen in <1 hr | 15 working days |
| Hydroxyproline* | 24 Hour urine collection | None | 50 | 5L Urine container | Refer to section 4.3.2 Store urine in a refrigerator (+4°C) between each urination. At the end of collection, send the urine to the lab without delay DIET: within 48 hours prior to the assay, avoid consuming collagen-rich foods (meat, jelly or gelatin, ice creams, confectioneries, cold meats, etc.) | 15 working days |
| Hypocretin (Orexin)* | CSF | None | 1.0 | Sterile Universal | Request this test using CSF form PL012. | Up to 60 working days |

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| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|---|---------------|---|---------------------------|-----------------------|---|----------------------------|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| IgA Immuno-globulin A | Blood | None  | 4.9 Adult 1.3 Paed | Blood Tube | None | 1 working day Mon - Fri |
| IgA Subclasses* | Blood | None  | 4.9 | Blood Tube | None | 15 working days |
| IgD Im-muno-globulin D* | Blood | None  | 4.9 | Blood Tube | None | 15 working days |
| IgE Immuno-globulin E | Blood | None  | 4.9 | Blood Tube | None | 10 working days |
| IgG Immuno-globulin G | Blood | None  | 4.9 Adult 1.3 Paed | Blood Tube | None | 1 working day Mon - Fri |
| IgG Immuno-globulin G Subclasses* | Blood | None  | 4.9 | Blood Tube | None | 15 working days |
| IgM Immuno-globulin M | Blood | None  | 4.9 Adult 1.3 Paed | Blood Tube | None | 1 working day Mon – Fri |
| Immune Complexes* | Blood | None  | 4.9 | Blood Tube | None | 20 working days |
| Immunoreactive Trypsin* | Blood | None  | 4.9 | Blood Tube | Bring to lab immediately, must be frozen in <1 hr | 15 working days |
| Infliximab Antibodies* | Blood | None  | 4.9 | Blood Tube | Please right click link to access Request Form choose 'Copy link address' from dropdown menu and paste the link into the Chrome browser, press return. Review request form for infliximab antibodies and levels and send with the sample. Sample must be taken before the next dose – trough level. | 15 working days |
| Insulin Like Growth Factor 1 (IGF1)* | Blood | None  | 4.9 | Blood Tube | Freeze serum < 4 hrs of specimen collection | 15 working days |
| Insulin Like Growth Factor 2 (IGF2)* | Blood | None  | 4.9 | Blood Tube | Freeze serum < 4 hrs of specimen collection | 28 working days |

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BIOCHEMISTRY TESTS – General.....cont'd

| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|--|------------------------|---|---------------------------|-----------------------|--|-----------------|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| Insulin Like Growth Factor Binding Protein 3 (IGF BP3)* | Blood | None  | 4.9 | Blood Tube | Freeze serum < 4 hrs of specimen collection | 20 working days |
| Interleukin-6* | Blood | None  | 4.9 | Blood Tube | Bring to lab immediately, must be frozen in <4 hrs | 15 working days |
| Iron (Fe) | Blood | None  | 4.9 | Blood Tube | None | ♦ 120 mins ϕ |
| Serum Kappa / Lambda (including Kappa Lambda Ratio) | Blood | None  | 4.9 | Blood Tube | Do not request a Kappa/ Lambda ratio if it is less than one month since the previous order for this test. In exceptional circumstances, please contact the Biochemistry Lab if a sample needs to be processed in a stated timeframe. | 10 working days |
| 24 hr Urine Kappa & Lambda* | 24 hr urine collection | None | 20 | 5L Urine Container | Do not request a urine Kappa/ Lambda if it is less than one month since the previous order for this test. In exceptional circumstances, please contact the Biochemistry Lab if a sample needs to be processed in a stated timeframe. | 15 working days |

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BIOCHEMISTRY TESTS – General.....cont'd




| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|------------------------------------|---------------|---|--------------------------------|-----------------------|---|-----------------|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| Lactate | Blood | Fluoride EDTA (Adult Glucose Tube) | 2.7 | Blood Tube | Transport to Biochemistry immediately on ice. | ♦ 120 mins φ |
| Lactate (Paediatric) | Blood | Fluoride EDTA (Adult Glucose Tube) | 1.3 | Blood Tube | Transport to Biochemistry immediately on ice. | ♦ 120 mins φ |
| Lactate | CSF | Fluoride EDTA (Adult Glucose Tube) | Refer to form PL012 for volume | Blood Tube | Transport to Biochemistry immediately on ice. | ♦ 120 mins φ |
| Lactate Dehydrogenase (LDH) | Blood | None  | 4.9 | Blood Tube | Do not transport by the Pneumatic Tube System | ♦ 120 mins φ |

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| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|---|--------------------------|---|-------------------------------------|-----------------------|--|-----------------|
| | | <u>Additive Required</u> | <u>Volume Required</u> <u>mL</u> | <u>Container Type</u> | | |
| Lactate (Blood Gas) | Blood | Heparinsed Syringe | 1.0 | Blood Gas Syringe | Do not transport by the Pneumatic Tube System. Please walk sample immediately to Lab and ideally transport on ice. Sample must be assayed within 10 mins. Please state if venous or arterial as reference interval is different. | ♦ 60 mins ϕ |
| Lactate Dehydrogenase (LDH Isoenzymes)* | Blood | None  | 4.9 | Blood Tube | Do not transport by the Pneumatic Tube System | 15 working days |
| Lactate Dehydrogenase (LDH Fluid) | Fluid (pleural etc) | None | 2 | Sterile Universal | Do not transport by the Pneumatic Tube System | ♦ 120 mins ϕ |
| Lamotrigine* | Blood | None | 4.9 | Blood Tube | Bring to lab immediately, must be frozen in <4 hrs | 15 working days |
| Laxative Screen (Faeces)* | Faeces | None | 20 | Sterile Universal | None | 25 working days |
| Laxative Screen (Urine)* | Urine | None | 20 | Sterile Universal | None | 25 working days |
| Lead* | Blood | Potassium EDTA  | 2.7 x 2 | Blood Tube x 2 | Whole blood, do not centrifuge | 15 working days |
| Lead* | 24 hour urine collection | None | 30 | 5L Urine container | Refer to section 4.3.2 | 15 working days |
| Lipase* | Blood | None  | 4.9 | Blood Tube | None | 15 working days |

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


BIOCHEMISTRY TESTS – General.....cont'd

| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|-------------------------------------|--|--|---------------------------|---|--|----------------------------|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| Urinary Lipase* | 24 hour urine collection preferably, otherwise single specimen | None | 10 | 5L Urine container preferably otherwise Sterile Universal | Refer to section 4.3.2 | 15 working days |
| Lipoprotein (a)* | Blood | None  | 4.9 | Blood Tube | None | 15 working days |
| Magnesium | Blood | None  | 4.9 | Blood Tube | None | ♦ 120 mins ϕ |
| Magnesium (Urine) | 24 Hour Urine collection | 50mL 2M HCl | 10 | 5L Urine container | Refer to section 4.3.2 | 1 working day Mon – Fri |
| Manganese* | Blood | Lithium Heparin  | 7.5 | Trace Metal-Free Tube | None | 15 working days |
| Mercury* | Blood | Lithium Heparin  | 7.5 | Trace metal free tube (Bio) | None | 20 working days |
| Mercury* | Spot Urine | None | 100 | Sterile Universal | None | 15 working days |
| Metanephrines (Plasma)* | Blood | Lithium Heparin  | 4.9 x 2 | Blood Tube x 2 | Bring to lab immediately, must be frozen in <30 mins | 25 working days |
| Methaemoglobin | Blood | Lithium Heparin  | 4.9 | Blood Tube | Venous sample taken into blood gas syringe | <60 mins |
| Methylhistamine* | Urine | 50mL 2M HCl | 100 | 5L Urine Container | None | 15 working days |
| Methyl Malonic Acid – Plasma | Blood | Lithium Heparin  | 4.9 | Blood Tube | Bring to lab immediately, must be frozen in <60 mins | 20 working days |

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| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|---|---------------|--|-------------------------------------|--|--|----------------------------|
| | | <u>Additive Required</u> | <u>Volume Required</u> <u>mL</u> | <u>Container Type</u> | | |
| Microalbumin Creatinine Ratio | Spot Urine | None | 10 | Sterile Universal | Please note that the drugs Flucytosine and Eltrombopage can cause interference with the Creatinine method. Please inform the Laboratory if the patient is on either of these medicines so that Creatinine can be measured by a different method. | 1 working day |
| Beta 2 Microglobulin | Blood | None  | 4.9 | Blood Tube | None | 1 working day Mon – Fri |
| Molybdenum* | Blood | Lithium Heparin  | 7.5 | Trace Metal-Free Tube | None | 15 working days |
| Myoglobin* | Spot Urine | None | 20 | Sterile Universal | Sample needs to be frozen within 1 hour. | 15 working days |
| Myoglobin Serum* | Blood | None  | 4.9 | Blood Tube | None | 15 working days |
| NTx (N-telopeptide)* | Spot Urine | None | 5 | Sterile Universal | Freeze on day of collection, send to referral lab frozen. | 15 working days |
| Occupational Health Drugs of Abuse Screen* | Urine | None | 50 | x 2 marked "A" and "B" Sterile Universal | Chain of custody kits (bottles A+B) must be ordered in advance via Biochemistry. | 15 working days |
| Oligoclonal banding* | CSF | None | Refer to form PL012 for volume | Sterile Universal | Clotted blood taken simultaneously | 20 working days |
| Oligoclonal banding (Small Volume)* | CSF | None | Refer to form PL012 for volume | Sterile Universal | Clotted blood taken simultaneously | 15 working days |

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| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|-----------------------|--------------------------|---|---------------------------|-----------------------|---|---|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| Organic Acids* | Spot Urine | None | 10 | Sterile Universal | Must be frozen. Send to lab immediately. Check pH in lab and if > 8.5 then a repeat sample must be obtained. Click here to access Metabolic Investigations Request Form and send with the sample | 20 working days |
| Osmolality | Blood | None  | 4.9 | Blood Tube | None | ♦ 1 working day 2 Hrs when taken as part of water deprivation test |
| Osmolality | Spot Urine | None | 5 | Sterile Universal | Send to lab immediately | ♦ 1 working day 2 Hrs when taken as part of water deprivation test |
| Osteocalcin* | Blood | None  | 4.9 | Blood tube | Non Haemolysed Frozen serum required | 10 working days |
| Oxalate* | 24 hour urine collection | 50 mL 20% HCl | 20 | 5L Urine container | Refer to section 4.3.2 | 15 working days |

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BIOCHEMISTRY TESTS – General.....cont'd

| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|-----------------------------------|--------------------------|--|---------------------------|-----------------------|--|----------------------------|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| Oxalate/ Creatinine Ratio* | Spot Urine | None | 5 | Sterile Universal | Please note that the drugs Flucytosine and Eltrombopage can cause interference with the Creatinine method. Please inform the Laboratory if the patient is on either of these medicines so that Creatinine can be measured by a different method. | 15 working days |
| Oxalate* | Blood | None  | 4.9 x 2 | Blood tube x 2 | Separated and frozen within 3 hours | 15 working days |
| pH in Pleural Fluid | Fluid | Lithium Heparin  | 1 | Blood Gas Syringe | None | <60 mins ♦ |
| Phosphate (Inorganic) | Blood | None  | 4.9 | Blood Tube | None | ♦ 120 mins φ |
| Phosphate (Urinary) | 24 hour urine collection | 50 mL 2M HCl | 10 | 5L Urine Container | Refer to section 4.3.2 | 1 working day Mon - Fri |
| Phytanic Acid* | Blood | Potassium EDTA  | 2.7 x 2 | Blood Tube x 2 | Bring to lab immediately, must be frozen in <1 hr | 15 working days |
| Porphyrins (Blood)* † | Blood | Potassium EDTA  | 7.5 | Blood Tube | Whole Blood. Protect from light (cover with tin foil and store at 4°C). Click here to access Porphyrin Request Form and send with the sample | 15 working days |

* These specimens/ samples are referred to external laboratories for testing.







♦ Tests provided in the emergency out of hours service.

† These tests require special patient preparation or conditions. For clarification purposes, please contact Biochemistry department before taking sample.

φ Note: 95% of results reported for this test will achieve the turnaround time as stated.

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BIOCHEMISTRY TESTS – General.....cont'd

| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|--|---------------------|---|--------------------------------------|-----------------------|---|-----------------|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| Porphyryns (Faeces)* † | Faeces | None | 10g | Sterile Universal | Protect from light, refrigerate. Click here to access Porphyrin Request Form and send with the sample | 15 working days |
| Porphyryns (plasma)* † | Blood | Potassium EDTA  | 7.5 | Blood Tube | Protect from light. Click here to access Porphyrin Request Form and send with the sample | 15 working days |
| Porphyryns (urine)* † | Urine | None | Random urine. Minimum volume of 5mL. | Sterile Universal | Protect from light (cover with tin foil and store at 4°C). Click here to access Porphyrin Request Form and send with the sample | 15 working days |
| Potassium (K) | Blood | None  | 4.9 | Blood Tube | None | ♦ 120 mins φ |
| | Urine | | | Sterile Universal | | |
| Prealbumin* | Blood | None  | 4.9 | Blood Tube | None | 15 working days |
| Procollagen Type-1 Pro Peptide* | Blood | None  | 4.9 | Blood Tube | None | 15 working days |
| Procollagen 3* | Blood | None  | 4.9 | Blood Tube | Freeze serum <4 hrs after collection | 20 working days |
| Protein Total | Blood | None  | 4.9 | Blood Tube | None | ♦ 120 mins φ |
| Protein Total (CSF) | CSF | None | Refer to form PL012 for volume | Sterile Universal | None | ♦ 120 mins φ |
| Protein Total (Fluid) | Fluid (pleural etc) | None | 2 | Sterile Universal | None | ♦ 120 mins φ |

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BIOCHEMISTRY TESTS – General.....cont'd








| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|---------------------------------|--------------------------|---|--|--------------------------------|--|-----------------|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| Protein (Urinary) | 24 hour urine collection | None | 10 | 5L urine container | Refer to section 4.3.2 | 1 working day |
| Protein Creatinine Ratio | Urine | None | 10 | Sterile Universal | Please note that the drugs Flucytosine and Eltrombopage can cause interference with the Creatinine method. Please inform the Laboratory if the patient is on either of these medicines so that Creatinine can be measured by a different method. | 1 working day |
| PTH-Related Protein* | Blood | Potassium EDTA and Aprotinin  | 7.5 mL tube with Aprotinin available from Specimen Reception | Blood Tube | Send on ice. Freeze plasma within 1 hr of specimen collection | 25 working days |
| Pyruvate*† | Blood | Perchloric Acid | 2 | Special tube from Biochemistry | Contact Biochemistry for special tube before performing test. | 25 working days |
| Red Cell Folate* | Blood | Potassium EDTA  | 2.7 x 1 | Blood Tube | Send both whole blood and serum for analysis | 15 working days |
| | | None  | 4.9 x 1 | Blood Tube | | |
| Salivary Cortisol* | Saliva | None | 1 | Salivette | Contact Biochemistry for special containers. | 20 working days |
| Selenium* | Blood | Lithium Heparin  | 7.5 | Trace Metal Free Tube | None | 15 working days |

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BIOCHEMISTRY TESTS – General.....cont'd

| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|---|---------------|--|---------------------------|-----------------------|---|-----------------|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| Sodium (Na) | Blood | None  | 4.9 | Blood Tube | None | ♦ 120 mins ϕ |
| | Urine | | | Sterile Universal | | |
| Sperm Antibodies* | Blood | None  | 4.9 | Blood Tube | None | 15 working days |
| Sulphonylureas* | Urine | None | 2 | Sterile Universal | Freeze within 4 hrs | 15 working days |
| Thiopurine-S-Methyl Transferase (Red Cell) (TPMT)* | Blood | Potassium EDTA  | 2.7 | Blood Tube | Whole blood keep at 4°C | 20 working days |
| Transferrin | Blood | None  | 4.9 | Blood Tube | None | ♦ 120 mins ϕ |
| Transferrin Glycoform Analysis* | Blood | Lithium heparin  | 1.3 | Blood Tube | None | 25 working days |
| Transferrin Saturation (Calculation) | Blood | None  | 4.9 | Blood Tube | None | 120 mins ϕ |
| Triglycerides | Blood | None  | 4.9 | Blood Tube | If fasting triglyceride is required collect sample after 12 hrs fasting | ♦ 120 mins ϕ |
| Triglycerides (Fluid) | Fluid | None | 2 | Sterile Universal | None | ♦ 120 mins ϕ |






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BIOCHEMISTRY TESTS – General.....cont'd

| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|--|--------------------------|--|---------------------------|-----------------------|---|----------------------------|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| Troponin-I | Blood | Lithium heparin  | 4.9 | Blood Tube | Samples to be taken in accordance with Troponin Algorithm. Click here to access Troponin Algorithm. | ♦ 120 mins ϕ |
| Tryptase* | Blood | None  | 4.9 | Blood Tube | When investigating anaphylactic shock take sample immediately after event and again at 2 and 4 hours. | 20 working days |
| Tumour Necrosis Factor – Alpha (TNF Alpha)* | Blood | None  | 4.9 | Blood Tube | Bring to lab immediately. Must be frozen in < 4 hr | 28 working days |
| Urate (uric acid) | Blood | None  | 4.9 | Blood Tube | None | ♦ 120 mins ϕ |
| Urate (Urinary) | 24 hour urine collection | 50 mL 2.5 M NaOH | 10 | 5L Urine Container | Refer to section 4.3.2 | 1 working day Mon - Fri |
| Urate (Fluid) | Fluid | None | 2 | Sterile Universal | None | ♦ 120 mins ϕ |
| Urea | Blood | None  | 4.9 | Blood Tube | None | ♦ 120 mins ϕ |
| | Urine | | | Sterile Universal | | |
| Urinary Cotinine* | Urine | None | 10 | Sterile Universal | None | 15 working days |
| Urinary Electrolytes (Na, K, Cl, Urea) | 24 hour urine collection | None | 10 | 5L Urine container | Refer to section 4.3.2 | 1 working day ♦ |
| Urinary Electrolytes (Spot) (Na, K, Cl, Urea) | Urine | None | 5 | Sterile Universal | None | 1 working day ♦ |
| Urinary Na, K, Osmolality | Urine | None | 5 | Sterile Universal | Transport to lab immediately | ♦ 120 mins ϕ |



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BIOCHEMISTRY TESTS – General.....cont'd

| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|---|---------------|--|---------------------------|-----------------------|--|----------------------------|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| Urine Purine and Pyrimidine Screen* | Urine | None | 5 | Sterile Universal | Store sample at 4°C | 20 working days |
| Vedolizumab (Entyvio)* | Blood | None  | 4.9 | Blood Tube | Click here to access Request Form and levels and send with the sample. Sample must be taken before the next dose – trough level. | 15 working days |
| Vigabatrin* | Blood | None  | 4.9 | Blood Tube | Bring to lab immediately, must be frozen in <4 hrs | 15 working days |
| Viscosity* | Blood | Potassium EDTA  | 7.5 | Blood Tube | Keep at room temp | 10 working days |
| Vitamin A* (Retinol) | Blood | Lithium Heparin  | 4.9 | Blood Tube | Protect from light. Freeze plasma in <1 hour of specimen collection | 15 working days |
| Vitamin B₁* (Thiamine) | Blood | Potassium EDTA  | 7.5 | Blood Tube | Protect from light. Freeze blood within 4 hours of specimen collection | 20 working days |
| Vitamin B₆* (Pyridoxal Phosphate) | Blood | Potassium EDTA  | 2.7 | Blood Tube | Protect from light. Freeze blood within 4 hours of specimen collection | 20 working days |
| Vitamin B₁₂ | Blood | None  | 4.9 | Blood Tube | None | 1 working day Mon - Fri |
| Vitamin C* (Ascorbic Acid) | Blood | Lithium Heparin  | 9 | Blood Tube | Freeze plasma <1 hr of specimen collection. Store away from light. | 20 working days |

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BIOCHEMISTRY TESTS – General.....cont'd

| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|--|---------------|--|---------------------------|-----------------------|---|----------------------------|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| Vitamin D (25 OH Vitamin D) A measure of overall reserves, dietary intake and endogenous sunlight dependent synthesis. | Blood | None  | 4.9 | Blood Tube | None | 1 working day Mon - Fri |
| Vitamin D₃* (1, 25 (OH) ₂ Vitamin D) A measure of Vitamin D metabolism. | Blood | None  | 4.9 x 2 | Blood Tube | Sample to be sent to Lab immediately. Serum to be frozen within 4 hrs of specimen collection. | 20 working days |
| Vitamin E* (Tocopherol) | Blood | Lithium Heparin  | 4.9 | Blood Tube | Protect sample from light, bring to lab immediately, plasma to be frozen within 90 mins of collection | 15 working days |
| Vitamin K* (Phylloquinone) | Blood | None  | 4.9 | Blood Tube | Protect sample from light, bring to lab immediately, serum to be frozen within 90 mins of collection | 15 working days |
| Xylose | Blood | Fluoride  | 2.7 Adult | Blood Tube | Contact Biochemistry for details of test | 15 working days |
| Xylose | Urine | None | 10 | Sterile Universal | Contact Biochemistry for details of test | 15 working days |
| Zinc* | Blood | Lithium Heparin  | 7.5 | Trace Metal Free Tube | None | 15 working days |

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9.6 BIOCHEMISTRY PROFILES

On every patient where a serum creatinine has been ordered an estimated GFR is reported, using the CKD-EPI equation. It must be noted that this estimation of GFR is **not** suitable for the calculation of chemotherapy dosages where the existing eGFR will continue to be used. Where a patient has an eGFR requested the CKDEPI value will not be reported, in order to avoid any potential confusion.

Full information on the use of this equation and the interpretation of results can be found at http://www.kidney.org/professionals/cls/pdf/12-10-4004_KBB_FAQs_AboutGFR-1.pdf.

| ADMISSION PROFILE INCLUDING FBC |
|---|
| Sodium, Potassium, Chloride, Urea, Creatinine, Urate, Calcium, Phosphate, Total Bilirubin, Direct Bilirubin, Alkaline Phosphatase (ALP), Aspartate Aminotransferase (AST), Alanine Aminotransferase (ALT), Gamma-Glutamyl Transferase (GGT), Total Protein, Albumin, Globulin, Glucose, Magnesium (Mg), FBC |
| Sample Requirements |
| Blood Tube (4.9mL) containing no additive Blood Tube (2.7mL) containing EDTA Glucose Tube Fluoride (2.7 adult, 1.3 paed) |
| Turn Around Time |
| ♦ 120 mins φ Note: 95% of results reported for this test will achieve the turnaround time as stated. |

| CHILDREN'S WARD BIOCHEMISTRY |
|---|
| Sodium, Potassium, Chloride, Urea, Creatinine, Urate, Calcium, Magnesium (Mg), Total Protein, Albumin, Globulin, Alkaline Phosphatase (ALP), Alanine Aminotransferase (ALT), Aspartate Aminotransferase (AST), C – Reactive Protein (CRP), Random Glucose |
| Sample Requirements |
| Blood Tube (1.3mL) containing no additive |
| Turn Around Time |
| ♦ 120 mins φ Note: 95% of results reported for this test will achieve the turnaround time as stated. |

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BIOCHEMISTRY PROFILES.....cont'd

| LIPOPROTEIN PROFILE (FASTING) |
|---|
| Cholesterol, Triglycerides, High Density Lipoprotein (HDL), Low Density Lipoprotein (LDL) |
| Sample Requirements |
| Blood Tube (4.9mL) containing no additive |
| Special Requirements |
| Patient should be fasting for at least 12 hours before Phlebotomy (Fluids are allowed) |
| Turn Around Time |
| ♦ 120 mins φ Note: 95% of results reported for this test will achieve the turnaround time as stated. |

| LIPID PROFILE (NON-FASTING) |
|---|
| Cholesterol, Triglycerides, High Density Lipoprotein (HDL), Low Density Lipoprotein |
| Sample Requirements |
| Blood Tube (4.9mL) containing no additive |
| Turn Around Time |
| ♦ 120 mins φ Note: 95% of results reported for this test will achieve the turnaround time as stated. |

| LIVER/ RENAL / BONE PROFILE |
|---|
| Sodium, Potassium, Chloride, Urea, Creatinine, Urate, Calcium, Phosphate, Total Bilirubin, Direct Bilirubin, Alkaline Phosphatase (ALP), Alanine Aminotransferase (ALT), Aspartate Aminotransferase (AST), Gamma-Glutamyl Transferase (GGT), Total Protein, Albumin, Globulin, Magnesium (Mg) |
| Sample Requirements |
| Blood Tube (4.9mL) containing no additive |
| Turn Around Time |
| ♦ 120 mins φ Note: 95% of results reported for this test will achieve the turnaround time as stated. |

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BIOCHEMISTRY PROFILES.....cont'd

| LIVER FUNCTION TESTS |
|---|
| Total Bilirubin, Direct Bilirubin, Alkaline Phosphatase (ALP), Alanine Aminotransferase (ALT), Aspartate Aminotransferase (AST), Gamma-Glutamyl Transferase (GGT), Total Protein, Albumin, Globulin |
| Sample Requirements |
| Blood Tube (4.9mL) containing no additive |
| Turn Around Time |
| ♦ 120 mins φ Note: 95% of results reported for this test will achieve the turnaround time as stated. |

| PATIENT PROFILE (NON FASTING) |
|--|
| Sodium, Potassium, Chloride, Urea, Creatinine, Urate, Calcium, Phosphate, Cholesterol, Total Bilirubin, Direct Bilirubin, Alkaline Phosphatase (ALP), Alanine Aminotransferase (ALT), Aspartate Aminotransferase (AST), Gamma-Glutamyl Transferase (GGT), Total Protein, Albumin, Globulin |
| Sample Requirements |
| Blood Tube (4.9mL) containing no additive |
| Turn Around Time |
| ♦ 120 mins φ Note: 95% of results reported for this test will achieve the turnaround time as stated. |

| PATIENT PROFILE (+LPP - FASTING) |
|--|
| Sodium, Potassium, Chloride, Urea, Creatinine, Urate, Calcium, Phosphate, Cholesterol, Triglycerides, High Density Lipoprotein (HDL), Low Density Lipoprotein (LDL), Total Bilirubin, Direct Bilirubin, Alkaline Phosphatase (ALP), Alanine Aminotransferase (ALT), Aspartate Aminotransferase (AST), Gamma-Glutamyl Transferase (GGT), Total Protein, Albumin, Globulin |
| Sample Requirements |
| Blood Tube (4.9mL) containing no additive |
| Special Requirements |
| Patient should be fasting for at least 12 hours before Phlebotomy (Fluids are allowed) |
| Turn Around Time |
| ♦ 120 mins φ Note: 95% of results reported for this test will achieve the turnaround time as stated. |

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BIOCHEMISTRY PROFILES.....cont'd

| RENAL / BONE PROFILE |
|--|
| Sodium, Potassium, Chloride, Urea, Creatinine, Urate, Calcium, Albumin, Phosphate, Alkaline Phosphatase (ALP), Total Protein, Globulin, Magnesium (Mg) |
| Sample Requirements |
| Blood Tube (4.9mL) containing no additive |
| Turn Around Time |
| ♦ 120 mins ϕ Note: 95% of results reported for this test will achieve the turnaround time as stated. |

| THYROID FUNCTION TESTS |
|---|
| Free Thyroxine (FT4), Thyroid Stimulating Hormone (TSH) |
| Sample Requirements |
| Blood Tube (4.9mL) containing no additive |
| Turn Around Time |
| 1 Working Day (Monday to Friday) |

| UREA / ELECTROLYTES / CREATININE |
|--|
| Sodium, Potassium, Chloride, Urea, Creatinine |
| Sample Requirements |
| Blood Tube (4.9mL) containing no additive |
| Turn Around Time |
| ♦ 120 mins ϕ Note: 95% of results reported for this test will achieve the turnaround time as stated. |

| UREA / ELECTROLYTES / CREATININE & LIVER |
|--|
| Sodium, Potassium, Chloride, Urea, Creatinine, Total Bilirubin, Direct Bilirubin, Alkaline Phosphatase (ALP), Alanine Aminotransferase (ALT), Aspartate Aminotransferase (AST), Gamma-Glutamyl Transferase (GGT), Total Protein, Albumin, Globulin |
| Sample Requirements |
| Blood Tube (4.9mL) containing no additive |
| Turn Around Time |
| ♦ 120 mins ϕ Note: 95% of results reported for this test will achieve the turnaround time as stated. |

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BIOCHEMISTRY PROFILES.....cont'd

| COUNTERWEIGHT PLUS FASTING PROFILE |
|---|
| Sodium, Potassium, Chloride, Urea, Creatinine, Urate, Calcium, Phosphate, Total Bilirubin, Direct Bilirubin, Alkaline Phosphatase (ALP), Aspartate Aminotransferase (AST), Alanine Aminotransferase (ALT), Gamma-Glutamyl Transferase (GGT), Total Protein, Albumin, Globulin, Cholesterol, Triglyceride, High Density Lipoprotein (HDL), Low Density Lipoprotein (LDL), *HbA1c, Full Blood Count (FBC), Ferritin*, Free T4*, Thyroid Stimulating Hormone (TSH), Glucose Tolerance Test |
| Sample Requirements |
| Blood Tube (4.9mL) containing no additive Blood Tube (2.7mL) containing EDTA x 2 Glucose Tube Fluoride (2.7mL) x 2 if GTT requested |
| Turn Around Time |
| 120 mins φ Note: 95% of results reported for this test will achieve the turnaround time as stated. 1 working day for tests marked with * |

- ◆ Tests provided in the emergency out of hours service.
- φ Note: 95% of results reported for this test will achieve the turnaround time as stated.

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9.7 **FLUID BATTERIES**

9.7.1 Tests are ordered on non-standard body fluids (those other than blood, urine and CSF) depending on the site of the fluid as follows:-

| Fluid Type | Tests |
|-------------------|--|
| Pericardial | Total protein LDH Glucose |
| Peritoneal | Total protein Albumin Glucose |
| Pleural | Total protein Albumin LDH Glucose |
| Synovial | Total protein Albumin LDH Glucose |

A clotted blood sample must be sent at the same time.

9.7.2 The appearance of the fluid sample will also be noted. In addition, if required the following tests are also available on fluid samples:-

- Alkaline phosphatase
- Amylase
- Cholesterol
- Creatinine
- Glucose
- Lactate
- Triglycerides
- Urate

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9.8 Troponin Algorithm

In May 2014 a new high sensitivity Troponin I (hsTnI) method was introduced into routine use. The algorithm in Table 1 shows a suggested use of this assay in the evaluation of acute chest pain.

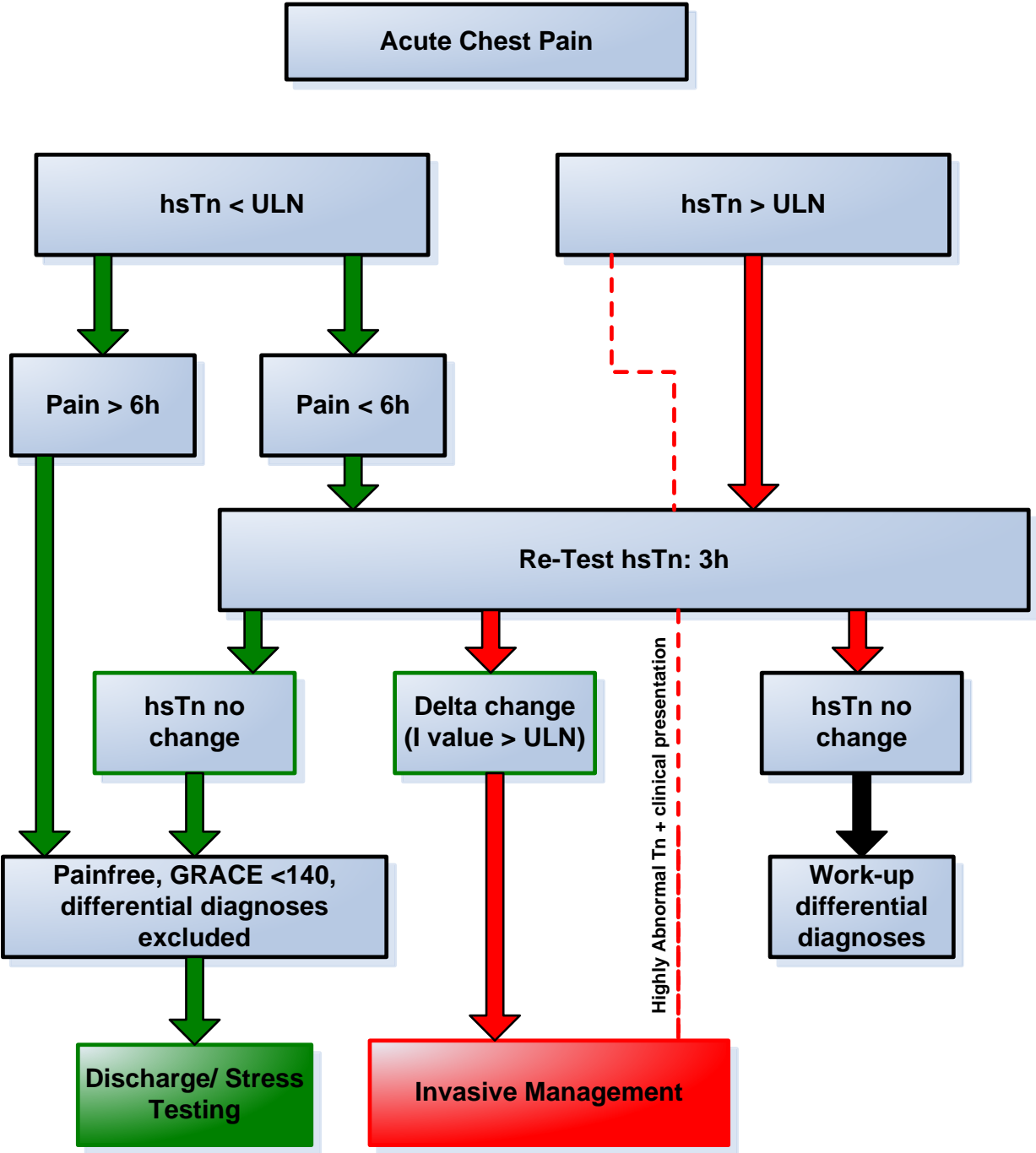
Please note that the new method differs from the previous assay in a number of ways:-

1. The units of hsTnI are ng/L (also equivalent to pg/mL) as opposed to µg/L. The numerical results change by a factor of 1000 so 0.04 µg/L will now be expressed as 40 ng/L.
2. There is a gender difference in the new method.
Male: < 34 ng/L and Female: < 16 ng/L when reported to the nearest whole number.
The reference interval for the existing method is < 0.05 µg/L (=> 50 ng/L) and does not differentiate male and female,
3. The new Algorithm can be used in the evaluation of Acute Chest Pain. This requires a measurement at presentation and again three hours later. The interpretation of the result is based not only on the absolute concentration but also on the change (delta) between the baseline and the second value. A delta change of 50% over the Baseline value and/or a value greater than the gender specific Upper Limit of Normal (ULN) is significant and should be investigated.
4. An abnormally high value of > 262 ng/L at presentation also supports the diagnosis of MI in an appropriate clinical context.

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Table 1



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10.0 BLOOD TRANSFUSION

10.1 BLOOD TRANSFUSION TESTS

ABO incompatible transfusion can be a fatal but avoidable event. Erroneous patient identification and specimen labelling account for many errors that lead to ABO mismatched transfusion. If the blood in the tube used for pretransfusion compatibility testing is not that of the patient identified on the label i.e., ‘Wrong blood in tube’ (WBIT), this may lead to catastrophic outcomes, such as death from ABO-incompatible red cell transfusion. WBIT is a National Haemovigilance Office reportable incident.



Two-Sample Rule: The two-sample rule is a well-established guideline to improve patient safety by identifying cases of ‘Wrong blood in tube’ (WBIT). The Blood Bank must ensure that there are TWO distinct samples from a patient that have generated the same blood group from both samples.

If the Blood Bank already has a historic blood group registered on the Laboratory Information System, then only one Group and Screen/Crossmatch sample is required. If the patient has no previous records in Blood Bank, BSHC and is for crossmatch, then the Group and Screen/Crossmatch must be repeated with a second sample. Only one transfusion sample, whether for a Group and Screen or Group and Crossmatch, should be taken at initial phlebotomy.

It is **NOT** acceptable to take two samples at one venepuncture event and send them to Blood Bank on separate request forms as this does not improve patient safety.

If a Crossmatch request is received, or if a Group and Screen is converted into a Crossmatch, the Transfusion Laboratory will inform the clinical area if a second transfusion sample is required to confirm the patient’s blood group. Where time does not permit a second sample to be taken, Group O red cells will be issued.

BLOOD TRANSFUSION TESTS



| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|---------------------------------|---------------|---|---------------------------|-----------------------|----------------------|-----------------|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| Cold Agglutinins | Blood | Potassium EDTA  | 7.5 | Blood Tube | None | 2 working days☀ |
| Direct Antiglobulin Test | Blood | Potassium EDTA  | 2.7 | Blood Tube | None | 4 hours♦ |

♦ Tests provided in the emergency out of hours service.

☀ These tests may not be performed until the following routine day if received after 16.00 hours on Monday to Friday

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



BLOOD TRANSFUSION TESTS.....cont'd

| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|-----------------------------|---------------|---|---------------------------|-----------------------|--|---|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| Group and Crossmatch | Blood | Potassium EDTA  | 7.5 2.7 Paed | Blood Tube | Urgent requests, contact Blood Bank Laboratory See also section 10.3 titled "Life Threatening Haemorrhage". All samples are stored for 28 days only. After 28 days, a new sample must be taken for Group and Crossmatch. Note: If the patient has been transfused or pregnant in the last 3 mths, a new sample is required after 3 days. CHECK BLOOD TRACK FOR AVAILABILITY OF BLOOD | Routine: 4 hours (except pre-assessment, see note below) ^ Note: testing is prioritised according to patient need ♦# |
| Group and Screen | Blood | Potassium EDTA  | 7.5 2.7 Paed | Blood Tube | Urgent requests, contact Blood Bank Laboratory See also section 10.3 titled "Life Threatening Haemorrhage". All samples are stored for 28 days only. After 28 days, a new sample must be taken for Group and Screen or Crossmatch. Note: If the patient has been transfused or pregnant in the last 3 mths, a new sample is required after 3 days. | Routine: 1 day. Note: testing is prioritised according to patient need ♦# |

- # Turnaround times may be significantly increased if:
- Antibodies are present
 - Samples are referred to external site
 - Patient is on treatment that affects serological transfusion tests e.g. Daratumumab
- ^ Crossmatch samples for pre-assessment clinic patients are taken up to **28 days** in advance of surgery. The crossmatch will be done at a time that ensures the blood is available for the date and time stated on the request form.
- ♦ Tests provided in the emergency out of hours service.

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BLOOD TRANSFUSION TESTS.....cont'd


| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|--|---------------|---|---------------------------|-----------------------|---|--|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| Human Leucocyte Antigen (HLA) Typing* | Blood | Potassium EDTA  | 5 - 10 | Blood Tube | Samples should be received Mon to Thurs am. | 10 working days |
| Human Leucocyte Antigen Antibodies* | Blood | None  | 4.9 | Blood Tube | Samples should be received Mon to Thurs am. | 10 working days |
| Transfusion Reaction Investigation | Blood | Potassium EDTA  | 7.5 | Blood Tube | Refer to Hospital Transfusion Handbook (BSC/HV/MAN/001) | 10 working days Serological results will be available on the next routine day ∇ |
| Platelet Antibodies* | Blood | None  | 4.9 x 2 | Blood Tube | Samples should be received Mon to Thurs am. | 10 working days |
| Platelet Genotyping by DNA* | Blood | Potassium EDTA  | 10–20 Adult 1-2 Paed | Blood Tube | Samples should be received Mon to Thurs am. | 10 working days |

∇ Tests provided in the emergency out of hours service if authorised by a Consultant.

* These specimens/ samples are referred to external laboratories for testing.

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10.2 BLOOD PRODUCTS/COMPONENTS FOR TRANSFUSION

| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|--|---------------|---|---------------------------|-----------------------|--|--|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| Albumin | None | None | None | None | Refer to Hospital Transfusion Handbook (BSC/HV/MAN/001) | Immediately ♦ |
| Anti D Immunoglobulin | None | None | None | None | Refer to Hospital Transfusion Handbook (BSC/HV/MAN/001) | Routine: 1 hour Urgent: Immediately ♦ |
| Coagulation Factor Concentrates | None | None | None | None | Refer to Hospital Transfusion Handbook (BSC/HV/MAN/001) | Not normally held in the Blood Bank. Advance notice during routine hours is required |
| Fibrinogen Concentrate | None | None | None | None | Refer to Hospital Transfusion Handbook (BSC/HV/MAN/001) See also section 10.3 titled “Life Threatening Haemorrhage” | Routine: 1 hour Urgent: Immediately ♦ |
| Prothrombin Concentrate Complex (PCC) | None | None | None | None | Refer to Hospital Transfusion Handbook (BSC/HV/MAN/001) | Routine: 1 hour Urgent: Immediately ♦ |
| Platelets● | Blood | Potassium EDTA  | 7.5 2.7 Paed | Blood Tube | Urgent: Contact the Blood Bank. Platelets are not held in the Blood Bank routinely. They must be brought in from the IBTS. Refer to Hospital Transfusion Handbook (BSC/HV/MAN/001). See also section 10.3 titled “Life Threatening Haemorrhage” | 4 hours ♦ |

- ♦ Tests provided in the emergency out of hours service.
- The blood specimen is required if the blood group has not been previously tested by the laboratory.

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BLOOD PRODUCTS/COMPONENTS FOR TRANSFUSION.....cont'd

| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|--|---------------|---|---------------------------|-----------------------|--|--|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| Solvent Detergent Plasma (SDP) (Frozen Plasma)● | Blood | Potassium EDTA  | 7.5 2.7 Paed | Blood Tube | Refer to Hospital Transfusion Handbook (BSC/HV/MAN/001) See also section 10.3 titled “Life Threatening Haemorrhage” | 1 hour if blood group already know by laboratory otherwise 2 hours ♦ |
| Red Cells● | Blood | Potassium EDTA  | 7.5 2.7 Paed | Blood Tube | Refer to Hospital Transfusion Handbook (BSC/HV/MAN/001) See also section 10.3 titled “Life Threatening Haemorrhage” | 4 hours (routine) ♦ See 10.1 Group and Screen and Group and Crossmatch |
| For details on other products, refer to Hospital Transfusion Book. | | | | | | |

- ♦ Tests provided in the emergency out of hours service.
- The blood specimen is required if the blood group has not been previously tested by the laboratory.

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10.3 **LIFE-THREATENING HAEMORRHAGE**

Contact the Blood Bank Laboratory immediately Ext 1721 or front desk for Medical Scientist on call. State Massive Haemorrhage Protocol Activated in Location XXXX for patient YYYY.

Refer to the Hospital Transfusion Handbook BSC/HV/MAN/001 Life-Threatening Haemorrhage Protocol.

Where a clinical condition dictates that a transfusion is required prior to the completion of testing, the transfusion support may vary depending on the following:-

- the degree of clinical urgency
- the prior availability of the patient's sample and a validated blood group report in the hospital blood bank
- the presence of antibodies
- complex transfusion requirements where the sample must be referred to the Irish Blood Transfusion Service for serological investigations and crossmatch

NOTE:

2 Emergency O Neg red cell units and 2 x 1g Emergency Fibrinogen Concentrate are normally available in the Blood Bank Issue Fridge.

1 Emergency O Neg red cell unit and 2 x 1g Emergency Fibrinogen Concentrate are normally available in the Satellite Blood Fridge.

Uncrossmatched Group O or group specific blood can be issued in 15 minutes. Risks associated with use are identical to risks associated with use of O RhD negative uncrossmatched blood, but uses resources better i.e. unnecessary use of O RhD negative blood is avoided.

Crossmatched blood will be available in 40 – 60 minutes, provided a sample has been received or is already in the Blood Bank, and the patient does not have antibodies.

RhD Negative red cells are a scarce resource and small stocks (minimum 5 units) are held in the hospital. Female patients of childbearing potential are prioritised for transfusion of RhD Negative red cells. The decision to switch other patients to RhD positive red cells is made in consultation with the patient's treating doctor and Consultant Haematologist, where clinical need for transfusion must be prioritised over transport time of further RhD Negative blood from IBTS, or after transfusion of 10 units of RhD Negative red cells (IBTS policy).





Platelets are not normally held in the Blood Bank, BSHC and must be transported from MRTC, IBTS, St. Finbarr's Hospital, Cork. This takes 30 – 45 minutes from time of receipt of order, provided the patient's blood group is known. Group A platelets will be ordered if the patient's blood group is unknown.

Frozen plasma (SDP) is held in the Blood Bank, BSHC and takes 40 minutes to thaw and issue to a patient. Thawed Group AB SDP will be issued if the patient's blood group is unknown.

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LIFE-THREATENING HAEMORRHAGE..... cont'd

| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|------------------------------|---------------|---|---------------------------|-----------------------|--|---|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| Group and Screen● | Blood | Potassium EDTA  | 7.5 2.7 Paed | Blood Tube | Contact Blood Bank Laboratory to check if blood sample is required | 45 minutes ♦# From time of receipt of sample |
| Group and Crossmatch● | Blood | Potassium EDTA  | 7.5 2.7 Paed | Blood Tube | Contact Blood Bank Laboratory to check if blood sample is required and estimated time of availability of red cells | 40 – 60 minutes from time of receipt of sample. Emergency O Neg available immediately Uncrossmatched blood – 15 minutes ♦# See above for further details |
| Red Cells● | Blood | Potassium EDTA  | 7.5 2.7 Paed | Blood Tube | Contact Blood Bank Laboratory to check if sample is required and estimated time of availability of red cells | 40 – 60 minutes if there is a valid sample already in the blood bank. Emergency O Neg available immediately Uncrossmatched blood – 15 minutes See Group and Crossmatch# |
| Platelets● | Blood | Potassium EDTA  | 7.5 2.7 Paed | Blood Tube | Contact Blood Bank Laboratory to check if sample is required and estimated time of availability of platelets. Platelets are not held in the Blood Bank routinely. See above for further details | 30- 45 minutes ♦ from time of receipt of order See above for further details |

- # Turnaround times may be significantly increased if:
- Antibodies are present
 - Samples are referred to external site
 - Patient is on treatment that affects serological transfusion tests e.g. Daratumumab
- ♦ Tests provided in the emergency out of hours service.
- The blood specimen is required if the blood group has not been previously tested by the laboratory.

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LIFE-THREATENING HAEMORRHAGE..... cont'd

| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|--|---------------|---|---------------------------|-----------------------|---|--|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| Solvent Detergent Plasma (SDP) (Frozen Plasma)● | Blood | Potassium EDTA  | 7.5 2.7 Paed | Blood Tube | Contact Blood Bank Laboratory to check if sample is required. SDP needs to be thawed before it can be issued from the Blood Bank | 40 minutes ♦ from time of receipt of order |
| Fibrinogen Concentrate | None | None | None | None | Contact Blood Bank Laboratory | Unlabelled – immediately Labelled for patient – 10 minutes ♦ from time of receipt of order |
| HAEMATOLOGY TESTS DURING LIFE-THREATENING HAEMORRHAGE | | | | | | |
| Full Blood Count (FBC) ☐ | Blood | Potassium EDTA  | 2.7 1.3 Paed | Blood Tube | Contact Haematology Laboratory | 10 Min ♦ from time of receipt of order ϕ |
| INR and APTT | Blood | Sodium Citrate  | 3 | Blood Tube | Contact Haematology Laboratory | 15-20 Min ♦ from time of receipt of order ϕ |
| Fibrinogen | Blood | Sodium Citrate  | 3 | Blood Tube | Contact Haematology Laboratory | 1 st request 35-40 Min further requests thereafter 15-20 Min ♦ from time of receipt of order ϕ |
| D-Dimer | Blood | Sodium Citrate  | 3 | Blood Tube | Contact Haematology Laboratory | 15-20 Min ♦ from time of receipt of order ϕ |

- ♦ Tests provided in the emergency out of hours service.
- The blood specimen is required if the blood group has not been previously tested by the laboratory on 2 separate occasions.
- ☐ In the event of the performance of manual blood films turnaround time is 2 hrs for Full Blood Counts.
- ϕ Note: 95% of results reported for this test will achieve the turnaround time as stated.

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11.0 HAEMATOLOGY TESTS

| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|--|-----------------|---|---------------------------|-----------------------|--|--|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| 11.1 BLOOD COUNTS AND ESR | | | | | | |
| Erythrocyte Sedimentation Rate (ESR) | Blood | Potassium EDTA  | 2.7 Adult 1.3 Paed | Blood Tube | None | 2 hours Performed on-call at request of the Consultant Haematologist only |
| Full Blood Count (FBC) | Blood | Potassium EDTA  | 2.7 Adult 1.3 Paed | Blood Tube | None | ◆ 90 mins ϕ |
| Full Blood Count including Manual Blood Film Examination (FBC) | Blood | Potassium EDTA  | 2.7 Adult 1.3 Paed | Blood Tube | None | ◆ 120 mins ϕ |
| Reticulocyte Count | Blood | Potassium EDTA  | 2.7 Adult 1.3 Paed | Blood Tube | None | 90 mins ☼ ϕ |
| 11.2 COAGULATION | | | | | | |
| Activated Partial Thromboplastin Time (APTT) | Blood | Sodium Citrate  | 3 | Blood Tube | Testing must be complete within 4 hours of specimen collection | ◆ 100 mins ϕ |
| D-Dimer | Blood | Sodium Citrate  | 3 | Blood Tube | Testing must be complete within 4 hours of specimen collection | ◆ 100 mins ϕ |
| DOAC Level* | Citrated Plasma | Sodium Citrate  | 7.5 | Blood Tube | None | 10 Working Days |
| Fibrinogen | Blood | Sodium Citrate  | 3 | Blood Tube | Testing must be complete within 4 hours of specimen collection | ◆ 100 mins ϕ |
| International Normalised Ratio (INR) | Blood | Sodium Citrate  | 3 | Blood Tube | Testing must be complete within 6 hours of specimen collection | ◆ 100 mins ϕ |

* These specimens/ samples are referred to external laboratories for testing.






◆ Tests provided in the emergency out of hours service.

☼ These tests may not be performed until the following routine day if received after 17.30 hours on Monday to Friday and after 12.30 on Saturdays.

ϕ Note: 95% of results reported for this test will achieve the turnaround time as stated.

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HAEMATOLOGY TESTS.....cont'd

| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|--|----------------------|--|---------------------------|-----------------------|--|--|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| 11.2 COAGULATION | | | | | | |
| Heparin Induced Thrombocytopenia Screen (HITs) | Blood | 2 Clotted Samples  | 10 | Blood Tube | Contact Laboratory. Samples must be received in lab immediately post Phlebotomy. The type and amount of Heparin administered must be stated on request form. Click here to access St. James's Test Request Form. All positive screens will be referred to SJH for confirmatory testing. | 2 hours Samples referred to SJH 20 working days |
| Prothrombin Time (PT) | Blood | Sodium Citrate  | 3 | Blood Tube | Testing must be complete within 6 hours of specimen collection | ◆ 100 mins ϕ |
| 11.3 SPECIAL STAINING PROCEDURES | | | | | | |
| Bone Marrow Aspirate | Bone Marrow Aspirate | None | N/A | Slides | Do not fix aspirate on ward. Aspirate will be fixed by staff in the laboratory. | 4 working days if accompanied with biopsy. 3 working days if stand-alone |
| 11.4 MISCELLANEOUS HAEMATOLOGY TESTS | | | | | | |
| EMA Screen for Hereditary Spherocytosis* | Blood | Potassium EDTA  | 7.5 + 2.7 | Blood Tube | Monday to Wednesday mornings only | 15 working days |
| Infectious Mononucleosis Screening Test (Monospot) | Blood | None  | 4.9 | Blood Tube | None | 4 hours ◆ |
| | | Potassium EDTA  | 2.7 | Blood Tube | None | |

- * These specimens/ samples are referred to external laboratories for testing.
- ♦ Tests provided in the emergency out of hours service.

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HAEMATOLOGY TESTS.....cont'd

| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|---|---------------|---|---------------------------|-----------------------|---|-----------------|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| 11.4 MISCELLANEOUS HAEMATOLOGY TESTS | | | | | | |
| Malaria Parasites Screen (This includes Blood Film Examination and Screening Test for Plasmodium Antigens) | Blood | Potassium EDTA  | 2.7 | Blood Tube | Where possible thick and thin smears should be prepared by Lab personnel. During out of hours the laboratory staff must be directly contacted. | 6 hours ♦ |
| Urinary hCG (Pregnancy Test) | Urine | None | 5-20 | Sterile Universal | None | 1 hour ♦ |
| Sickle Cell Screen (HBS)* | Blood | Potassium EDTA  | 2.7 | Blood Tube | Order FBC and Ferritin when requesting HBS Screen | 24 hours |
| 11.5 COAGULATION TESTS REFERRED | | | | | | |
| Coagulation Factor Assays* | Blood | Sodium Citrate  | 3 | Blood Tube | Samples should be received Mon to Thurs am | 10 working days |
| Factor Xa activity (Heparin Assay)* | Blood | Sodium Citrate  | 3 | Blood Tube | Samples should be received Mon to Thurs am. Specimens must be taken between 2 and 4 hrs post last administration of Heparin. Indicate on request form if patient is on Clexane or Innohep. | 10 working days |
| Inhibitor Screen* | Blood | Sodium Citrate  | 3 | Blood Tube | Samples should be received Mon to Thurs am | 10 working days |
| Lupus Anticoagulant* | Blood | Sodium Citrate  | 1 x 10 1 x 3.0 | Blood Tube | Clinical information must be provided. If a patient is on anticoagulation therapy, it must be discontinued for 1 month prior to sample collection. Samples must be received into the Lab within 1 hr of collection. | 10 working days |

* These specimens/ samples are referred to external laboratories for testing.

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HAEMATOLOGY TESTS.....cont'd

| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|---|---------------|---|---|-----------------------|--|-----------------|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| 11.5 COAGULATION TESTS REFERRED | | | | | | |
| Platelet Function Assessment (PFA 100)* | Blood | Sodium Citrate  | 3 x 3 | Blood Tube | Samples should be received Mon to Thurs am and by appointment with the laboratory | 10 working days |
| Thrombophilia Screen* | Blood | Sodium Citrate  And Potassium EDTA  1 Clotted Sample  | 1 x 10 1 x 3.0 2 x 2.7 1 x 4.9 | Blood Tube | Clinical information must be provided. Click here to access Eurofins Biomnis Consent Form and send with the sample. If a patient is on anticoagulation therapy, it must be discontinued for 1 month prior to sample collection. Samples must be received into the Lab within 1 hr of collection. | 15 working days |
| Von Willebrands Screen* | Blood | Sodium Citrate  | 3 x 3 | Blood Tube | Samples should be received Mon to Thurs am. | 15 working days |

* These specimens/ samples are referred to external laboratories for testing.

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HAEMATOLOGY TESTS.....cont'd

| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|---|----------------------|--|---------------------------|-----------------------|---|-----------------|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| 11.6 SPECIALISED HAEMATOLOGY TESTS (REFERRED) | | | | | | |
| ADAMTS 13 Factor Willibrand Cleavage Protease* | Peripheral Blood | Sodium Citrate  | 3 x 3 | Serum | Sample to be taken before any treatment, transfusion of plasma or any plasma exchange. Click here to access Biomnis Test Request Form and send with the sample. | 20 working days |
| CD34 (Marker)* | Blood | Potassium EDTA  | 2.7 | Blood Tube | Samples should be received Mon to Thurs am. | 10 working days |
| Glucose 6 Phosphate Dehydrogenase (G6PD)* | Blood | Potassium EDTA  | 2.7 1.3 (Paed) | Blood Tube | Samples should be received Mon to Thurs am. | 10 working days |
| Haemoglobin Screening (Abnormal)* | Blood | Potassium EDTA x 2  | 1.3 | Blood Tube | Samples should be received Mon to Thurs am. Order Ferritin and FBC assays also. | 10 working days |
| Leucocyte Immunopheno-Typing*♣ (Flow Cytometry) | Bone Marrow or Blood | Potassium EDTA  | 2.7 | Blood Tube | Samples should be received Mon to Thurs am. | 10 working days |
| Leucocyte Immunopheno-Typing* (Flow Cytometry) | CSF | Not applicable | 1 | Sterile Universal | CSF can only be taken Mon, Tues and up to 12 noon Wed | 10 working days |
| PNH (Flow Cytometry)* | Blood | Potassium EDTA  | 5-10 | Blood Tube | Samples can only be taken Mon, Tues, Wed and up to 12 noon Thurs | 5 working days |
| Pyruvate Kinase* | Blood | ACD Whole Blood supplied by Biomnis  | 5 | Blood Tube | Samples should be received Mon to Thurs am. | 10 working days |



* These specimens/ samples are referred to external laboratories for testing.

♣ Please contact the Haematology Laboratory, if requesting Leucocyte Immunopheno-Typing in the investigation of leukaemia.

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HAEMATOLOGY TESTS.....cont'd

| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|--|---------------|---|---------------------------|-----------------------|---|-----------------|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| 11.6 SPECIALISED HAEMATOLOGY TESTS (REFERRED) | | | | | | |
| T & B Lymphocytes* | Blood | Potassium EDTA  | 2.7 | Blood Tube | Samples should be received Mon to Thurs am. | 10 working days |
| T Cell Lymphocyte (T4, T8) Subset Levels* (CD4, CD8) | Blood | Potassium EDTA  | 2.7 | Blood Tube | Samples should be received Mon to Thurs am. | 10 working days |

* These specimens/ samples are referred to external laboratories for testing.

11.7 HAEMATOLOGY PROFILES

| COAGULATION SCREEN |
|---|
| Prothrombin Time (PT) International Normalised Ratio (INR) Activated Partial Thromboplastin Time (APTT) |
| Sample Requirements |
| Blood Tube (3mL) containing Sodium Citrate |
| Turn Around Time |
| ♦ 100 mins |

| LUPUS ANTICOAGULANT |
|--|
| International Normalised Ratio (INR) Activated Partial Thromboplastin Time (APTT) Fibrinogen APTT with Lupus Sensitivity (AFSL) Lupus Anticoagulant Screen |
| Sample Requirements |
| Patients should not be on any anticoagulant therapy or have anticoagulant therapy discontinued for 1 month. |
| Turn Around Time |
| 10 working days |

♦ Tests provided in the emergency out of hours service.

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11.7 HAEMATOLOGY PROFILEScont'd

| THROMBOPHILIA SCREEN |
|--|
| International Normalised Ratio (INR) Activated Partial Thromboplastin Time (APTT) Fibrinogen APTT with Lupus Sensitivity (AFSL) Lupus Anticoagulant Screen Activated Protein C Ratio (APCR) Anti-Thrombin 3 (AT3) Protein C (Pr C) Protein S (Pr S) Prothrombin Gene Mutation (PGM) |
| Sample Requirements |
| Patients should not be on any anticoagulant therapy or have anticoagulant therapy discontinued for 1 month. |
| Turn Around Time |
| 10 working days |

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12.0 HISTOPATHOLOGY

- 12.1**
- The “part type” description of a specimen as documented on the Histopathology request form by the Clinician is required for appropriate examination procedures relating to it.
 - This description also forms part of the final diagnostic report and consequently historical medical record of a patient. Therefore, the accuracy of this description is an **essential** requirement.
 - The Specimen Reception department in liaison with the Histopathology department may require clarification from the source of the specimen in cases where the description is absent, ambiguous or incomplete.
 - Note that the specimen description should **not** include the procedure, previous diagnosis or clinical history. There is a separate area on the request form for this information.
 - The time required to receive clarification may result in a delay of the final report. This can be avoided by the initial accurate completion of the specimen description on the request form.

The Histopathology National Quality Improvement (NQI) Programme divides Histopathology and Cytology specimens into categories according to the procedure (p) code within which turnaround times (TATs) are analysed.

It is presently increasingly difficult for Histopathology Departments nationally, including the BSHC Histopathology Department, to meet the NQI Target TAT for some routine cases. A process is in place to improve staffing and resources in the laboratory as we work towards achieving NQI Target TAT for all sample types. Our aim is to meet the NQI target TATs for all urgent cases. A realistic BSHC Target TAT for routine cases is included below.

| Test/Profile/P Code | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|--|---------------|----------------------------|--|---|----------------------|------------------------|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container /Type</u> | | |
| Small Biopsy (P01) | Fixed Tissue | Prefilled 10% Formalin Pot | Sufficient to fully immerse the specimen | Pre-filled container with lid firmly closed | None | 80% of cases by Day 10 |
| GI Endoscopic Biopsy (P02) | Fixed Tissue | Prefilled 10% Formalin Pot | Sufficient to fully immerse the specimen | Pre-filled container with lid firmly closed | None | 80% of cases by Day 10 |
| Non-Biopsy Cancer Resection (P03) | Fixed Tissue | Prefilled 10% Formalin Pot | Sufficient to fully immerse the specimen | Pre-filled container with lid firmly closed | None | 80% of cases by Day 7 |

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| Test/Profile/P Code | Specimen Type | Specimen Requirements | | | Specimen Requirements | Turnaround Time |
|---|-----------------------------|--|--|---|---|------------------------|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container /Type</u> | | |
| Non-Biopsy Other (P04) | Fixed Tissue | Prefilled 10% Formalin Pot | Sufficient to fully immerse the specimen | Pre-filled container with lid firmly closed | None | 80% of cases by Day 14 |
| Non Gynae Cytology FNA (P06) | Fluid/FNA/ Fixed slide | Neat or in Saline or Cytospin Collection fluid | Approx 10mls | Standard sterile Universal container | None | 80% of cases by Day 10 |
| Non Gynae Cytology Exfoliative (P07) | Fluid/Brush/ Fixed Slide | Neat or in Saline or Cytospin Collection fluid | Approx 10mls | Standard sterile Universal container | None | 80% of cases by Day 10 |
| Frozen Section (Q007) | Fresh Tissue | None | N/A | Dry/ empty Histopathology Container | <ul style="list-style-type: none"> Tissue for frozen sections must be hand delivered to Specimen Reception between 9 - 4.00pm Mon – Fri. 3 hrs notice to the Lab is required for planned frozen sections. | 20 mins |

Factors that may impact target TAT include the following:-

- Requirement to obtain additional clinical/radiological information
- Requirement for longer fixation in some cases (TAT may take >= 20 working days)
- Requirement for specimen decalcification
- Requirement to examine a large number of blocks/slides in a case
- Requirement for ancillary testing including levels, immunohistochemistry, special stains and Molecular Pathology
- Requirement for Intradepartmental Consultation

For cases sent externally for reporting (e.g. Gynae cytology cases, cases that require direct immunofluorescence), our target TAT is as per the target TAT stated in the Histopathology laboratory manual for the department reporting the case.

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12.2 Molecular Genetic Testing on Tumours

| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|--|--------------------------------|--------------------------|---------------------------|------------------------|--|-----------------|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container /Type</u> | | |
| Colorectal Panel* | Paraffin Embedded Fixed Tissue | N/A | N/A | N/A | Pathologist selected material upon request | 10 working days |
| Gastric Her2* | Paraffin Embedded Fixed Tissue | N/A | N/A | N/A | Selected carcinomas requiring ISH confirmation. | 10 working days |
| GIST Panel* | Paraffin Embedded Fixed Tissue | N/A | N/A | N/A | Pathologist selected material upon request | 15 working days |
| Her 2 ISH Analysis (Breast Carcinomas)* | Paraffin Embedded Fixed Tissue | N/A | N/A | N/A | Selected breast carcinomas requiring ISH confirmation. Notify Lab of urgent cases. | 10 working days |
| Lung Panel* | Paraffin Embedded Fixed Tissue | N/A | N/A | N/A | Pathologist selected material upon request | 10 working days |
| Melanoma Panel* | Paraffin Embedded Fixed Tissue | N/A | N/A | N/A | Pathologist selected material upon request | 10 working days |
| Microsatellite Instability* | Paraffin Embedded Fixed Tissue | N/A | N/A | N/A | Pathologist selected material upon request. Requires MMR first, liaise with Pathologist. | 20 working days |
| Oncomine* | Paraffin Embedded Fixed Tissue | N/A | N/A | N/A | Pathologist selected material upon request | 20 working days |

* These specimens/ samples are referred to external laboratories for testing.

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12.3 CYTOPATHOLOGY













| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements |
|--|---------------|--------------------------------------|--------------------------------|-----------------------|--|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | |
| Biliary Brush Sample | Brush | Cytology Collection Fluid | N/A | Universal | None |
| Body Fluid (Various) | Fluid | None | N/A | Universal | Where possible, if Microbiology required, please send additional sample. |
| Bronchial Lavage | Fluid | None | N/A | Universal | Where possible, if Microbiology required, please send additional sample. |
| Cervical Specimen (Thin Prep) and HPV* | Fluid | Prefilled Thin Prep Collection Fluid | N/A | Thin Prep Vial | Click here to access Biomnis Cytology Test Request Form and send with the sample. |
| Cerebrospinal Fluid (CSF) | Fluid | None | Refer to form PL012 for volume | Universal | Where possible, if Microbiology required, please send additional sample. Request this test using CSF form PL012. |
| Fine Needle Aspirate (FNA) (Breast) | Slides | None | N/A | Slide Holder | Alcohol spray fix at time of preparation |
| Fine Needle Aspirate (FNA) (Breast) | Fluid | Cytospin Collection Fluid | N/A | Universal | None |
| Fine Needle Aspirate (FNA) (Not Breast) | Slides | None | N/A | Slide Holder | Alcohol spray fix at time of preparation |
| Fine Needle Aspirate (FNA) (Not Breast) | Fluid | Cytospin Collection Fluid | N/A | Universal | None |
| Hepatic Brush Sample | Brush | Cytospin Collection Fluid | N/A | Universal | None |
| Sputum | Sputum | None | N/A | Universal | Where possible, if Microbiology required, please send additional sample. |

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13.0 IMMUNOLOGY














| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|---|---------------|---|---------------------------|-----------------------|--|---|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| Adrenal Antibody* | Blood | None  | 4.9 | Blood Tube | None | 30 working days |
| ANCA (Anti-Neutrophil Cytoplasmic Antibodies) | Blood | None  | 4.9 | Blood Tube | Notify Immunology lab @ 1997 of all urgent ANCA requests | Urgent - 1 working day Routine BSHC - 2 working days Routine other sites – 3 working days |
| Antinuclear Antibody (ANA) | Blood | None  | 4.9 | Blood Tube | None | 5 working days |
| Aquaporin 4 Antibody* | Blood | None | 4.9 | Blood Tube | None | 20 working days |
| Autoantibody Screen (ANA, AMA, LKM, SMA, GPCA) | Blood | None  | 4.9 | Blood Tube | None | 5 working days |
| Avian Precipitins* | Blood | None  | 4.9 | Blood Tube | None | 15 working days |
| Bullous Antibodies* | Blood | None  | 4.9 | Blood Tube | None | 20 working days |
| CASPR2 Antibody* | Blood | None  | 4.9 | Blood Tube | None | 20 working days |
| CASPR2 Antibody* | CSF | None | 0.5 | Universal | Request this test using CSF form PL012. | 20 working days |
| Cyclic Citrullinated Peptide Antibody (CCP) | Blood | None  | 4.9 | Blood Tube | None | 3 working days |
| dsDNA Antibody | Blood | None  | 4.9 | Blood Tube | None | 5 working days |
| Endomysial IgA Antibody | Blood | None  | 4.9 | Blood Tube | None | 4 working days |
| Extractable Nuclear Antigen (ENA Screen) | Blood | None  | 4.9 | Blood Tube | None | 3 working days |
| GABA and AMPA Receptor Antibodies* | Blood | None  | 4.9 | Blood Tube | None | 20 working days |
| GABA and AMPA Receptor Antibodies* | CSF | None | 0.5 | Universal | Request this test using CSF form PL012. | 20 working days |

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IMMUNOLOGY..... cont'd










| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|---|---------------|---|---------------------------|-----------------------|---|-----------------|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| Ganglioside Antibodies* (Anti GM1 and GQ1b) | Blood | None  | 4.9 | Blood Tube | None | 20 working days |
| Ganglioside Antibodies* (Anti GM1 and GQ1b) | CSF | None | 0.5 | Universal | None | 20 working days |
| Ganglionic Acetylcholine Receptor Antibody* | Blood | None  | 4.9 | Blood Tube | None | 40 working days |
| Glomerular Basement Membrane Antibody (GBM) | Blood | None  | 4.9 | Blood Tube | None | 5 working days |
| Gliadin Antibodies (IgA, IgG)* | Blood | None  | 4.9 | Blood Tube | None | 10 working days |
| Glycine Receptor Antibody* | Blood | None  | 4.9 | Blood Tube | None | 20 working days |
| Intrinsic Factor Antibody | Blood | None  | 4.9 | Blood Tube | None | 3 working days |
| Islet Cell Antibody* | Blood | None  | 4.9 | Blood Tube | None | 30 working days |
| LGI1 Antibody* | Blood | None  | 4.9 | Blood Tube | None | 20 working days |
| LGI1 Antibody* | CSF | None | 0.5 | Universal | Request this test using CSF form PL012. | 20 working days |
| MUSK Antibody* | Blood | None  | 4.9 | Blood Tube | None | 20 working days |
| Myelin Associated Glycoprotein Antibody (MAG)* | Blood | None  | 4.9 | Blood Tube | None | 10 working days |
| Myelin Oligodendrocyte Glycoprotein Antibody (MOG)* | Blood | None  | 4.9 | Blood Tube | None | 20 working days |
| Myositis Marker Antibodies (Anti Synthetase Antibodies)* | Blood | None  | 4.9 | Blood tube | None | 15 working days |
| Neuronal Antibodies* | Blood | None  | 4.9 | Blood Tube | None | 20 working days |

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IMMUNOLOGY..... cont'd

| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|--|---------------|---|--------------------------------|-----------------------|----------------------|-----------------|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| NMDA Receptor Antibody* | Blood | None  | 4.9 | Blood Tube | None | 20 working days |
| NMDA Receptor Antibody* | CSF | None | Refer to form PL012 for volume | Universal | None | 20 working days |
| Ovarian Antibody* | Blood | None  | 4.9 | Blood Tube | None | 20 working days |
| Phospholipid Antibodies* (IgG and IgM Cardiolipin and β 2 Glycoprotein Antibodies) | Blood | None  | 4.9 | Blood Tube | None | 20 working days |
| Rheumatoid Screen (RF Latex & CCP) | Blood | None  | 4.9 | Blood Tube | None | 3 working days |
| Thyroglobulin Antibody* | Blood | None  | 4.9 | Blood Tube | None | 10 working days |
| Thyroid Peroxidase Antibody (TPO) | Blood | None  | 4.9 | Blood Tube | None | 3 working days |
| TSH Receptor Antibody* | Blood | None  | 4.9 | Blood Tube | None | 10 working days |
| tTG IgA Antibody (Positive tTG will be followed by an Endomysial IgA Antibody Test) | Blood | None  | 4.9 | Blood Tube | None | 3 working days |
| Voltage Gated Calcium Channel (VGCC) Antibody* | Blood | None  | 4.9 | Blood Tube | None | 20 working days |

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14.0 MICROBIOLOGY TESTS

14.1 General Bacteriology

Turnaround times are based on routine specimens. In some cases, additional testing and/ or referral may be required. This will extend expected turnaround times.

Amies charcoal swabs are manufactured with raw material products which are Latex free.

| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|--|---|--------------------------|--|--|--|--|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| Acid Fast Bacilli Culture (AFB/ TB Culture) (Mycobacterium Species) | Sputum Pus Pleural fluid Other body fluids | None | 5-10 as available as available as available | Sterile universal container | Refer to section 14.3.1 | Up to 45 days Interim AAFB smear report available @ 1 working day |
| | Bone marrow | None | as available | Special bone marrow/ TB culture bottle | Bottle available from Microbiology dept. on request | |
| | Urine (EMU) | None | 10-20 | Sterile universal container | | |
| | Tissue | None | as available | Sterile universal container | Do not use Formalin with tissue | |
| | Bronchial lavage\brushings | None | as available | Specimen trap container | | |
| | Swabs | None | N/A | Charcoal Swabs | 2 pus swabs required or tissue accompanied by pus swab | |

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MICROBIOLOGY - General Bacteriology cont'd

| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|---|---|--------------------------|---|---|--|---|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| Blood Cultures for Adult | Blood | None | 10 added to each bottle | BacT Alert Bottles aerobic (green) and anaerobic (orange) | Refer to section 14.3.2 of this document | Positives: Gram Stain: 5 hrs Preliminary Culture: 1 working day from detection Full Report and Susceptibility: 2 working days from detection Negatives: 5 working days except in cases of infective endocarditis where it is 10 days. Updated reports available on PIMS ♦ |
| Blood Cultures for Babies and Children | Blood | None | 1-4 per bottle | Paediatric BacT Alert Bottle Paediatric (yellow) | Refer to section 14.3.2 of this document | Positives: Gram Stain: 5 hrs Preliminary Culture: 1 working day from detection Full Report and Susceptibility: 2 working days from detection Negatives: 5 working days except in cases of infective endocarditis where it is 10 days. Updated reports available on PIMS ♦ |
| Bacterial PCR* | Fluids/ Tissues from normally sterile sites | None | 0.5 mL or Fingernail size piece of tissue | Sterile Universal Container | None | 15 Working Days |

- * These specimens/ samples are referred to external laboratories for testing.
♦ Tests provided in the emergency out of hours service.

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MICROBIOLOGY - General Bacteriology cont'd

| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|---|-------------------------------|--------------------------|--------------------------------|---|---|--|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| Cerebrospinal Fluid for Cryptococcus Culture | CSF | None | Refer to form PL012 for volume | Sterile universal container | None | 4 working days |
| Cerebrospinal Fluid for Microscopy, Culture and Susceptibility | CSF | None | Refer to form PL012 for volume | Sterile universal container | Refer to section 14.3.3 of this document | 2 working days ♦ Interim verbal microscopy results available within 2 hrs |
| COVID 19 PCR | Naso-pharyngeal & Throat Swab | None | Viral UTM Swab | Viral UTM Swab  | Refer to section 14.3.18 | Up to 2 working days |
| Faeces For Clostridium difficile Toxin | Faeces | None | 5-10 | Sterile universal container | Detection of Clostridium difficile Toxin A and B in suspect pseudo-membranous colitis or antibiotic-associated diarrhoea or where indicated. | 1 working day |

♦ Tests provided in the emergency out of hours service.

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MICROBIOLOGY TESTS – General Bacteriology.....cont'd

| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|--|---------------|--------------------------|---------------------------|---|--|-----------------|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| Faeces for Clostridium difficile antigens – GDH Glutamate dehydrogenase | Faeces | None | 5-10 | Sterile universal container | Detection of Clostridium difficile antigens (predominately Glutamate dehydrogenase GDH) | 1 working day |
| Faeces for Cryptosporidium | Faeces | None | 5-10 | Sterile universal container | This test will routinely be performed on all samples from the children's ward and all fluid samples. | 2 working days |
| CPE/ESBL/VRE Screen | Rectal Swab | None | Not relevant | Copan FecalSwab  | Refer to section 14.3.14 VRE Screens | 4 working days |
| Faeces For Intestinal Pathogens | Faeces | None | 5-10 | Sterile universal container | Faecal Culture for Salmonella, Shigella, Campylobacter, E. coli 0157 and other gastrointestinal pathogens depending on clinical details supplied. Please include history of foreign travel and/ or consumption of shellfish. | 2 working days |

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MICROBIOLOGY TESTS – General Bacteriology.....cont'd

| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|---|--|--------------------------|---------------------------|---|--|---|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| Faeces Occult Blood | Faeces | None | 5-10 | Sterile universal container | None | 1 working day |
| Faeces For Ova Cysts and Parasites* | Faeces | None | 5-10 | Sterile universal container | See 14.3.15 Please state clinical history including foreign travel. | 10 working days |
| Faeces for Rotavirus and Adenovirus | Faeces | None | 5-10 | Sterile universal container | This test is performed routinely on children 5 years or age or less. | 2 working days |
| Fluid from Sterile Site for Culture and Susceptibility | Amniotic fluid Ascitic fluid Bursa fluid Pericardial fluid Peritoneal fluid Pleural fluid | None | 5-10 | Sterile universal container | Refer to sections 14.3.4 and 14.3.5 of this document | 2 working days |
| | Synovial (Joint) fluid | | | | | ◆ |
| Helicobacter pylori culture* | Two gastric biopsies should be taken, one from the antrum and one from the body of the stomach which are embedded in Portagerm pylori transport medium | None | Not relevant | Portagerm pylori medium is available from Micro | Micro must be notified 1 day in advance of biopsies being taken and samples must be transported to Microbiology ASAP | 15 working days |
| MRSA Screen | Miscellaneous Swabs | None | Not relevant | Charcoal Swab | Refer to section 14.3.12 MRSA screens | 2 working days |
| Mycology, Fungal Culture | Skin Scrapings, Hairs, Nail Clippings, Tissue | None | 5 pieces | Sterile Universal container or Dermapak | Refer to section 14.3.5 of this document | Up to 15 working days. Interim microscopy reports available @ 1 working day |

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These specimens/ samples are referred to external laboratories for testing.

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| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|--|--|--------------------------|---------------------------|-----------------------------|--|--|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| Pharmacy Culture | TSA plates and SAB plates | None | N/A | Agar plate | Fungal growth phoned directly to Pharmacy | 2 working days for TSA plates 7 working days for SAB plates |
| Sputum for Culture and Sensitivities Bronchial Lavage for culture and sensitivity | Sputum Bronchial lavage/aspirate/brushings/washings/alveolar lavage Transthoracic/ Transtracheal aspirate | None | 5-10 | Sterile universal container | Refer to section 14.3.6 of this document | 2 working days |

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| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|---|----------------|--------------------------|---------------------------|--|--|----------------------|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| Swabs for Culture and Sensitivity including ear, eye, mouth, throat, nasal, high vaginal swab (HVS), penile, cervical, urethral, wound and ulcer | Swabs | None | Not relevant | Special adult ear swab (charcoal) available from Microbiology dept.  | Refer to sections 14.3.7 Eye 14.3.8 Throat 14.3.9 Nasal 14.3.10 HVS, penile and cervical 14.3.11 Wound 14.3.5 Fungal | 2 working days |
| Swabs for Whooping Cough (Bordetella pertussis) | Per nasal Swab | None | Not Relevant | Special pernasal swab available from Microbiology dept.  | None | Up to 7 working days |

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MICROBIOLOGY TESTS – General Bacteriology.....cont'd







| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|--|---|--------------------------|---------------------------|--|--|------------------|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| Tissue for Culture and Sensitivity | Tissue | None | Not Relevant | Universal container filled with 0.85% saline and ballotini beads | Refer to section 14.3.5 Do not use Formalin | 2 working days |
| Tips for culture and sensitivity | CVP or Hickman lines Central/arterial /portacath/ venous tips Cannula tips | None | Not Relevant | Sterile universal container | The culture of urinary catheter tips is not clinically useful and they will not be processed in the laboratory. For central/ arterial / portacath tips please send distal 4 cms (cut using sterile scissors) | 2 working days |
| Urine for Microscopy, culture and sensitivities | Urine MSU CSU CATCH BAG Supra Pubic Ileal Conduit | None | 10 | Sterile universal container | Refer to section 14.3.13 of this document | 2 working days ♦ |
| Urine Bilirubin | Urine | None | 5-20 | Sterile Universal Container | None | 1 working day ♦ |

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14.2 **MICROBIOLOGY TESTS – Serology**

*Please see section 14.3.17 for general notes and requesting guide for Virology/
Serology tests*








| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|---|---------------|---|---------------------------|-----------------------|---|--|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| Aciclovir Pre Levels* | Blood | None  | 4.9 | Blood Tube | None | 15 working days |
| Aciclovir Post Levels* | Blood | None  | 4.9 | Blood Tube | Post dose sample should be taken either 1 hr after the end of IV administration or 2 hrs after oral administration. | 15 working days |
| Adenovirus Antibodies* | Blood | None  | 4.9 | Blood Tube | None | 15 working days |
| Antibiotic assay Aminoglycosides including Amikacin* Gentamicin Tobramycin | Blood | None  | 4.9 | Blood Tube | Refer to Gentamicin & Tobramycin Once Daily Guideline Refer to Endocarditis Infection Guideline. Refer to Vancomycin Dosing & Monitoring Guideline Refer to Amikacin Dosing and Monitoring Guideline | Gentamicin 3 hours ♦ Vancomycin 3 hours ♦ |
| Glycopeptides including Vancomycin Teicoplanin* | | | | | Amikacin Teicoplanin | 1 working day 5 working days |
| Anti Streptolysin – O Titre (ASOT) | Blood | None  | 4.9 | Blood Tube | Useful for the investigation of Group A Beta-haemolytic Streptococcal infection or post-streptococcal disorder e.g. rheumatic fever. Normal Findings: Titre of 0 - 200 Todd units per ml. | 1 working day |
| Amoebiasis Antibodies* | Blood | None  | 4.9 | Blood Tube | None | 15 working days |

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MICROBIOLOGY TESTS – Serology.....cont'd

| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|--|---------------|---|---------------------------|-----------------------|---|-----------------|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| Aspergillus Antibodies* | Blood | None  | 4.9 | Blood Tube | Aspergillus <u>antibody</u> titres may be useful in the investigation of allergic broncho-pulmonary aspergilloma. | 15 working days |
| Aspergillus Antigen (Galactomannan (GM))* (Acute Infection) | Blood | None  | 4.9 | Blood Tube | Aspergillus <u>antigen</u> detection may be useful in invasive aspergillosis. See section 14.3.17. | 15 working days |
| Beta Glucan Antigen* | Blood | None  | 4.9 | Blood Tube | Aspergillus <u>antigen</u> detection may be useful in invasive aspergillosis. See section 14.3.17. | 15 working days |
| Bartonella henselae Antibodies (CATSCRATCH FEVER)* | Blood | None  | 4.9 | Blood Tube | Special form available in the Microbiology dept. | 15 working days |
| Blastomyces Antibodies* | Blood | None  | 4.9 | Blood Tube | Clinical requests for this test need to have a travel history. Click here to access referral lab request form and send with the sample. | 15 working days |
| Bordetella pertussis Anti-toxin Antibodies* | Blood | None  | 4.9 | Blood Tube | None | 15 working days |
| Brucella Serology (Titre)* | Blood | None  | 4.9 | Blood Tube | Clinical information is essential. Please complete relevant clinical details on back of Serology test request form. | 15 working days |

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| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|---|---|---|-------------------------------------|---|--|-----------------|
| | | <u>Additive Required</u> | <u>Volume Required</u> <u>mL</u> | <u>Container Type</u> | | |
| Campylobacter Antibodies* | Blood | None  | 4.9 | Blood Tube | Note: Campylobacter serology is NOT available for the diagnosis of acute Campylobacter infection. The test is valid only for possible sequelae arising from previous Campylobacter infections. | 15 working days |
| Chikungunya Serology* | Blood | None  | 4.9 | Blood Tube | Test is only performed following prior discussion with the Reference Laboratory | 15 working days |
| Chlamydia pneumoniae IgM* | Blood | None  | 4.9 | Blood Tube | Refer to section 14.3.17 Restricted test: will only be referred following approval from Consultant Microbiologist. | 15 working days |
| Chlamydia sp. specific Antibodies* (includes C. trachomatis, C. pneumoniae and C. psittaci) | Blood | None  | 4.9 | Blood Tube | None | 15 working days |
| Chlamydia trachomatis/ Gonorrhoea/ Trichomonas (PCR)* | Urethral Swab Cervical Swab Eye Swab Urine | None | Not relevant | Special swabs available from Micro Sterile universal container | None | 10 working days |
| Creutzfeldt-Jakob Disease* (14-3-3 Protein) | CSF | None | CSF no. 3 with min volume of 1.5 | Sterile universal container | Click here to access CJD Request Form and send with the sample. Samples need to be sent to the Lab within 30 mins of aspiration. It is essential that clinical details, MRI and EEG results are included. | 35 working days |

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MICROBIOLOGY TESTS – Serology.....cont'd

| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|--|---|---|--------------------------------|----------------------------------|--|-----------------|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| Cytomegalovirus CMV Culture* | Viral Throat Swab, Urine, BAL | None | 5 | Swab Sterile universal container | None | 15 working days |
| Cytomegalovirus (CMV) PCR* | Blood | Potassium EDTA Whole Blood, Plasma  | 4.9 | Blood Tube | Please send to Laboratory immediately as sample must be frozen | 15 working days |
| | CSF Urine Bronchial Lavage Throat Swab | | 0.5 | Sterile universal container | For CSF specimens, document request on form PL012. | |
| CMV IgG* | Blood | None  | 4.9 | Blood Tube | None | 10 working days |
| CMV IgM | Blood | None  | 4.9 | Blood Tube | None | 3 working days |
| Coccidioides Antibodies* | Blood | None  | 4.9 | Blood Tube | Clinical requests for this test need to have a travel history. Click here to access referral lab request form and send with the sample. | 15 working days |
| Coxiella burnetii* (Q Fever Antibodies) | Blood | None  | 4.9 | Blood Tube | Refer to section 14.3.17. Please complete relevant clinical details on back of Serology test request form. | 15 working days |
| Cryptococcal Antigen* | Blood | None  | 4.9 | Blood Tube | None | 15 working days |
| | CSF | N/A | Refer to form PL012 for volume | Sterile Universal Container | | |
| Dengue Virus Serology* | Blood | None  | 4.9 | Blood Tube | None | 15 working days |

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| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|-----------------------------------|---|---|---|---|--|-----------------|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| Diphtheria IgG (Immunity)* | Blood | None  | 4.9 | Blood Tube | None | 15 working days |
| Echinococcus Antibodies* | Blood | None  | 4.9 | Blood Tube | None | 15 working days |
| Enterovirus Culture* | Faeces Pleural Fluid CSF | None | 10-20 Refer to form PL012 for volume | Sterile Universal Container | None | 15 working days |
| | Viral Throat Swab | None | 1 | Viral Swab | | |
| Enterovirus PCR* | CSF Stool Respiratory Excretions Viral Skin Swab Viral Throat Swab Vesicle Fluid | None | 0.5 | Sterile Universal Container Viral Swab | For CSF specimens, document request on form PL012. | 15 working days |
| Epstein-Barr virus IgG* | Blood | None  | 4.9 | Blood Tube | None | 15 working days |
| Epstein-Barr virus IgM* | Blood | None  | 4.9 | Blood Tube | None | 15 working days |
| Epstein-Barr virus PCR* | Blood | Potassium EDTA  | 4.9 | Blood Tube | Transport to the lab immediately. Specimen must be spun and separated within 6 hrs. Specimen is sent frozen to the reference lab. | 15 working days |
| Farmers Lung Antibodies* | Blood | None  | 4.9 | Blood Tube | None | 15 working days |

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| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|--|---------------|--|---------------------------|-----------------------------|--|-----------------|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| Filaria Serology* | Blood | None  | 4.9 | Blood Tube | None | 15 working days |
| Francisella tularensis* | Blood | None  | 4.9 | Blood Tube | None | 15 working days |
| Haemophilus influenzae Antibodies* | Blood | None  | 4.9 | Blood Tube | None | 15 working days |
| Haemophilus influenzae PCR* | Blood | Potassium EDTA  | 2.7 (1mL minimum) | Blood Tube | Sample taken pre-antibiotic. Transport to the lab immediately. | 10 working days |
| | CSF | None | 0.5 | Sterile Universal Container | For CSF specimens, document request on form PL012. | |
| Helicobacter pylori IgG Antibodies* | Blood | None  | 4.9 | Blood Tube | None | 15 working days |
| Hepatitis A Antibodies Total* | Blood | None  | 4.9 | Blood Tube | None | 10 working days |
| Hepatitis A IgM | Blood | None  | 4.9 | Blood Tube | None | 2 working days |
| Hepatitis B Surface Antibodies (Immunity Check) Occupational Health Additional Method | Blood | None  | 4.9 | Blood Tube | None | 3 working days |
| | Blood | None  | 4.9 | Blood Tube | None | 15 working days |
| Hepatitis B Core Antibodies | Blood | None  | 4.9 | Blood Tube | None | 2 working days |
| Hepatitis B DNA* | Blood | None  Potassium EDTA  | 4.9 | Blood Tube | Transport to the lab immediately. Specimen must be spun and separated within 6 hrs. Specimen is sent frozen to the reference lab. | 15 working days |
| Hepatitis B (VRL Markers)* | Blood | None  | 4.9 | Blood Tube | None | 15 working days |

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| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|--|---------------|--|--------------------------------|-----------------------|---|-----------------|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| Hepatitis B surface Antigen (HbsAg) | Blood | None  | 4.9 | Blood Tube | None | 2 working days |
| Hepatitis C Antibodies | Blood | None  | 4.9 | Blood Tube | None | 2 working days |
| Hepatitis C PCR (Polymerase Chain Reaction)* | Blood | Potassium EDTA Whole Blood Plasma  | 9 | Blood Tube | Deliver specimen to laboratory immediately | 10 working days |
| Hepatitis D Antibodies* | Blood | None  | 4.9 | Blood Tube | None | 15 working days |
| Hepatitis E Antibodies* | Blood | None  | 4.9 | Blood Tube | None | 15 working days |
| Herpes simplex Antibodies* | Blood | None  | 4.9 | Blood Tube | Please discuss with Microbiologist | 15 working days |
| Herpes simplex PCR* | Viral Swab | N/A | N/A | Viral Swab | None | 15 working days |
| | CSF | None | Refer to form PL012 for volume | Sterile Universal | None | 15 working days |
| Histoplasma Antibodies* | Blood | None  | 4.9 | Blood Tube | Clinical requests for this test need to have a travel history. Click here to access referral lab request form and send with the sample. | 15 working days |

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|-----------------------------------|--|---|--|--|--|---|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| HIV PCR* | Blood | Potassium EDTA  | 4.9 | Blood Tube | Transport to the lab immediately. Specimen must be spun and separated within 6 hrs. Specimen is sent frozen to the reference lab. | 20 working days |
| HIV Antigen/ Antibody Test | Blood | None  | 4.9 | Blood Tube | None | 2 working days |
| HTLV 1 and 2* | Blood | None  | 4.9 | Blood Tube | None | 15 working days |
| Human Herpes virus 6* | Blood | None  | 4.9 | Blood Tube | None | 15 working days |
| Influenzae A and B | Naso-pharyngeal Aspirate or nasal and throat swab only | None | 1mL Naso-pharyngeal Aspirate Viral UTM Swab | Sterile Universal Container or Viral UTM Swab  | Refer to section 14.3.17 | 3 hours during routine hours Test performed between 9am – 11pm 7 days a week ♦ |
| Extended Resp. Panel | Nasal and throat swab only | None | Viral UTM Swab | Viral UTM Swab  | Standard test for CCU/Oncology. All other requests Consultant to Consultant Microbiologist only. Refer to section 14.3.17 | 1 working day |

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| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|---|---------------------------------|---|---------------------------|---|--|-----------------|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| Legionella pneumophila urinary antigen test | Urine | None | 20 | Sterile Universal Container | Refer to section 14.3.17 | 2 working days |
| Legionella Culture* | Sputum or Bronchial Lavages | None | 4 | Sterile Universal Container | None | 15 working days |
| Leishmania Antibodies* | Blood | None  | 4.9 | Blood Tube | None | 15 working days |
| Leishmania Detection* | Skin Bone Marrow Biopsies | None | N/A | Sterile Universal Container | Transport to the Laboratory immediately | 15 working days |
| Leptospira Antibodies* | Blood | None  | 4.9 | Blood Tube | None | 10 working days |
| Lymes Disease (Borrelia burgdorferi) Antibodies* | Blood | None  | 4.9 | Blood Tube | Please complete relevant clinical details on back of Serology test request form. | 15 working days |
| | CSF | None | 500µl | Sterile Universal Container | | 15 working days |
| Lymphoma Viral Screen | Blood | None  | 4.9 | Blood Tube | Refer to section 14.3.17 | 2 working days |
| Measles IgG* | Blood | None  | 4.9 | Blood Tube | None | 15 working days |
| Measles IgM* | Blood | None  | 4.9 | Blood Tube | Click here to access Oral Fluid Specific Request Form | 15 working days |
| | Saliva | Oracol Swab (i.e. Salivary Swab) | N/A | Special foam salivary swabs available in Micro. | | |
| Measles RNA PCR* | Saliva | Oracol Swab (i.e. Salivary Swab) | N/A | Special foam salivary swabs available in Micro. | Click here to access Oral Fluid Specific Request Form | 15 working days |
| CSF PCR Panel | CSF | None  | 0.5 | Sterile Universal Container | Document request on form PL012. | 1 working day |

* These specimens/ samples are referred to external laboratories for testing.

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| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|---|-------------------------------------|---|---------------------------|---|--|---|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| Mumps IgG* | Blood | None  | 4.9 | Blood Tube | None | 15 working days |
| Mumps IgM* | Blood | None  | 4.9 | Blood Tube | None | 15 working days |
| | Saliva | Salivary Swab | N/A | Special foam salivary swabs available in Microbiology | Click here to access Oral Fluid Specific Request Form | |
| Mycobacterium TB PCR* | CSF or other normally sterile fluid | None | 0.5 | Sterile Universal Container | For CSF specimens, document request on form PL012 | 15 working days |
| Mycoplasma pneumoniae IgM | Blood | None  | 4.9 | Blood Tube | None | 1 working day |
| Occupational Blood/ Body Fluid Exposure <u>Source Tests:</u> <ul style="list-style-type: none"> Hepatitis B Surface Antigen Hepatitis C Antibodies HIV (if patient consented) <u>Recipient Tests:</u> Hepatitis B Surface Antibodies | Blood | None  | 4.9 | Blood Tube | Refer to Hospital Policy OCC0009 State clearly on form source or recipient of Occupational Blood/ Body Fluid Exposure injury | "Verbal" results are available within 4 hours ♦ |
| | Blood | None  | 4.9 | Blood Tube | | |
| Neisseria meningitidis Antibodies* | Blood | None  | 4.9 | Blood Tube | Convalescent sample 2-3 weeks post onset | 15 working days |
| Parvovirus B19 IgG (Non Acute) Serology* | Blood | None  | 4.9 | Blood Tube | None | 15 working days |




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♦ Tests provided in the emergency out of hours service.

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| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|--|--|---|---|-----------------------------|---|-----------------|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| Pneumococcal Antibodies IgG* | Blood | None  | 4.9 | Blood Tube | None | 15 working days |
| Neisseria meningitidis PCR* | Blood | Potassium EDTA  | 2.7 (0.5mL minimum) | Blood Tube | Sample taken pre-antibiotic (Specimens should be received in the Lab. before midday for transport to the reference Lab. the next day). Click here to access specific request form. | 10 working days |
| | CSF | None | >0.5 Request this test using CSF form PL012. | Sterile Universal Container | | |
| PCR for Norovirus (previously SRSV, “Winter Vomiting Bug”)* | Faeces | None | 10-20 | Sterile Universal Container | None | 1 working day |
| Parainfluenzae virus Immunofluorescence* | Naso-pharyngeal aspirate or mouth washings | None | 2 | Sterile Universal Container | None | 15 working days |
| Parvovirus B19 IgM (Acute) Serology* | Blood | None  | 4.9 | Blood Tube | None | 15 working days |
| Pneumococcal Urinary Antigen | Urine | None | 2 | Sterile Universal container | None | 2 working days |
| Pneumocystis jiroveci PCR* (Formally known as Pneumocystis carinii (PCP)) | Bronchial Lavage/ Sputum | None | 5 | Sterile Universal Container | None | 15 working days |

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| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|--|----------------------------|---|---------------------------|-----------------------------|---|-----------------------|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| Polyoma Virus (JC)* | Blood | None  | 4.9 | Blood Tube | Transport to the lab immediately. Blood specimen needs to be frozen within 24 hrs. | 15 working days |
| | CSF | | 0.5 | Sterile Universal Container | Request this test using CSF form PL012. | |
| | Urine | | 10 | Sterile Universal Container | | |
| Polyoma BK Virus* | Blood | None  | 4.9 | Blood Tube | Transport to the lab immediately. Blood specimen needs to be frozen within 24 hrs. | 15 working days |
| | Urine | | 10 | Sterile Universal Container | | |
| Procalcitonin | Blood | None  | 4.9 | Blood Tube | None | 2 working days |
| Quantiferon test for MTB Complex | Whole Blood | Heparin anti-clotting agent | 4 x 1 bottles | Blood tubes in box supplied | Special Tubes available from Phlebotomy by request. | 2 working days |
| Reagent Strip Urinalysis | Urine | None | 1 | Sterile Universal Container | Reagent Strip Urinalysis is performed if requested as part of a urine culture but may be requested without urine culture. | 1 working days |
| Respiratory Syncytial Virus Antigen (RSV) | Nasophar - yngeal aspirate | None | 2 | Sterile Universal Container | None | 2 working days ♦ |
| Rickettsia Antibodies* | Blood | None  | 4.9 | Blood Tube | None | 15 working days |

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|---|---------------|---|---------------------------|-----------------------------|---|-----------------|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| Rubella IgG (Immunity) | Blood | None  | 4.9 | Blood Tube | None | 2 working days |
| Rubella IgM* | Blood | None  | 4.9 | Blood Tube | None | 15 working days |
| Saccharomyces cerevisiae Antibodies* | Blood | None  | 4.9 | Blood Tube | None | 15 working days |
| Schistosomal Antibodies* | Blood | None  | 4.9 | Blood Tube | None | 15 working days |
| Strep. Pneumoniae PCR* | Blood | Potassium EDTA  | 2.7 (1mL minimum) | Blood Tube | Sample taken pre-antibiotic (Specimens should be received in the Lab. before midday for transport to the reference Lab. the next day). Request this test using CSF form PL012. | 10 working days |
| | CSF | None | >0.5 | Sterile Universal container | | |
| Group B Streptococcal PCR* | Blood | Potassium EDTA  | 2.7 (1mL minimum) | Blood Tube | Sample taken pre-antibiotic. Transport to the lab immediately. Performed on patients <3 mths old. Please contact Lab if patient is >3 mths old. Request this test using CSF form PL012. | 10 working days |
| | CSF | None | >0.5 | Sterile Universal Container | | |
| Strongyloides Antibodies* | Blood | None  | 4.9 | Blood Tube | None | 15 working days |
| Total Syphilis Antibodies* | Blood | None  | 4.9 | Blood Tube | None | 10 working days |
| Tetanus IgG (Immunity)* | Blood | None  | 4.9 | Blood Tube | None | 15 working days |

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|--|-------------------------------|---|---------------------------|---|--|-----------------|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| Toxocara Antibodies* | Blood | None  | 4.9 | Blood Tube | Special form available in the Microbiology dept. | 15 working days |
| Toxoplasma IgM | Blood | None  | 4.9 | Blood Tube | None | 2 working days |
| Toxoplasma IgG* | Blood | None  | 4.9 | Blood Tube | None | 10 working days |
| Trichinosis Antibodies* | Blood | None  | 4.9 | Blood Tube | None | 15 working days |
| Urine for Pneumococcal Antigen | Urine | None | 2 | Sterile Universal container | None | 2 working days |
| Varicella zoster DNA* | Viral swab of vesicular fluid | None | None | Viral UTM Swab  | None | 15 working days |
| Varicella zoster IgG Antibodies | Blood | None  | 4.9 | Blood Tube | None | 2 working days |
| Viral Culture* | Faeces | None | 1-20 | Sterile Universal Container | Special viral swab available in the Microbiology dept. | 15 working days |
| | other body fluids Swab | None | None | Viral UTM Swab  | | |
| West Nile Virus IgM* | Blood | None  | 4.9 | Blood Tube | None | 15 working days |

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14.3 **SPECIAL REQUIREMENTS FOR MICROBIOLOGY SAMPLING AND TESTING**

14.3.1 **MYCOBACTERIA TUBERCULOSIS (TB) STAINING AND CULTURE (AFB CULTURE)**

Specimens Required

| | |
|-----------------------------|--|
| Sputum / Urine (EMU): | Three early morning specimens |
| Pus: | As much pus as possible |
| Other Body Fluids: | As available |
| Bone Marrow: | As much material as possible. Special bottle for collection of bone marrow available in the Microbiology department, please contact. |
| Bronchial Brushings (slide) | |

Please note 'Swab' samples are not useful for the demonstration or isolation of mycobacteria. Fluid or tissue samples should be sent.

Please note that whilst the demonstration of acid-fast bacilli in smears is diagnostic, positive samples may have few bacilli and are smear negative.

14.3.2 **BLOOD CULTURE**

Only take blood for culture when there is a clinical need to do so and not as routine. ([Click here](#) to access the policy entitled 'Indication for Taking Blood Cultures' for further information). Blood cultures should be taken after identification of possible sepsis and, before the administration of antibiotics (refer to policy D&TP0016, Integrated Care Pathway for the management of Sepsis Shock, in Q-Pulse). This should not prevent prompt administration of antimicrobial therapy.

If a patient is on antibiotics, blood cultures should be taken immediately before the next dose with the exception of paediatric patients. Drawing more than one set of cultures (at separate times from separate sites) can help to distinguish true bacteraemia from contaminated cultures (Shafazand and Weinaker, 2002). Samples should not be taken from existing peripheral cannulas but if the patient has an existing Central Venous Access Device a sample should be taken from this following collection of a separate peripheral sample.

Bottles

The blood culture bottles and system in use are the BacT/Alert (Biomérieux) system. Bottles should be kept at a cool room temperature in the wards. The number of bottles stored in each ward should be limited to their general usage and excessive stocks avoided.

Three bottles are available:-

- **green top (aerobic)**
- **orange top (anaerobic)**
- **yellow top (paediatric) bottle.**

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COLLECTION PROCEDURE:

Please refer to all relevant areas of Hospital Venepuncture Policy (HOS/65)

Blood cultures should be collected using the Blood Culture Adapter sets for peripheral and line cultures available on the ward as per Bon Secours Hospital, Cork, Collection of Blood Culture Specimens Policy (Q Pulse Ref NUR0178). Skin should be disinfected with a 3mL Chloraprep applicator swab prior to collection of peripheral blood. Blood Culture bottles are filled from peripheral sites via a safety multi-fly needle, the adapter cap and associated BCF Adapter (14.1110). Blood is collected from vascular access devices via Sarstedt multi-adapter (ref 12.1205.100). Please reference Procedure as per blood sampling in Hospital Policy Management and Care of Central Venous and Arterial Catheters (Ref No: NUR0122). **Add up to 10 ml of blood per adult bottle and up to 4 ml per pediatric bottle.**

If other blood tests are required, always collect the blood culture first. The BC adapter cap with attached BCF Adapter is disconnected from the butterfly and discarded to a sharps container and the additional blood bottles filled in order as per the Lab Manual.

Ensure additional labels do not cover the bottle bar code. Do not remove any part of the barcode from the bottles as these are used in the laboratory during processing.

Record the procedure, including indication for culture, time, site of venipuncture, and any complications. Transport specimens to the laboratory immediately in the bag attached to the Microbiology request form **using the pneumatic chute system at all times**. Fill out clinical details, antibiotic, time & site of venepuncture on the Microbiology request form.

Time to Report

Most organisms will be detected within 24-48 hrs and normally blood cultures are incubated for up to 5 days but this time may be extended to 10 days in some cases e.g. infective endocarditis. Please contact the Microbiology Department to discuss such cases.

POSITIVE CULTURES ARE NOTIFIED TO THE WARD INVOLVED IMMEDIATELY ON DETECTION BY THE MICROBIOLOGY/ ON CALL STAFF THEREFORE THERE IS NO NEED FOR WARD STAFF TO CONTACT THE MICROBIOLOGY LABORATORY TO DETERMINE IF A BLOOD CULTURE IS POSITIVE.

14.3.3 CEREBROSPINAL FLUID (C.S.F.) – INVESTIGATION FOR INFECTION

Samples should be transported to the lab as soon as possible.

Bacteraemia is sometimes seen associated with meningitis, and a blood culture should be taken when meningitis is suspected.

If in doubt, the Consultant Microbiologist should be contacted for advice.

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Specimen Required

CSF sample - as much sample as possible divided as per the CSF Request Form PL012:-

- Send a blood glucose sample (to compare with CSF glucose value)
- Send blood culture as outlined in section 14.3.2
- Send EDTA blood sample for PCR for meningococcus if this is suspected.
- Send a throat swab for meningococcus if this is suspected.

Ordering the correct test and providing the correct volume of sample will ensure that all tests are completed.

Normal Findings

| Cell Type | | | |
|--------------------|--|----------------|--------------------|
| WBC (Leucocytes) | Neonates | <28 days | 0-30 cells per cmm |
| | Infants | 1 to 12 months | 0-15 cells per cmm |
| | Children/Adults | >1 year | 0-5 cells per cmm |
| RBC (Erythrocytes) | No RBC's should be present in normal CSF | | |

Protein: 0.15 - 0.45 G/L

Viral Meningitis

If clinical and laboratory findings suggest viral meningitis, a CSF PCR Panel should be requested on the CSF. A faeces and throat swab sample may also be sent for Enterovirus. Additional viruses such as EBV may also be excluded by referral to the National Virus Reference Laboratory. Depending on clinical suspicion, consider sending a serum sample for HSV, VZV, Mumps, Measles, EBV and Enterovirus IgM. Results are phoned to the ward as soon as they are available.

14.3.4 FLUIDS FROM SITES NORMALLY STERILE

Specimens should be transported and processed as soon as possible.

The volume of specimen influences the transport time that is acceptable. Large volumes of purulent material maintain the viability of anaerobes for longer.

| Volume of aspirated material | Optimal time for transport to laboratory |
|------------------------------|--|
| <1ml | <10min |
| 1ml | <30min |
| >2ml | <3h |

The recovery of anaerobes is compromised if the transport time exceeds 3 hours.

If processing is delayed, refrigeration is preferable to storage at ambient temperature. Delays of over 48 hours are undesirable.

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Samples Required

A sample for **culture** in a sterile universal container. In addition to the sterile universal container, an EDTA (red) sample for **white cell count** may be of diagnostic value.

Normal Findings:

| | |
|----------------------------|--------------------------------|
| Peritoneal / Ascitic Fluid | White Cell Count: 0 - 200 /cmm |
| Synovial / Joint Fluid | White Cell Count: 0 - 200 /cmm |

14.3.5 MYCOLOGY (FUNGAL) EXAMINATION AND CULTURE

Systemic Fungal Infections

The isolation of fungi from blood and deep tissue is difficult. If systemic fungal disease is suspected please contact the Laboratory or the Consultant Microbiologist to discuss case and ensure that fungal culture is specifically requested on the request form.

Superficial/ Skin Fungal Infections

Specimens Required: Skin, Hair, Nails, Tissue

Taking of Samples

1. Preliminary cleansing of the lesion with 70% alcohol reduces bacterial contamination.
2. Scales of skin are scraped with a scalpel or the side of a microscope slide from the active periphery of the inflamed area. Infected nails should be clipped off for examination and scrapings taken from deeper areas of the nail bed. Infected hairs must be carefully chosen to avoid submitting healthy hair.
3. All samples should be sent to the laboratory in a sterile universal container or "Dermapak" available from the Microbiology department.
4. A 5 day fungal culture will be performed on swabs and fluid specimens when requested or where clinically indicated.

14.3.6 SPUTUM CULTURE/ BRONCHO-ALVEOLAR LAVAGE/ BRONCHIAL WASHINGS/ ANTRAL WASHOUT

Sample Required

A good quality sputum sample should be submitted.

Salivary or muco-salivary samples may give misleading results as these samples will be contaminated by normal mouth flora.

Sputum and Antral washout are cultured for all likely Lower Respiratory Tract (LRT) pathogens.

Broncho-Alveolar Lavage (BAL's) are routinely cultured for bacterial pathogens as well as Mycobacteria and fungi and are referred if indicated for examination for CMV, other viruses and Pneumocystis carinii.

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14.3.7 EYE SWABS

Samples Required

1. For investigation of **keratitis** a corneal scrape is required. Please notify laboratory prior to sampling to ensure prompt processing.
2. For investigation of **conjunctivitis** a swab should be taken as follows:-
Retract the lower eye lid and stroke the tarsal conjunctiva with a transport swab and remove all purulent material. Place swab in transport medium case. If chlamydial infection is suspected, please contact the laboratory beforehand to obtain chlamydial transport swab. Break the swab into this medium and replace cap. Send swabs to the laboratory immediately.
3. For investigation of endophthalmitis an aspirate from the aqueous humour is required. Please notify the laboratory prior to sampling to ensure prompt processing.

14.3.8 THROAT SWAB

Sample required

Using a tongue depressor take a vigorous swab sample from the tonsil or inflamed area. Replace the swab in the Amies transport medium charcoal.

14.3.9 NASAL SWAB

A nasal swab is not usually useful for the investigation of sinusitis.

Antral lavage or pus from sinus should be sent if acute maxillary sinusitis is suspected. Nasal swabs are useful for the investigation of carriage of Staphylococcus species, including MRSA.

Sample required

Rotate one swab twice round each of the anterior nares.

14.3.10 GENITAL INFECTIONS

Sexually Transmitted Diseases

Samples Required

Females: Cervical or High Vaginal Swabs, Urethral Swabs, Urine (Chlamydia, gonorrhoea and trichomonas)

Males: Urethral Swab and Urethral Smear, Urine (Chlamydia, gonorrhoea and trichomonas)

Genital Tract Swabs

Cervical and high vaginal swabs should be taken with the aid of a speculum. It is important to avoid vulval contamination of the swab. For *trichomonas*, the posterior fornix, including any obvious candidal plaques should be swabbed. If pelvic infection, including gonorrhoea, is suspected, the cervical os should be swabbed. These samples should be collected using the aptima collection device.

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High Vaginal Swabs

After the introduction of the speculum, the swab should be rolled firmly over the surface of the vaginal vault. The swab should then be placed in Amies transport medium with charcoal.

Cervical Swabs

After introduction of the speculum into the vagina, the swab should be rotated inside the endocervix. The swab should then be placed in Amies transport medium with charcoal.

Urethral Swabs

Contamination with micro-organisms from the vulva or the foreskin should be avoided. Thin swabs are available for collection of specimens.

The patient should not have passed urine for at least 1 hour. For males, the swab is gently passed through the urethral meatus and rotated. Place the swab in Amies transport medium with charcoal.

Intrauterine Contraceptive Devices (IUCDs)

The entire device should be sent in a sterile universal container.

Rectal Swabs

Rectal swabs should be taken via a proctoscope.

Transport all swabs immediately to the laboratory.

If Chlamydial infection is suspected, please contact the laboratory beforehand for Chlamydial Transport Swabs.

If Herpes simplex infection is suspected, please contact the laboratory beforehand for Viral Transport medium.

Urine Samples

Sterile universal container

14.3.11 PUS SAMPLES/ WOUND SWABS

Wound swabs should only be taken when signs of clinical infection are present.

Deep swabs rather than superficial will give more accurate representation of bacteria/ fungi present which may be causing infection. If viral infection is suspected please refer to section 14.3.16.

Please indicate clearly on the request form and the swab the site of the wound otherwise interpretation of culture results may be difficult.

Specimens Required

1. Pus sample (always preferable to a wound or pus swab) in sterile universal container.
2. Wound swab in transport swab. (Please note on form and the swab, the site of the wound and clinical details).

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Wound or Pus samples are screened for all likely bacterial pathogens and, if present, these organisms and their antibiotic sensitivity results are reported. The inclusion of relevant clinical information on the request form assists in deciding the relevance of some bacterial isolates.

14.3.12 MRSA SCREENS

Staff and/or patient screens for colonisation by Meticillin-Resistant Staph aureus (MRSA) are necessary from time to time in the control of infection with this organism. The Consultant Microbiologist and the infection control Clinical Nurse Manager will initiate and monitor this screening as deemed necessary.

A full MRSA screen consists of:-

Nasal Swab: Rotate **one** swab twice round each of the anterior nares.

Throat Swab: Using a tongue depressor take a vigorous swab sample from the tonsil or back of pharynx.

Perineal Swab: Use one swab to sample.

Axillary Swab: Use one swab for each axilla.

Groin Swab: Use one swab for each groin.

If present, also send swab if wounds, sites of damaged or abnormal skin, intravenous line insertion sites, catheter urine samples, and sputum if expectorating.

14.3.13 URINE

A clean mid-stream specimen is essential. In urinary tract infection (UTI) the bacterial count exceeds 100,000 organisms/ml in the majority of cases. Two samples should ideally be taken to make a diagnosis.

Urine acts as a culture medium and therefore specimens should be stored at 4°C to prevent subsequent multiplication of bacteria after collection of the patient sample which would invalidate the bacterial count. **Any sample which may be subject to delay of more than 2 hrs before being sent to the lab should be refrigerated.**

Samples Required

MSU A midstream urine is the recommended sample and requires careful collection.

CSU Samples may be from patients who have had a catheter passed for a one-off urine sample or who have in-dwelling catheters. In patients with a long term indwelling catheter samples should only be sent if clinically indicated i.e. patient symptomatic or systemically unwell or having a catheter change or urinary tract instrumentation.

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BSU Bag specimen of urine. A sterile collection bag is applied to the cleansed perineum to catch urine, which must then be drained into a sterile universal container. This is commonly used in infants. Culture results are difficult to interpret as contamination is common with this method of specimen collection.

Tests Available

1. Microscopy White cell count:
Normal range 0-10 / c.mm
Red cell count (normally absent)
Comment on the presence of organisms, casts, crystals, etc.
2. Bacterial Colony Count:
should be less than 1,000 orgs/ml ($<10^3$ orgs/ml)
1,000 - 100,000 orgs/ml may indicate UTI
>100,000 orgs/ml is usually indicative of UTI

14.3.14 CPE/ ESBL/VRE SCREENS

The Consultant Microbiologist and the infection prevention and control clinical nurse manager will recommend and monitor patient and environmental screening as deemed necessary. CPE/ ESBL/ VRE screening is ideally carried out on a rectal swab or occasionally if this is not possible on a faeces sample. All 3 tests may be ordered on one rectal swab.

14.3.15 OVA CYSTS AND PARASITES

Please include as much clinical information as possible, particularly any history of foreign travel.

Sellotape Slides

Diagnosis of “Pinworm” (*Enterobius vermicularis*) infestation in children may also be made using the sellotape slide technique. Apply a piece of sellotape to the anal margin at night (the female worm crawls out of the anus at night and lays eggs in the anal margin). Then remove the sellotape and attach it to a glass slide which is sent to the laboratory. Please label the slides with the patient's name and hospital number.

14.3.16 ANTIBIOTIC ASSAY

Refer to the Antimicrobial Guidelines in the Public Documents Folder on the desktop under [Antimicrobial Guidelines](#).

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14.3.17 SEROLOGY\VIROLOGY GENERAL COMMENTS AND REQUESTING GUIDE

Please discuss with the Consultant Microbiologist if tropical or obscure infection is suspected. If viral infection is suspected in immunocompromised patients, please contact the Consultant Microbiologist for advice.

Identification of patients with invasive fungal infection

The diagnosis of invasive fungal infection still poses many challenges and there is no single diagnostic test that is both sensitive and specific. In the majority of cases the use of other data e.g. clinical predisposition and findings, radiological investigations remains an important component of diagnostic strategy and the tests listed below may be used as a adjunct to appropriate clinical and radiological evaluation.

Routine microbiological tests:

Use of routine microscopy and culture of appropriate samples e.g. blood, BAL, tissue remain an important part of the diagnostic workup.

Histological examination

Histological examination of tissue should be considered in certain cases e.g. invasive aspergillosis of the lung, sinuses etc,

(1→3)-β-D-Glucan test (BG)

(1→3)-β-D-Glucan is a structural component of the cell wall of various medically important fungi. Testing for this protein detects the presence of fungal infection some days before clinical signs or symptoms and before other biomarkers become positive. It only indicates the presence of fungus. It cannot specify yeast or mould or the infecting species. Infections detected by this test include Candidiasis (except *C. parapsilosis*), fusariosis, trichoporosis and aspergillosis. It also has the potential to detect invasive fungal infections due to *Pneumocystis jiroveci*.

False positive results can occur due to contamination of the specimen with fungal spores or contact with items like cellulose filters, gauze and cotton swabs or in patients who have had recent surgery, have dressings, are on dialysis, are in receipt of blood products or are receiving IV amoxicillin/clavulanic acid

Fungal species including *Cryptococcal* species, *Absidia corymbifera*, *Rhizopus*, *Rhizomucor* and *Mucor* species have very little (1→3)-β-D-Glucan and so may give false negative results.

A minimum of 0.5ml of reffridgerated serum is required.

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Aspergillus Antigen/Galactomannan (GM)

Galactomannan (GM) is a cell wall polysaccharide which is released by *Aspergillus* species during hyphal growth in tissues and is detectable in blood at a median of 5-8 days (range, 1-27 days) before the clinical manifestation of Aspergillosis or the demonstration of abnormalities in high-resolution CT scan. The concentration of circulating GM corresponds with the fungal tissue burden and so may be used to monitor the patient's response to antifungal treatment.

Detection of Aspergillus antigen as a screening test is unlikely to be beneficial or cost-effective if the pre-test probability of the test is low. Testing should be reserved for high-risk populations including allogenic stem cell transplant patients, acute myeloid leukaemia patients and patients undergoing aggressive chemotherapeutic regimens for relapsed disease.

Serial testing is required (twice weekly) to achieve acceptable sensitivity.

False positives may occur, especially in patients treated with certain β lactam antibiotics (piperillin/tazobactam, amoxicillin/clavulanic acid, ampicillin and phenoxymethylpenicillin). This false positive result can persist for over 10 days post administration of the β lactam antibiotics. Patients undergoing dialysis and patients undergoing liver transplantation for autoimmune liver disease or patients with chronic graft-versus-host disease after allogenic bone marrow transplant may also have false positive results.

False negative results can occur if there is previous exposure to antifungal drugs. A false negative result may also occur in patients with non or minimally invasive manifestation of aspergillosis (Aspergilloma or tracheobronchitis). GM detection is not useful for patients suffering from chronic cavity pulmonary aspergillosis or allergic bronchopulmonary aspergillosis.

This test has poor sensitivity, good specificity and an excellent negative predictive value in the detection of invasive aspergillosis in high-risk patients.

Patient Testing Strategy

In addition to other routinely used diagnostic tests in patients with relevant risk factors and suspected invasive aspergillosis perform a GM test twice weekly, confirm a positive with a follow up sample and a request for BG also.

In addition to other routinely used diagnostic tests in patients with relevant risk factors and suspected invasive candidiasis perform a BG test twice weekly and C. albicans antigen test to determine if the cause is C. albicans or non-albicans.

In addition to other routinely used diagnostic tests in patients with relevant risk factors and suspected fungal infection other than the above perform a BG test twice weekly.

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Atypical pneumoniae and Respiratory Infections

1. **Chlamydia pneumoniae**

ELISA tests for IgM antibodies that appear within 2 to 4 weeks of onset of illness. Restricted test: will only be referred following approval from Consultant Microbiologist.

2. **Legionella pneumophila**

In an acute setting, please send urine for urinary antigen. This test is useful in the early detection of Legionnaires disease.

Positive Legionella screening tests are confirmed at the Atypical Pneumoniae Unit, Respiratory and Systemic Infection Laboratory, CPHL, 61 Colindale, London NW9 5HT. This results in an additional turnaround time of 2 weeks.

3. **Mycoplasma pneumoniae**

This is an EIA test for the detection of IgM antibodies. IgM antibodies usually appear 10 days after onset of illness.

4. **Coxiella burnetii**

ELISA test for IgM, IgG and IgA antibodies that appear within 2 to 4 weeks of onset of illness.

5. **Influenzae A and B and Swine Flu**

Serology not appropriate. During off season periods, a nasopharyngeal aspirate (NPA) is required. In season, please send a nasal and throat viral swab or a nasopharyngeal aspirate.

SEROLOGY: General Notes

Serological Tests

For a serological diagnosis, i.e. antibody tests based on appearance of IgG, acute (as early as possible in the illness) and convalescent sera (2-3 weeks after on-set) should be taken for antibody titration. A four-fold rise in titre is considered significant. Single samples for serology are of limited value unless used for detection of IgM antibody when such tests are available.

VIROLOGY: General notes

Virus Isolation

Where virus isolation is attempted, specimens must be taken early in the illness in the correct manner (contact Microbiology department).





Please Note: Samples marked 'virus studies' or 'viral screen' will not be processed. Please specify requests and give clinical details. Failure to supply the required information will lead to delays in reporting.

The patient's date of birth must be supplied in all cases.







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Requesting Guide







The following “Requesting Guide” may assist in identifying possible agents with a certain clinical syndromes. This guide is derived from guidelines issued by the National Virus Reference Laboratory, Dublin.

| | TESTS TO BE ORDERED | |
|--|--|---|
| Provisional Diagnosis/ Symptoms | Possible Virus/ Agent/ Disease | Specimen Type |
| Respiratory Infection URTI and LRTI (Atypical pneumonia) (Refer to section 14.3.16) | Mycoplasma pneumoniae Ab IgM Chlamydia pneumoniae Ab IgM | Serum/ Clotted Blood  |
| | Legionella pneumophila (Urinary Antigen) | Urine |
| | Coxiella burnetii (Q Fever) Ab IgM Please request if clinically suspected. Not part of routine atypical pneumonia screen. | Serum/ Clotted Blood  |
| Respiratory Infection URTI and LRTI (Viral) (Refer to section 14.3.16) | Influenza A Influenza B | Nose and throat swab in viral transport media |
| | Respiratory Syncytial Virus (RSV) Human Metapneumovirus Seasonal testing for Parainfluenza and Adenovirus | Nasopharyngeal Aspirate (NPA) or Bronchial Lavage |
| | Cytomegalovirus (CMV) in immuno compromised patients | Bronchial Lavage |
| Arthralgia | Parvovirus B19 Mycoplasma pneumoniae Borrelia burgdorferi (Lyme Disease) | Serum/ Clotted Blood  |
| | Consider Brucella. Please request if clinically suspected. Not part of routine arthralgia screen. | |
| Exanthem (Skin Rash) | Measles IgM Rubella IgM Parovirus B19 Borrelia burgdorferi (Lyme Disease) (if clinically suspected) HIV (if risk factors) Hepatitis B (if risk factors) Dengue (if risk factors) | Serum/ Clotted Blood  Oracol Swab (Saliva Oral Fluid for Measles only) |






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| TESTS TO BE ORDERED | | |
|--|---|--|
| Provisional Diagnosis/ Symptoms | Possible Virus/ Agent/ Disease | Specimen Type |
| Exanthem (Skin Rash) | Herpes Simplex Virus (HSV) Varicella Zoster Virus (VZV) | Viral swab Vesicle Fluid/ |
| | Molloscum Contagiosum Orf Virus | Scrapings on Slide Vesicle Fluid/ Scrapings on Slide |
| | Enterovirus | Stool/ Viral Throat Swab/ Vesicle Fluid/ Scrapings on Slide |
| Central Nervous System | Mumps virus | Serum/ Clotted Blood or  salivary swab or CSF |
| | Measles IgM | Oracol Swab (Salivary Swab) |
| | Herpes simplex virus Varicella zoster virus | Viral UTM Copan Swab Respiratory Secretions or CSF |
| | Enterovirus (Coxsackie Echo) | CSF/ Stool/ Viral Throat Swab/ Pleural Fluid |
| | Dengue (if risk factors) West Nile Virus (if risk factors) | Serum/ Clotted Blood  |
| Hepatitis Screen | Hepatitis A, B, C, Cytomegalovirus (CMV IgM) Epstein-Barr Virus (EBV IgM) Leptospirosis antibodies | Serum/ Clotted Blood  Plasma  |
| CABG/HEP screen | Hepatitis B & C | Serum/ Clotted Blood  |
| Genital Infection | Herpes Simplex Virus | Viral Swab |
| | Chlamydia trachomatis/ Gonorrhoea/ Trichomonas | Aptima collection device |
| | Syphilis (Treponema pallidum) | Serum/ Clotted Blood  |
| Diarrhoea/Vomiting | Rotavirus Adenovirus Astrovirus Calicivirus Norovirus (small round structured virus) | Stool |

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| TESTS TO BE ORDERED | | |
|--|---|---|
| Provisional Diagnosis/ Symptoms | Possible Virus/ Agent/ Disease | Specimen Type |
| Intra-Uterine Infection | Toxoplasma IgM Cytomegalovirus (CMV IgM) Rubella IgM Parvovirus B19 Varicella Zoster Virus (VZV) | Serum/ Clotted Blood  Viral UTM Copan Swab Respiratory Secretions |
| Inflammatory Bowel Disease Profile | Quantiferon, HIV Screening Test Hepatitis B Surface Antigen Hepatitis B Core Antibodies Hepatitis B Surface Antibodies Hepatitis C Antibodies TPMT Autoantibody Screen Varicella Zoster IgG | Serum/ Clotted Blood  Blood Tube (4.9mL) containing no additive x 3 Blood Tube (7.5mL) containing EDTA Quantiferon Blood Collection Tube |
| Biologics Profile | Quantiferon Hepatitis B surface antigen HIV Screening Test Hepatitis C Antibodies Hepatitis B Core Antibodies Varicella Zoster Virus IgG Hepatitis B Surface Antibodies | Serum/ Clotted Blood  Blood Tube (4.9mL) containing no additive x 3 Blood Tube (7.5mL) containing EDTA Quantiferon Blood Collection Tube |
| Lymphoma Viral Screen | Hepatitis B surface antigen HIV Screening Test Hepatitis C Antibodies Anti Hepatitis B Core Varicella Zoster IgG | Serum/ Clotted Blood  |
| Organ Donor | Syphilis RPR/ TPPA Cytomegalovirus (CMV IgM) Toxoplasma Total Hepatitis B Surface Antigen HIV Hepatitis C Antibodies HTLV | Serum/ Clotted Blood  |
| Ante Natal | Rubella IgG Hepatitis B surface Antigen HIV (Must be requested) Syphilis RPR/ TPPA | Serum/ Clotted Blood  |
| Pleurodynia | Coxsackie Group B viruses | Stool/ Viral Throat Swab or Pleural Fluid |

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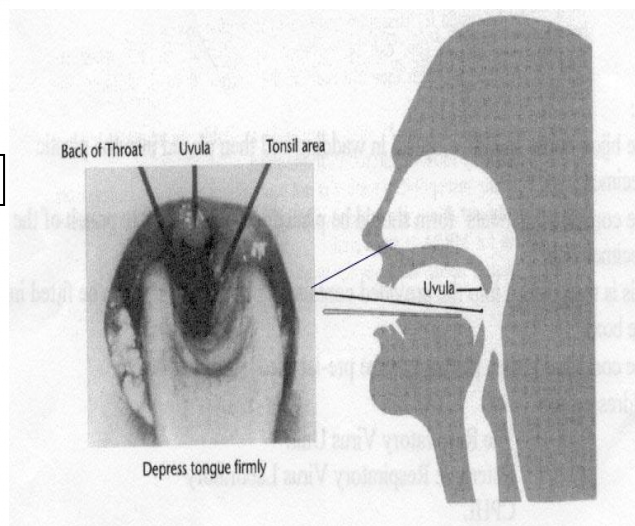
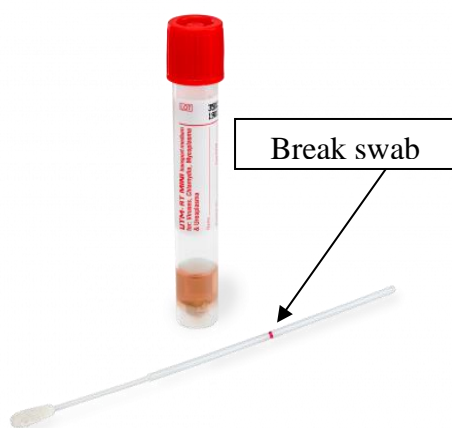
| | TESTS TO BE ORDERED | |
|--|--|--|
| Provisional Diagnosis/ Symptoms | Possible Virus/ Agent/ Disease | Specimen Type |
| Pericarditis/ Myocarditis | Coxsackie Group B viruses | Stool/ Viral Throat Swab or Pleural Fluid |
| | Coxiella burnetii (Q Fever) Chlamydia Group Mycoplasma pneumoniae | Serum/ Clotted Blood  |
| Lymphadenopathy & Glandular Fever | Epstein Barr Virus (EBV IgM) Cytomegalovirus (CMV IgM) Toxoplasma IgM Bartonella Order HIV separately if requested | Serum/ Clotted Blood  |
| Paraparesis | Human T-Lymphotropic Virus (HTLV) | Serum/ Clotted Blood  |
| | Enterovirus (Coxsackie Echo) | Stool |
| Conjunctivitis | Adenovirus Herpes Simplex Virus (HSV) Enterovirus (Coxsackie Echo) | Viral Eye Swabs |
| | Chlamydia trachomatis | Swab in Chlamydia transport media |
| Stomatitis | Herpes Simplex Virus (HSV) | Swab in viral transport media Serum/ Clotted Blood  |
| | Enterovirus (Coxsackie Echo) | Stool/ Throat Swab |
| Hand, Foot and Mouth Disease | Coxsackie A16 Virus | Stool |
| Immune Status | Rubella IgG Mumps IgG Measles IgG Parvovirus IgG Hepatitis B Surface Antibodies Hepatitis A IgG Antibodies Hepatitis C | Serum/ Clotted Blood  |
| Haemorrhagic cystitis | Adenovirus Polyoma | |

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14.3.18 COVID 19 TESTING

The procedure for collecting nasal and throat swabs for investigation of respiratory viruses including COVID-19.

- Explain procedure to the patient
- Collect **both nasal and throat swab** using the one swab
- Insert swab into patients open mouth
- The swab is used first to abrade the tonsils and pharynx (see diagram below on right). This will induce a gag reflex.
- This swab is then inserted into the nostril and rubbed against and above the nasal turbinate (swab to be advanced almost 4 inches into the nostril. This will bring a tear to the eye)
- Repeat for the other nostril.
- **Place the swab into the red container** containing the transport medium.
- Break off the swab at the moulded breakpoint on the swab shaft (see picture below on the left)
- **Close lid securely.** Ensure that the bottle lid is secured tightly onto the bottle to prevent leakages.
- **Label specimen** for COVID19 PCR and write on clinical details if patient is symptomatic.
- Can be sent in POD system (clean POD inside and outside with 70% Alcohol wipe before sending)



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Diagnosis of Acute Infection (PCR):

PCR is the ‘gold standard’ recommended for the diagnosis of COVID-19 during the acute phase of infection.

False negative results can occur if testing takes place in the initial incubation period following infection. The minimum duration from infection to a positive test remains uncertain. SARS-CoV-2 viral RNA can be detected one-to-two days prior to symptom onset in upper respiratory tract samples (Figure 1). Viral load peaks around the time of symptom onset, the level of virus in nasopharyngeal secretions declines progressively over time after onset of symptoms becoming undetectable approximately two weeks following symptom onset.

The diagnostic window for using PCR to detect acute infection with SARS-CoV-2 therefore ranges from approximately three days following exposure to the virus until two weeks following symptom onset (Figure 1).

Testing in BSHC is performed using one of the following assays:

- GeneXpert
- BioFire FilmArray

Interpretation of PCR Results:

When SARS-CoV-2 is **DETECTED**, this indicates that virus is present.

Note: Viral RNA may be detected for a period when replicating virus is no longer present. Detection of virus RNA does not necessarily mean that the person is infectious.

When SARS-CoV-2 is **NOT DETECTED**, in a well taken sample during the symptomatic period this makes it much less likely that the person has COVID-19. However, in some people with symptomatic infection virus has been undetectable in nasopharyngeal samples swabs.

Note: A negative PCR result is only an indication of the patient’s status at a point in time. If self-isolation is required from a public health perspective, a negative result may not negate the need for self-isolation.

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14.4 MICROBIOLOGY PROFILES

| INFLAMMATORY BOWEL DISEASE (IBD) PROFILE |
|---|
| Quantiferon, HIV, Hepatitis B Surface Antigen, Hepatitis B Core Antibodies, Hepatitis B Surface Antibodies, Hepatitis C Antibodies, TPMT, Autoantibody Screen, Varicella Zoster IgG |
| Sample Requirements |
| Blood Tube (4.9mL) containing no additive x 3 Blood Tube (7.5mL) containing EDTA Quantiferon Blood Collection Tube |
| Turn Around Time |
| Refer to individual tests for turnaround time in this document |

| BIOLOGICS PROFILE |
|--|
| Quantiferon, HIV, Hepatitis B Surface Antigen, Hepatitis B Core Antibodies, Hepatitis B Surface Antibodies, Hepatitis C Antibodies, Varicella Zoster IgG |
| Sample Requirements |
| Blood Tube (4.9mL) containing no additive x 3 Blood Tube (7.5mL) containing EDTA Quantiferon Blood Collection Tube |
| Turn Around Time |
| Refer to individual tests for turnaround time in this document |

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15.0 CHROMOSOME ANALYSIS/ DNA GENETIC SCREENING

15.1 Chromosome Analysis and Genetic Testing is a very specialised Laboratory technique presently not performed by the Bon Secours Pathology Department. However, we do refer specific requests to other specialised Laboratories. These tests are specific to a particular syndrome or abnormality. **It is essential that the sample request form clearly identifies the test(s) required by the Clinician** as it is only these requests that will be investigated. The relevant clinical history must be provided as it is of benefit in the performance of these tests. The list below includes the majority of the more commonly requested tests, but it is impractical to list all Chromosome Analysis / Genetic tests that may be requested. If a particular test is required, but does not appear on the following list, please contact the Chief Medical Scientist, Histopathology Department for further details.

Consent is required for the listed genetic tests, refer to the links in special requirements to access the referral Laboratory consent forms where available. Please print Genetic Test Request Forms and complete including signatures of the patient and requesting Doctor.

Please note it is critical that patient details on the request form and specimen match, including spelling. Any discrepancies will result in the sample not being tested by the referral Laboratory.





BIOCHEMISTRY - Chromosome Analysis/ DNA Genetic Screening

| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|-------------------------------|---------------|---|---------------------------|-----------------------|---|-----------------|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| DNA GENETIC SCREENING | | | | | | |
| Alpha-1 Antitrypsin Genotype* | Blood | Potassium EDTA  | 7.5 | Blood Tube | Bring to Lab immediately. Preferably Monday to Wednesday am. Consent required, click on link for form. Click here to access Eurofins Biomnis Consent Form and send with the sample. Whole blood, do not centrifuge. | 30 working days |
| | | None  | 4.9 | Blood Tube | | |
| APO E Lipoprotein Genotyping* | Blood | Potassium EDTA  | 7.5 | Blood Tube | Preferably Monday to Thursday am. Consent required, click on link for form. Click here to access Eurofins Biomnis Consent Form and send with the sample. Whole blood, do not centrifuge. | 20 working days |

* These specimens/ samples are referred to external laboratories for testing.

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BIOCHEMISTRY - Chromosome Analysis/ DNA Genetic Screening.....cont'd

| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|--|---------------|---|---------------------------|--------------------------|---|-----------------|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| DNA GENETIC SCREENING | | | | | | |
| Cystic Fibrosis DNA Studies* | Blood | Potassium EDTA  | 7.5 (Adult) 4 (Paed) | Blood Tube | Preferably Monday to Thursday am. Consent required, click on link for form. Click here to access Eurofins Biomnis Consent Form and send with the sample. Whole blood, do not centrifuge. | 25 working days |
| Dihydropyrimidine dehydrogenase Gene Screen (DPD)* | Blood | Potassium EDTA  | 2.7 | Blood Tube (Whole Blood) | Whole blood. | 20 working days |
| FABRY Disease Screen* (Alpha Galactosidase plus Genetic Screen as indicated) | Blood | Potassium EDTA  | 2.7 | Blood Tube | Preferably Monday to Wednesday am. Consent required, click here to access Centrogene Consent Form and send with the sample. Whole blood, do not centrifuge. | 30 working days |
| Familial Hypercholesterol-aemia Genetics* | Blood | Potassium EDTA  | 7.5 | Blood Tube | Consent required, click here to access form. | 25 working days |

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BIOCHEMISTRY - Chromosome Analysis/ DNA Genetic Screening.....cont'd





| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|--|---------------|---|---------------------------|-----------------------|---|-----------------|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| DNA GENETIC SCREENING | | | | | | |
| GCH1* | Blood | Potassium EDTA  | 2 | Blood Tube | Preferably Monday to Wednesday am. Click here to access Eurofins Biomnis Consent Form and send with the sample. Whole blood, do not centrifuge. | 30 working days |
| Haemochromatosis DNA* | Blood | Potassium EDTA  | 7.5 | Blood Tube | Preferably Monday to Thursday am. Consent required, click on link for form. Click here to access Eurofins Biomnis Consent Form and send with the sample. Whole blood, do not centrifuge. | 25 working days |
| Hereditary Transthyretin Mediated Amyloidosis Gene (h ATTR) (Screening Test for Cardiac Amyloidosis)* | Blood | Potassium EDTA  | 2 x 2.7 | Blood Tube | Preferably Monday to Wednesday am. Consent required, click here to access request/ consent form. Whole blood, do not centrifuge. | 42 working days |
| Maternally Inherited Diabetes and Deafness Genetic Screen* | Blood | Potassium EDTA  | 2 x 7.5 | Blood Tube | Preferably Monday to Wednesday am. Consent by patient required and specific request form fully completed by requesting clinician. Click on link for form. Click here to access MIDD Genetics Request Form and send with the sample. Whole blood, do not centrifuge. | 50 working days |

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BIOCHEMISTRY - Chromosome Analysis/ DNA Genetic Screening.....cont'd







| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|---|---------------|---|---------------------------|-----------------------|--|-----------------|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| DNA GENETIC SCREENING | | | | | | |
| MEN Type 2 Genetics* | Blood | Potassium EDTA  | 7.5 | Blood Tube | Preferably Monday to Wednesday am. Consent required, click on link for form. Click here to access TDL Consent Form and send with the sample. Whole blood, do not centrifuge. | 40 working days |
| MODY Genetics* | Blood | Potassium EDTA  | 2 x 7.5 | Blood Tubes | Preferably Monday to Wednesday am. Consent by patient required and specific request form fully completed by requesting clinician. Click on link for form. Click here to access MODY Genetics Request Form and send with the sample. Whole blood, do not centrifuge. | 50 working days |
| PNPO* Pyrimidine 5' Phosphate Oxidase | Blood | Potassium EDTA  | 2 | Blood Tube | Preferably Monday to Wednesday am. Click here to access Eurofins Biomnis Consent Form and send with the sample. Whole blood, do not centrifuge. | 45 working days |
| UGT1A1 Genetic Test* (Alternate name Gilberts Syndrome) | Blood | Potassium EDTA  | 2 x 7.5 | Blood Tube | Whole blood. Consent required, click on link for form. Click here to access Eurofins Biomnis Consent Form and send with the sample. Complete patient's history and family history. Please also confirm if the mutation has been described in the patient's family, if so please confirm which relative. Whole blood, do not centrifuge. | 25 working days |

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IMMUNOLOGY - Chromosome Analysis/ DNA Genetic Screening





| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|----------------------------|---------------|---|---------------------------|--|---|------------------|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| GENETICS | | | | | | |
| Angelman Syndrome* | Blood | Potassium EDTA  Lithium Heparin  | 7.5 4.9 | Blood Tube (Do not use tubes containing beads) | Monday OR Tuesday before midday. Consent required, click on link for form. Click here to access Eurofins Biomnis Consent Form B12-INTGB and send with the sample. Whole blood, do not centrifuge. | 25 working days. |
| Chromosome Analysis Blood* | Blood | Lithium Heparin  | 4.9 | Blood Tube | Monday OR Tuesday before midday. Consent required, click on link for form. Click here to access Eurofins Biomnis Consent Form B13-INTGB and send with the sample. Whole blood, do not centrifuge. | 25 working days |
| Di George Syndrome* | Blood | Potassium EDTA  Lithium Heparin  | 7.5 4.9 | Blood Tube | Monday OR Tuesday before midday. Consent required, click on link for form. Click here to access Eurofins Biomnis Consent Form B22-INTGB and send with the sample. Click here to access Eurofins Biomnis Consent Form B34-INTGB and send with the sample. Whole blood, do not centrifuge. | 40 working days. |
| Fragile X DNA Studies* | Blood | Potassium EDTA  | 7.5 | Blood Tube (Do not use tubes containing beads) | Monday OR Tuesday before midday. Consent required, click on link for form. Click here to access Eurofins Biomnis Consent Form B12-INTGB and send with the sample. Whole blood, do not centrifuge. | 25 working days |

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IMMUNOLOGY - Chromosome Analysis/ DNA Genetic Screening.....cont'd

| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|------------------------|---------------|---|---------------------------|---|--|-----------------|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| GENETICS | | | | | | |
| Huntington's Disease* | Blood | Potassium EDTA  | 7.5 | Blood Tube | Monday to Wednesday am. Consent required, click on link for form. Click here to access Eurofins Biomnis Consent Form B12-INTGB and send with the sample. Whole blood, do not centrifuge. | 40 working days |
| Myotonic Dystrophy* | Blood | Potassium EDTA  | 3 x 7.5 | Blood Tube | Monday to Wednesday am. Consent required, click on link for form. Click here to access Eurofins Biomnis Consent Form B12-INTGB and send with the sample. Whole blood, do not centrifuge. | 50 working days |
| Prader-Willi Syndrome* | Blood | Potassium EDTA  Lithium Heparin  | 7.5 4.9 | Blood Tube (Do not use tubes containin g beads) | Monday OR Tuesday before midday. Consent required, click on link for form. Click here to access Eurofins Biomnis Consent Form B12-INTGB and send with the sample. Whole blood, do not centrifuge. | 25 working days |

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HAEMATOLOGY- Chromosome Analysis/ Karyotyping/ Cytogenetics



| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|---|----------------------|--|---------------------------|-----------------------|--|-----------------|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| CHROMOSOME ANALYSIS/ KARYOTYPING/ CYTOGENETICS | | | | | | |
| AML Cytogenetic Panel* | Bone Marrow or Blood | Potassium EDTA  | 10 | Blood Tube | Check with Haematology Lab | 15 working days |
| BCR-ABL Fusion Gene* | Blood | Potassium EDTA  | 2.7 | Blood Tube | Samples should be received Mon to Thurs midday. | 10 working days |
| BCR-ABL Transcripts (Quantitation)* | Blood | Potassium EDTA  | 2.7 | Blood Tube | Samples should be received Mon to Thurs midday. | 10 working days |
| Bone Marrow FISH* | Bone Marrow | Lithium Heparin  | 4.9 | Blood Tube | Samples should be received Mon to Thurs midday. | 15 working days |
| CALR Mutation* | Bone Marrow or Blood | Potassium EDTA  | 2.7 | Blood Tube | Samples should be received Mon to Thurs midday. | 15 working days |
| Chromosome Analysis Bone Marrow (Cytogenetics Bone Marrow)* | Bone Marrow | Lithium Heparin  | 4.9 | Blood Tube | Samples should be received Mon to Thurs midday. | 15 working days |
| CLL Panel (TP53, 17pdel, IGHV)* | Bone Marrow or Blood | Potassium EDTA  | 2.7 | Blood Tube | Samples should be received Mon to Thurs midday. | 15 working days |
| Immunoglobulin Gene Mutation* (IgVH Mutation) | Bone Marrow or Blood | Potassium EDTA  | 2.7 | Blood Tube | Samples should be received Mon to Thurs midday. | 20 working days |
| JAK2* | Bone Marrow or Blood | Potassium EDTA  | 2.7 | Blood Tube | Samples should be received Mon to Thurs midday. | 10 working days |
| MDSNGS Panel* | Bone Marrow | Potassium EDTA  | 2.7 | Blood Tube | Samples should be received Mon to Thurs midday. Click here to access Eurofins Biomnis Consent Form and send with the sample | 20 working days |

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HAEMATOLOGY- Chromosome Analysis/ Karyotyping/ Cytogenetics.....cont'd

| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|--|----------------------|---|---------------------------|-----------------------|--|-----------------|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| CHROMOSOME ANALYSIS/ KARYOTYPING/ CYTOGENETICS | | | | | | |
| MPL Mutation Studies (MPLS)* | Bone Marrow or Blood | Potassium EDTA  | 2.7 | Blood Tube | Samples should be received Mon to Thurs midday | 15 working days |
| MPN Panel* (JAK2, CALR, MPN, BCR-ABL) | Blood/ Bone Marrow | Potassium EDTA  | 2.7 | Blood Tube | Samples should be received Mon to Thurs midday | 10 working days |

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HAEMATOLOGY – DNA Genetic Screening

| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|---|---------------|---|---------------------------|-----------------------|--|-----------------|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| DNA GENETIC SCREENING | | | | | | |
| Factor II Prothrombin Gene Mutation* | Blood | Potassium EDTA  | 7.5 | Blood Tube | Preferably Monday to Thursday am Consent required, click on link for form. Click here to access Eurofins Biomnis Consent Form and send with the sample. Whole blood, do not centrifuge. | 15 working days |
| Factor V Leiden DNA Studies* | Blood | Potassium EDTA  | 7.5 | Blood Tube | Preferably Monday to Thursday am. Consent required, click on link for form. Click here to access Eurofins Biomnis Consent Form and send with the sample. Whole blood, do not centrifuge. | 15 working days |
| MTHFR* (Methylene Tetrahydrofolate Reductase Deficiency)* | Blood | Potassium EDTA  | 2 x 7.5 | Blood Tube | Preferably Monday to Wednesday am. Consent required, click on link for form. Click here to access Eurofins Biomnis Consent Form and send with the sample. Whole blood, do not centrifuge. | 30 working days |

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16.0 POINT OF CARE/ NEAR PATIENT TESTING

The following Point of Care/ Near Patient Testing is performed and reported in designated clinical areas.

| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|---|--------------------------------|--------------------------|---------------------------|-----------------------|---|-----------------|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| ACT+ (Activated Clotted Time) ACT Low Range (LR) | Blood | N/A | N/A | N/A | Sample taken in approved areas . Refer to procedure BSC/POC/SOP/014. | <5 mins |
| Blood Gas CCU & MAU Venous/ Arterial | Blood | Heparinised | 1 | Blood Gas Syringe | For further information, contact POC/ NPT or Biochemistry dept. Refer to policy BSC/POC/SOP/018. Do not shake sample as haemolysis may cause an elevation in potassium concentration. | <15 mins |
| Clo Test for Helicobacter pylori (Endoscopy/ Theatre) | Tissue | Follow test kit protocol | | | Refer to policy ENDO 0017. | 30 mins |
| Creatinine (Capillary) (Radiology) | Blood capillary finger prick | Heparinised | 65µL | Capillary tube | Refer to procedure BSC/POC/SOP/017 | < 5 mins |
| Gastric pH (Childrens Ward) | Gastric Fluid | N/A | N/A | N/A | Refer to policy DIE0008 | <5 mins |
| Glucose (Capillary) (All Clinical Locations) | Blood Capillary (Finger Prick) | N/A | N/A | N/A | For further information, contact Diabetes Nurse Specialist. Refer to BSC/POC/SOP/020 and DIAB 003. | <5 mins |
| Oxyhaemoglobin (Cath Lab) | Blood | Heparinised | 50µl | Blood Gas Syringe | Sample taken in a controlled Cath Lab environment. Refer to procedure BSC/POC/SOP/016 | <5 mins |

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16.0 POINT OF CARE/ NEAR PATIENT TESTINGcont'd

| Test/Profile | Specimen Type | Specimen Requirements | | | Special Requirements | Turnaround Time |
|--|-----------------------------------|--------------------------|---------------------------|-----------------------------|---|-----------------|
| | | <u>Additive Required</u> | <u>Volume Required mL</u> | <u>Container Type</u> | | |
| Ketone (Capillary) (Approved wards) | Blood Capillary (Finger Prick) | N/A | N/A | N/A | For further information, contact Diabetes Nurse Specialist. Refer to policy NUR 0185. | <5 mins |
| Urinalysis (4 Panel) (All clinical locations) | Urine | N/A | 5-20 | Clean Container | Refer to policy NUR0052 | <5 mins |
| Urinalysis (10 Panel) (Approved wards) | Urine | N/A | 5-20 | Clean Container | Refer to policy BSC/POC/SOP/015 | <5 mins |
| Urinary hCG (Pregnancy Test) (Approved wards) | Urine | None | 5-20 | Sterile Universal Container | Refer to procedure BSC/POC/SOP/013 | <5 mins |

Note: Records of Point of Care/ Near Patient Test results are maintained in the patient's Medical Record.

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16.1 Point of Care/ Near Patient Testing Critical Test Result Values

| Parameter | Units of Measurement | Lower Limit | Higher Limit | Required Action by User |
|---------------------------|----------------------|----------------------|---|--|
| Blood Gas CCU - Potassium | mmol/L | ≤ 2.5 | ≥ 6.0 | Treat and Medical SHO contacted |
| Blood Gas CCU - Lactate | mmol/L | N/A | > 2.0 first occurrence > 4.0 all occurrences | Sepsis protocol initiated and Medical SHO Contacted |
| Blood Gas CCU - pH | pH | ≤ 7.3 | ≥ 7.6 | Contact NCHD/ SHO |
| Blood Gas CCU - Glucose | mmol/L | ≤ 3.0 | ≥ 20 | Contact NCHD/ SHO |
| Blood Gas CCU - Na | mmol/L | < 120 | > 160 | Contact NCHD/ SHO |
| Blood Gas CCU - iCa | mmol/L | < 0.5 | > 1.58 | Contact NCHD/ SHO |
| Capillary Glucose | mmol/L | ≤ 3.0 | ≥ 20.0 | If tested and result is <4.0, treat as per hospital policy. Repeat after 15minutes. If still <4.0 contact doctor to review. If tested and result is >20, contact the doctor within 30 minutes. Follow treatment pathway in form 2070, the Glucose Monitoring and Insulin Prescription Record. |
| Capillary Ketone | mmol/L | N/A | ≥ 1.0 | If between 1.0mmol/L and 3.0mmol/L contact House doctor and Diabetes Specialist Nurse. If >3.0mmol/L contact Consultant Endocrinologist and Diabetes Specialist Nurse. |
| Pregnancy | N/A | All positive results | | Inform relevant clinical personnel |

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17.0 Pregnancy Testing Fact Sheet

Testing hCG to Screen for Pregnancy

The Basis of the Test

In pregnancy, hCG is normally synthesised and secreted by blastocysts. Hence, implantation and placental development must occur before hCG can be detected in serum or in urine. Typically, implantation occurs about day-23 into the cycle. hCG is excreted into urine in pregnant women and levels of hCG are very similar in both serum and urine at any one time.

The production of hCG is highly variable between individuals for any gestational age.

It takes about 14 days from fertilization to reliably detect pregnancy in more than 99% of pregnancies. Therefore, a negative pregnancy test, whether performed in urine or serum, does not exclude early pregnancy before day post LMP+28 or so.

Comparison between Serum and Urine Tests

The *urine pregnancy hCG test (Alere Sure Step One Step hCG)* performed in the Bon Secours Hospital Cork sufficiently sensitive in most cases to show positive results in pregnancy around the first day of the first missed period in women with normal 28-day cycles. The levels of maternal urinary hCG at that stage are normally 50-250 mIU/ mL, which is easily detected as 'Positive' by the test.

The *serum hCG pregnancy test* performed in the Bon Secours Hospital Cork is also highly sensitive. Serum hCG levels range between 50-500 mIU/ mL by the first day of the first missed period in women with normal 28-day cycles. Results between 5-25 mIU/ mL are as equivocal for pregnancy.

Tests performed in reproductive age females before LMP+28 days may be negative whether or not the person is pregnant and whether the test is done in urine or serum. Urine hCG tests as performed in the Bon Secours Hospital are no less reliable and are very much faster than serum hCG tests.

Note that hCG testing is relatively unreliable in diagnosing ectopic pregnancy.

Regulations and Guidelines

The relevant EU directive (Council Directive 97/43/Euratom of 30 June 1997 on health protection of individuals against the dangers of ionizing radiation in relation to medical exposure, and repealing Directive 84/466/Euratom) and various Learned Society guidelines set out and emphasise the need to operate according to rules of precaution based on time from LMP. They do not appear to mandate or even recommend serum hCG testing when deciding on the advisability of radiological tests in women at risk of being pregnant.

Practicalities

A serum sample for an hCG pregnancy test received in the Biochemistry department will take up to **2 hours** to analyse and report because of fundamental factors involved in the analysis. The average turnaround time for a urinary hCG pregnancy test is **<30 mins** to analyse and report.

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18.0 Dynamic Function Tests Performed In Biochemistry

| Dynamic Test | Measurand(s) | No of samples |
|---|----------------------------|---------------|
| Dexamethasone Suppression Tests | Cortisol, ACTH | 2 sets |
| Synacthen (ACTH Stimulation) Test | Cortisol | 3 sets |
| Investigation of Acromegaly or Gigantism | Glucose, Growth Hormone | 3 sets |
| Oral Glucose Tolerance For The Diagnosis Of Diabetes Mellitus | Glucose | 2 |
| Water Deprivation Test | Serum and Urine Osmolality | 2 or 3 sets |
| TRH (Thyrotropin Releasing Hormone) Test | TSH, Prolactin | 3 |
| LHRH (Luteinising Releasing Hormone) Test | FSH, LH | 3 |
| Ischaemic Forearm Exercise | Lactate | 3 |

Each dynamic function test is described in detail in the following pages, including instructions for patient preparation and interpretation of results. For information on the specific tests, refer to the individual measurand information.

Where a drug is to be administered, it must be prescribed by authorised medical personnel and obtained from the pharmacy.

18.1 Dexamethasone Suppression Tests

Purpose:

Dexamethasone suppression tests determine whether the normal ACTH-dependent secretion of cortisol by the adrenal gland is suppressed in response to the administration of dexamethasone.

There are two different types of dexamethasone suppression tests:

- 1) the low-dose test and
- 2) the high-dose test

Before Performing A Dexamethasone Suppression Test:

Patients in whom a diagnosis of hypercortisolism (such as Cushing's syndrome) is considered should have a **24-hour urine free cortisol** measurement performed first. A normal result excludes hypercortisolism due to pituitary or adrenal disease.

However, very obese persons may over-secrete cortisol.

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Why Are The Tests Performed?

The **low-dose test** can help differentiate healthy (e.g. very obese) people from those who pathologically overproduce cortisol. The **high-dose test** can help determine if the abnormality originates in the pituitary (Cushing's Disease), in the adrenals or in an ectopic site.

The secretion of ACTH from the pituitary gland is normally regulated by the level of cortisol in blood plasma. ACTH stimulates the adrenal cortex to produce cortisol. As plasma cortisol levels increase, ACTH secretion is suppressed. As cortisol levels decrease, ACTH increases.

Dexamethasone is a synthetic steroid similar to cortisol, which suppresses ACTH secretion in normal people. Therefore, giving dexamethasone should reduce ACTH levels, resulting in decreased cortisol levels. People with pituitary glands which produce too much ACTH will have an abnormal response to the low-dose test, but a normal response to the high dose.

Preparation of the Patient for the Test:

Drugs that can affect test results include corticosteroids, oestrogens, oral contraceptives, phenytoin, spironolactone, barbiturates, and tetracyclines. Please discuss the test with the Biochemistry Department in advance if these drugs are being taken.

How The Test Is Performed:

1) The low-dose overnight method:

- Take blood for cortisol and ACTH measurement at 8 a.m. on the first morning of the test.
- Give the patient 1 mg of dexamethasone orally at 11 p.m. and allow the patient to sleep overnight.
- Take blood at 8 a.m. on the second morning for cortisol and ACTH measurement.

2) The high-dose overnight method:

- Take blood for cortisol and ACTH measurement at 8 a.m. on the first morning of the test and label with the actual time.
- Give the patient 8 mg of dexamethasone orally at 11 p.m. and allow the patient to sleep overnight.
- Take blood at 8 a.m. on the second morning for cortisol and ACTH measurement.

Interpretation of Results:

Low dose (1 mg method): Normal people should have serum cortisol concentrations of less than 124 nmol/L following dexamethasone 1 mg. ACTH secretion should be suppressed completely.

High dose (8 mg method):

Serum cortisol concentrations should be less than 50% of the baseline value and ACTH suppressed in normal patients and in those with pituitary-dependant hypercortisolism, following 8 mg dexamethasone.

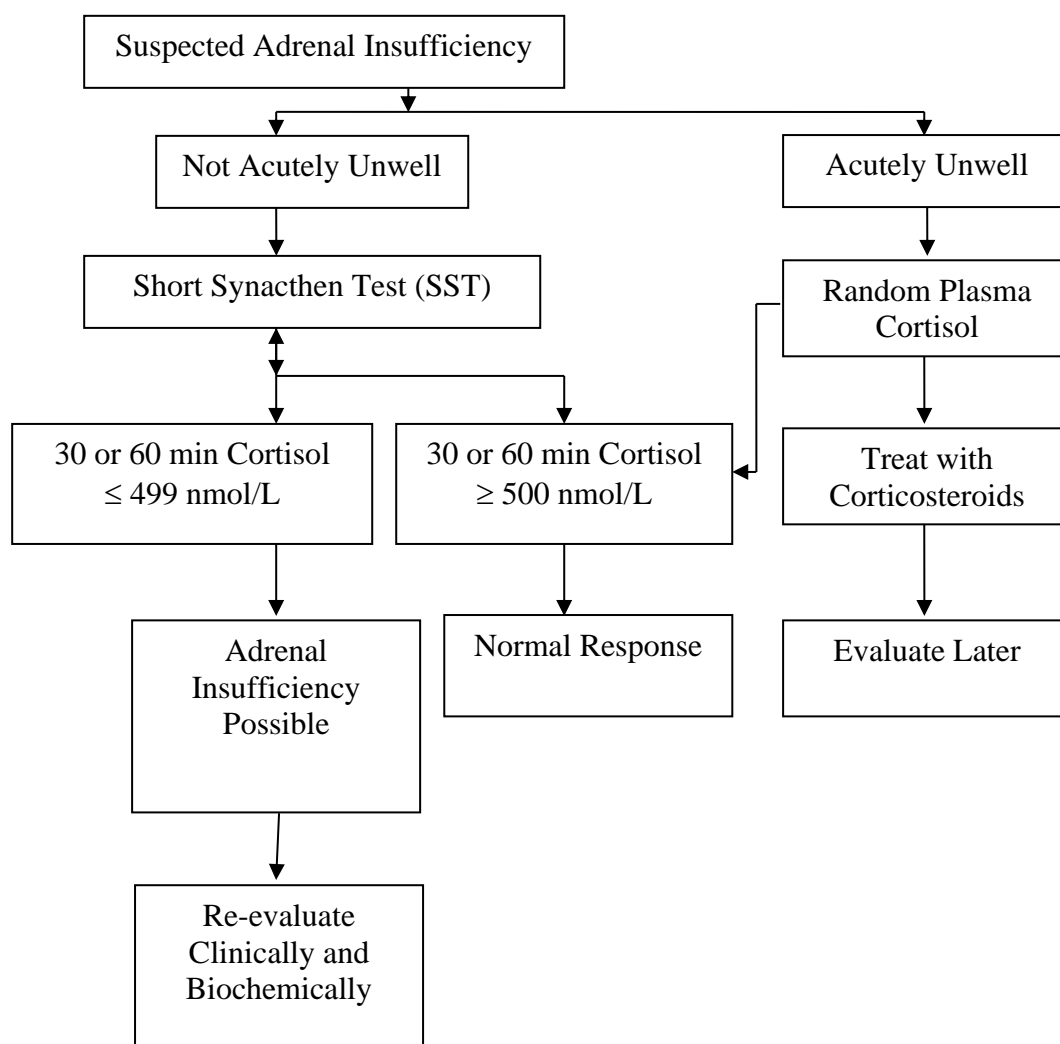
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18.1.1 Suspected Adrenocortical Insufficiency

Patients with adrenal failure may present acutely (with hypoglycaemia, hyponatraemia with hyperkalaemia and dehydration) or chronically with general malaise, anorexia, vomiting, intermittent abdominal pain and weight loss. Pigmentation may be seen on sun exposed areas and also sites of friction such as the palmar creases and buccal mucosa.

If the diagnosis is strongly suspected in acutely ill patients, there should be no delay in administering glucocorticoids as soon as blood has been taken for plasma cortisol and ACTH; the definitive diagnosis can wait. Dexamethasone should be given in this circumstance as the short Synacthen test can be performed the next day since dexamethasone does not interfere with cortisol assays.

Patients on long-term corticosteroid therapy become hypoadrenal due to adrenal atrophy. As a result they may have a poor response to exogenous ACTH, but there is no evidence that the response to ACTH is useful in tailoring glucocorticoid withdrawal.



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18.2 ‘Synacthen’ (ACTH Stimulation) Test

Definition

The Synacthen (ACTH stimulation) test measures the ability of the adrenal cortex to producing cortisol appropriately in response to ACTH.

Tetracosactide, the active substance of Synacthen, consists of the first 24 amino acids occurring in the natural ACTH sequence and displays the same physiological properties as ACTH. It stimulates adrenocortical production of glucocorticoids and mineralocorticoids and, to a lesser extent, of androgens.

Preparing The Patient For The Test:

The test **should not** be performed

- in patients on current regular oral or injected steroid therapy as the results cannot be interpreted.
- In pregnant or in lactating women

How The Test Is Performed:

Cortisol in the blood is measured before and again after an ACTH injection.

- Take a baseline venous blood sample for serum cortisol measurement and label “Time 0” and with the actual time.
- Inject 250 micrograms (the contents of one ampoule of “Synacthen”) intramuscularly into a large muscle, using standard I.M. injection technique.
- Take a second venous blood sample for serum cortisol measurement **30 minutes** after the Synacthen injection and label “30 min” and with the actual time.
- Take a third venous blood sample for serum cortisol measurement **60 minutes** after the Synacthen injection and label “60 min” and with the actual time.

Possible Undesirable Effects Related To Tetracosactide Injection:

Tetracosactide can provoke hypersensitivity reactions, which tend to be more severe in patients susceptible to allergies (especially asthma). Hypersensitivity reactions may include skin reactions at the injection site, dizziness, nausea, vomiting, urticaria, pruritus, flushing, malaise, dyspnoea, and angioneurotic oedema. Isolated cases of adrenal haemorrhage have been reported with Synacthen.

Interpretation:

A normal response to Synacthen is shown by a 30 or 60 min cortisol value of greater than or equal to 500 nmol/L. A cortisol value of less than or equal to 499 nmol/L indicates possible adrenal insufficiency and should be re-evaluated Clinically and Biochemically.

This definition only defines adrenal insufficiency. The definition of normality is problematic since there is considerable variation in healthy individuals and a significant overlap with patients who have adrenal insufficiency. Baseline and incremental cortisol values do NOT apply to women taking oral contraceptives or to pregnant women.

In ACTH deficiency the response to the short test may be normal or reduced. The response to Synacthen is not affected by obesity.

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18.3 Investigation of Acromegaly or Gigantism

IGF-1

The simplest screening test for excess growth hormone (hGH) secretion leading to acromegaly (or to pituitary gigantism in adolescents) is to measure insulin-related growth factor-1 (IGF-1; sometimes called somatomedin-C) in serum. Elevated IGF-1 is quite sensitive and specific for acromegaly in adults but elevated values should be further investigated by measuring the growth hormone response to a glucose load.

Oral GTT to Investigate Growth Hormone Excess:

Note: Random GH values are useless to exclude acromegaly since elevated GH may occur with stress and low values < 5 mIU/L are seen in up to 8% of acromegalic patients who are subsequently identified by the failure of GH to suppress during GTT. This **test is unnecessary** in diabetic patients who should already have a suppressed GH in the presence of hyperglycaemia.

Principle:

GH secretion is part of the counter-regulatory defence against hypoglycaemia and physiological GH secretion is inhibited by hyperglycaemia. In acromegaly, or gigantism, GH secretion is autonomous and does not suppress and may paradoxically rise with hyperglycaemia.

Side Effects:

Some subjects may feel nauseated and may have vasovagal symptoms during this test.

Preparation:

Patients must fast for a minimum of 10 hours before this test but may drink small volumes of water.

Note: The clinician may require samples to be taken at additional times to the standard OGTT used for the diagnosis for Diabetes Mellitus. It is important that the times are stated on the form and exactly what test(s) need to be taken at each time.

Requirements:

Oral Rapolose® OGTT Solution is used to provide the glucose load for this test.

Adults: In adults, a 300 mL dose of Rapolose® OGTT Solution is equivalent to 75g anhydrous glucose as recommended by the World Health Organisation

Children: In children weighing less than 43Kg, the amount of **Rapolose® OGTT Solution** given is related to the child's weight. For children, the recommended test load is 1.75g of glucose per kg body weight up to a total of 75g of glucose, this is equivalent to 7 mL of **Rapolose® OGTT Solution** per kg body weight up to a maximum of 300 mL of **Rapolose® OGTT Solution** (maximum load is 75g of anhydrous glucose). If the patient is a child, then please inform Phlebotomy of the weight of the subject so that the correct dose can be prepared.

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Procedure:

Venous blood samples are taken for GH and glucose (4-5 mL in plain & 1-2 mL in fluoride EDTA tubes). The RapiLOSE® OGTT Solution should be drunk within 5 minutes. Take further blood samples for GH and glucose at the times indicated by the clinician – see note above. Please **send blood both** in plain white and in yellow (Fluoride-EDTA) tubes **at all time points**.

Interpretation:

Normal subjects will exhibit suppression of GH to undetectable values during the test. Acromegalic subjects display either no suppression or a paradoxical rise in GH secretion during the test. Note however that a paradoxical rise in GH may also occur during GTT during normal adolescence.

18.4 Oral glucose tolerance test for the diagnosis of Diabetes Mellitus

Indication

The diagnosis of diabetes is made on the basis of repeatedly elevated fasting plasma glucose (> 7.0). The use of the oral glucose tolerance test is to clarify borderline elevations in fasting plasma glucose, and is required in a minority of patients.

Contraindications:

This test is only necessary if fasting glucose measurements are equivocal. i.e. 6.0 to 7.0 mmol/L.

This test should **not be performed** in patients who fulfil the criteria for diabetes mellitus. These are:

1. a fasting plasma glucose >7.0 mmol/L on two or more occasions and
2. clinical symptoms of diabetes *e.g.* polydipsia, polyuria, ketonuria and rapid weight loss with a random plasma glucose of >11.1 mmol/L).

This test should **not be performed** in patients who are under physical stress *e.g.* post surgery, trauma or infection or extreme psychological stress as these may give misleading results.

This test should **not be performed** in patients with periodic hypokalaemic paralysis.

Principle:

In normal individuals pancreatic insulin secretion maintains blood glucose within a tight concentration range following an oral glucose load. Failure of insulin secretion, or resistance to insulin action, will result in an elevation in blood glucose.

Side Effects:

Some subjects feel nauseated and may have vasovagal symptoms during this test.

Preparation:

Patients should eat a normal carbohydrate diet (>150g daily) for at least 3 days prior to the test and undertake normal physical activity.

Patients must fast for 10-14 hours prior to this test but may drink small volumes of plain water.

Smoking and physical exercise **are not allowed** in the morning prior to, and during, the test.

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Requirements:

Oral RapiLOSE[®] OGTT Solution is used to provide the glucose load for this test.

Adults: In adults, a 300 mL dose of RapiLOSE[®] OGTT Solution is equivalent to 75g anhydrous glucose as recommended by the World Health Organisation

Children: In children weighing less than 43Kg, the amount of **RapiLOSE[®] OGTT Solution** given is related to the child's weight. For children, the recommended test load is 1.75g of glucose per kg body weight up to a total of 75g of glucose, this is equivalent to 7.0 mL of **RapiLOSE[®] OGTT Solution** per kg body weight up to a maximum of 300mL of **RapiLOSE[®] OGTT Solution** (maximum load is 75g of anhydrous glucose). If the patient is a child, then please inform Phlebotomy of the weight of the subject so that the correct dose can be prepared.

Procedure:

- This test should be performed in the morning. Patients should remain at rest during the test.
 - Time 0 min: 2 mL blood should be taken in fluoride EDTA tubes (yellow cap)
 - The RapiLOSE[®] OGTT Solution should be drunk over 5 minutes.
 - Time 120 min: 2 mL blood should be taken in a fluoride EDTA tube (yellow cap)
- There is **no need to take urine** samples for glucose measurements.

Interpretation:

| | Plasma Glucose (mmol/L) | |
|----------------------------|-------------------------|------------|
| | 0 min | 120 min |
| Non diabetic | < 6.0 | < 7.8 |
| Impaired glucose tolerance | 6.1 - 6.9 | 7.9 - 11.0 |
| Diabetic | > 7.0 | > 11.1 |

WHO Diagnostic Criteria 2000:

Symptoms of diabetes (i.e. polyuria, polydipsia and unexplained weight loss) plus:

- a. random plasma glucose concentration > 11.1 mmol/L or
- b. fasting plasma glucose concentration > 7.0 mmol/L (fasting is defined as no calorie intake for at least 8 hours) or
- c. 2 h plasma glucose concentration > 11.1 mmol/L during an oral glucose tolerance test (OGTT). OGTT is not recommended for routine clinical use.

With no symptoms, diagnosis should not be based on a single plasma glucose determination. At least another plasma glucose on another day with a value in the diabetic range is essential, either fasting or at 120 min after a glucose load. If the fasting values are not diagnostic, the 120 min value should be used.

Impaired Fasting Glucose (IFG) And Impaired Glucose Tolerance (IGT):

- a. IFG: fasting plasma glucose > 6.0 mmol/L but <7.0 mmol/L.
- b. IGT: fasting plasma glucose < 7.0 mmol/L and 2 hour plasma glucose during an OGTT > 7.8 but < 11.1 mmol/L.
- c. All subjects with IFG should have an oral glucose tolerance test (OGTT).

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Gestational Diabetes Mellitus (GDM):

Gestational diabetes is defined as any degree of glucose intolerance with onset or first recognition during pregnancy. Strict glucose homeostasis is required during pregnancy in order to reduce the well-described GDM-associated perinatal morbidity and mortality, as well as the associated maternal complications.

Although the majority of women who develop GDM return to normal after delivery, progression to insulin dependent diabetes often occurs in younger, thinner women. Other women have an increased risk of developing NIDDM later in life. It is recommended that women with GDM are retested six weeks after delivery and classified as:

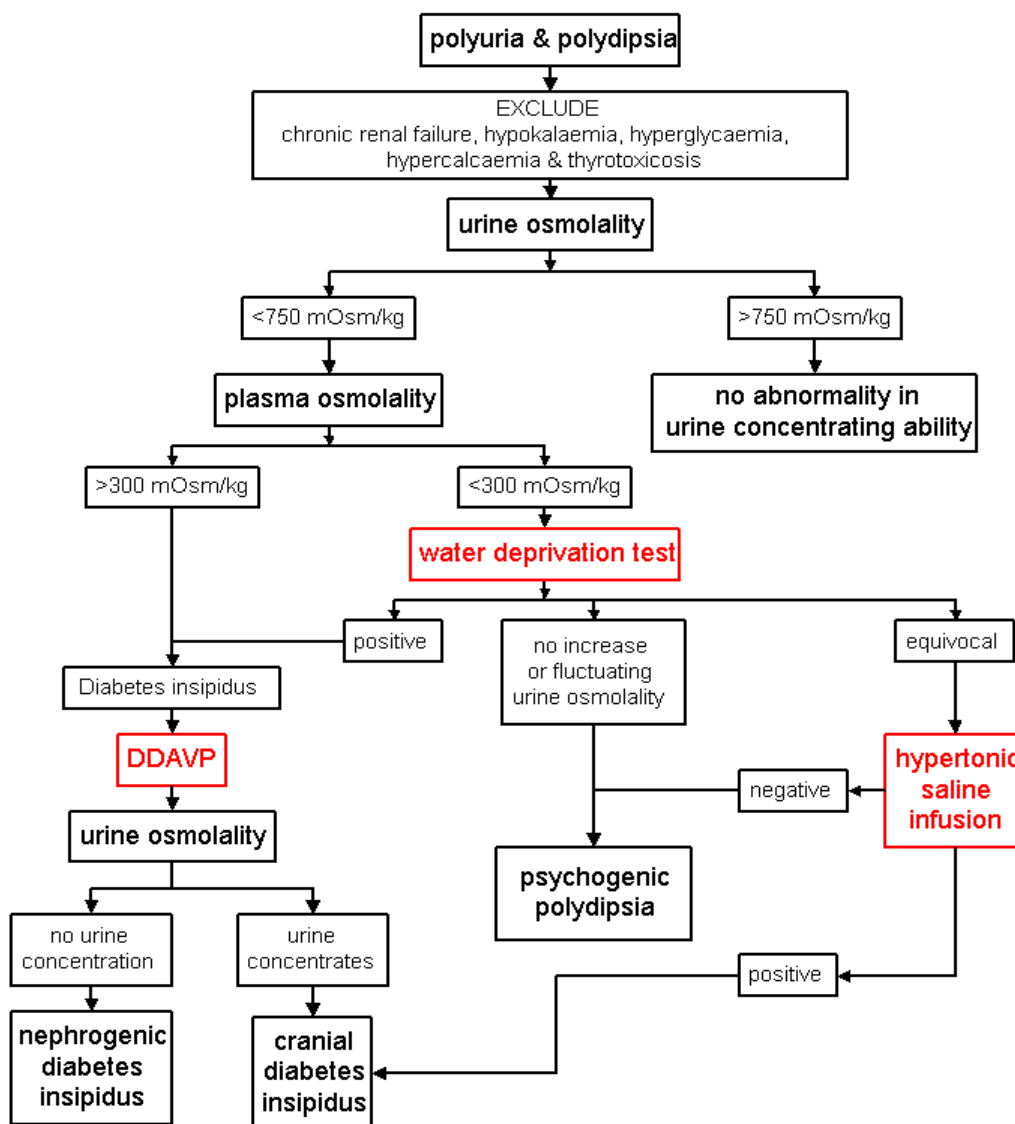
- normal
- IFG (impaired fasting glucose)
- IGT (impaired glucose tolerance)
- diabetic

18.4.1 Investigation of Polydipsia & Polyuria

Polydipsia and polyuria are subjective symptoms, which should be carefully explored in the history to distinguish them from dry mouth and from urinary frequency. A daily loss of > 2.5 L urine with persistent urine osmolalities < 300 mOsm/kg may be considered abnormal. The first line of investigation is to ascertain whether baseline values for urine volumes and plasma osmolality and sodium concentration are in fact abnormal. The next step is to determine if the increased urine production is driven by osmotically active substances excreted in the urine which cause obligate fluid loss *e.g.* glucose. It is then necessary to check if the water loss is due to either intrinsic tubular dysfunction or due to metabolic factors affecting tubular function *e.g.* hypokalaemia or hypercalcaemia. Polyuria is an infrequent manifestation of hyperthyroidism although a proportion of patients do complain of excessive thirst. Often the most difficult patients to diagnose are those with dipsogenic (psychogenic) polydipsia. Many of these patients are investigated with water deprivation tests that are characterised by fluctuating urine volumes and osmolalities that mirror their illicit drinks during the test.

It is important to consider drugs that cause dryness of the mouth as a cause of increased fluid intake.

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18.5 Water Deprivation Test

Indication:

Investigation of suspected cranial or nephrogenic diabetes insipidus and primary polydipsia.

Contraindications:

- The presence of other causes of polydipsia and polyuria.
 - In patients with heart failure and symptomatic ischaemic heart disease
- The **test is not required** if there is evidence for the ability to concentrate urine e.g. a spot urine osmolality > 750 mmol/kg.

Principle of Test:

Water restriction in the normal individual results in secretion of ADH by the posterior pituitary. Failure of this mechanism results in a rise in plasma osmolality owing to water loss, and a dilute urine of low osmolality. The two causes are a failure of ADH secretion and insensitivity of the renal tubules to ADH. They may be distinguished by the administration of DDAVP (synthetic ADH).

Side Effects:

Patients with true diabetes insipidus may become severely water depleted during water deprivation and **MUST** be carefully monitored throughout the procedure.

Requirements:

- Accurate weighing scales for weighing the patient.
- A volumetric cylinder for measuring urine volume.

Performing the Test:

Baseline:

- Weigh the patient
- Calculate and record 5% of this baseline body weight
- Take samples of serum (white plain tube) and urine (MSU tube) for osmolality
- Remove all sources of fluid and food from the patient's access.

Fluid restriction phase:

- Deprive the patient of all fluids and food.
- Hourly:** Weigh the patient, and measure urine output.
- When the patient is unable to tolerate fluid deprivation any longer, weigh, measure urine output, and take samples of serum (white plain tube) and urine (MSU tube) for osmolality. (Please mark actual time on all specimens).

Terminate this phase of the test if:

- the patient becomes distressed by thirst
- The patient loses >5% of baseline body weight
- Plasma osmolality exceeds 300 mOsm/kg

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DDAVP challenge phase

Following the fluid deprivation phase, but **before allowing the patient to drink**

- Administer DDAVP (Desmospray) 20 micrograms (two sprays) intra-nasally
- Measure urine volume and osmolality at 30 and 60 min. afterwards.
- Fluid may be given (200 mL) after 30 min. and should be restricted to less than 500 mL in the 8 hours after the test.

18.5.1 Interpretation of Water Deprivation and DDAVP Test

| Post-dehydration osmolality (mOsm/kg) | | Post DDAVP osmolality (mOsm/kg) | Diagnosis |
|---------------------------------------|-----------------|---------------------------------|--|
| plasma | urine | urine | |
| 283-293 | > 750 | > 750 | normal |
| > 293 | < 300 | < 300 | nephrogenic diabetes insipidus |
| > 293 | < 300 | > 750 | cranial diabetes insipidus |
| < 293 | 300-750 | < 750 | chronic polydipsia |
| < 293 | 300-750 | < 750 | partial nephrogenic DI or primary polydipsia |
| > 293 | 300-750 | > 750 | partial cranial DI |

NB: chronic primary polydipsia can dissipate the renal medullary osmotic gradient, thereby reducing the renal response to endogenous and exogenous AVP. In cranial DI, maximal urinary concentration may be achieved only after repeated DDAVP.

18.6 TRH (Thyrotropin Releasing Hormone) Test

Definition:

The TRH (Thyrotropin Releasing Hormone) Test measures the ability of the pituitary to producing TSH (thyrotropin) appropriately in response to TRH.

Indication:

The TRH test is indicated in the investigation of pituitary secretion of TSH and in distinguishing thyroid hormone resistance from 2^o hyperthyroidism in cases of high TSH and high thyroxine levels in plasma.

The TRH test may be combined with the GnRH (LHRH) test as a partial combined test of pituitary function.

Contraindications:

TRH can cause smooth muscle spasm and should be used with caution in patients with asthma or ischaemic heart disease. The TRH test should not be used in pregnant women. Patients should not have taken thyroxine for 3 weeks prior to this test.

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Principle:

TRH (thyrotropin releasing hormone) is a tripeptide secreted by the hypothalamus that stimulates the production and secretion of TSH by the anterior pituitary. TRH also stimulates prolactin release. Protirelin is a synthetic form of TRH.

Side Effects:

Most adult patients express an urgent need but inability to pass urine. Other side effects include flushing, dizziness and a metallic taste in the mouth.

Preparation:

No specific patient preparation is required

Requirements:

Protirelin (TRH) 200 mcg) for IV injection

The dose for children is 7 mcg/kg to a maximum 200 mcg.

Procedure:

| | |
|-------------|--|
| time 0 min | take 3 mL blood for TSH and Prolactin immediately give TRH I.V. as a bolus (dose as above) |
| time 30 min | take 3 mL blood for TSH and Prolactin |

Note: GnRH may be administered I.V. directly before or after TRH as part of a partial combined pituitary test.

Interpretation:

- Normal basal values of TSH should be 0.2-6 IU/mL. The normal increment in TSH at 30 min should be 5-30 (mean 15) IU/mL with a slight diminution at 60 min.
- Exaggerated TSH response is seen in primary hypothyroidism.
- A flat response is seen in primary hyperthyroidism; but also in some apparently euthyroid patients with ophthalmic Graves disease or multinodular goitre.
- A delayed response with the TSH concentration lower at 30 than 60 min may be seen in hypothalamic dysfunction.
- Various drugs can modify the TSH response.
- The TSH response is flat in most cases of TSHoma whereas in thyroid hormone resistance the TSH response is brisk.
- Prolactin secretion is normally stimulated by TRH administration.

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18.7 LHRH (Lutinsing Releasing Hormone) Test

Indications:

- To tell the difference between primary and secondary hypogonadism.
- To diagnose hypothalamic-pituitary disease in precocious and delayed puberty in both sexes in children with low basal gonadotropins.
- To evaluate low testosterone levels in men or low oestradiol levels in women

Principle:

GnRH (gonadotropin releasing hormone) is a decapeptide secreted by the hypothalamus, which stimulates the production and secretion of LH and FSH by the anterior pituitary.

Side Effects:

GnRH may rarely cause nausea, headache and abdominal pain.

Preparation:

Patients should not be on oestrogen therapy for four weeks prior to the test.

Requirements:

HRH[®] (recombinant LHRH) 100 micrograms.

Procedure:

| | |
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| time 0 min | Take 2 mL blood for LH & FSH Immediately give HRH [®] 100 mcg I.V. as a bolus (dose as above) |
| time 30 min | Take 2 mL blood for LH & FSH |

Interpretation:

- An exaggerated response is seen in primary & secondary gonadal failure.
- Following GnRH, the response may be considered normal if the basal values are in the reference range and there is at least a doubling at 20 min for LH and FSH. The response varies throughout the menstrual cycle: early (D4) < late follicular (D11) = "luteal" (D21), max response occurs at the mid-cycle (D14).
- Normal basal reference values in pre-pubertal children are:-
LH < 2.0 IU/L
FSH < 2.0 IU/L
- A flat response in gonadotropins (< 5 IU/L) occurs in pre-pubertal children and with pituitary and/or hypothalamic disease. However, a normal response does NOT exclude pituitary or hypothalamic disease since the response will be affected by the exact anatomy of the disorder.
- The magnitude of the LH response is proportional to the mean nocturnal LH and therefore the evolution of puberty

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18.8 Ischaemic Forearm Exercise Test

Indication:

The differential diagnosis of metabolic causes of muscle weakness, fatigue and cramps.

Principle of the Test:

Normal subjects exhibit a rise in lactate during ischaemic exercise. Glycogenolysis and glycolysis proceed to lactate during ischaemic exercise as oxygen is required for further oxidation in the Krebs cycle.

Side Effects:

This test is uncomfortable to perform and the subject will need encouragement to ensure that sufficient exercise is performed for a valid test.

Preparation:

The subject should rest for 30 min prior to the test.

Requirements:

- 4 special tubes for lactate (discuss with lab) and 4 plain white tubes for CK
- Manually operated Sphygmomanometer (not 'Dynamap'-type electronic model)
- A soft rubber ball

Procedure:

At each time point samples should be taken for lactate and CK (1 mL).

- A sphygmomanometer cuff is placed on the upper arm and an intravenous cannula inserted in the antecubital vein, which should be kept patent with saline.
- Allow three minutes to elapse after the cuff has been relaxed to allow free perfusion of the arm.
- Two **baseline** blood samples are drawn from the arm.
- The cuff is inflated above systolic pressure and the subject should squeeze the rubber ball once every few seconds for 2 minutes or as long as it can be tolerated. It is essential that the exercise should continue to produce muscle pain and that the fingers are extended fully between contractions. Patients may find it helpful and encouraging to be advised of the time they have exercised.
- The cuff is deflated.
- Two blood samples (**T0**) are immediately taken from the now hyperaemic arm as soon as blood has flushed through, and at 1 (**T1**) and 2 (**T2**) mins later.

Interpretation:

Normal subjects obtain relatively instantaneous relief of pain and can move their fingers immediately on release of the cuff. Patients with metabolic defects often cannot exercise for 2 minutes, develop a marked forearm cramp, and are unable to extend their fingers.

A normal response is shown by maximum rises (between baseline and peak values) in plasma lactate > 2.2 mmol/L. The absence of a venous blood lactate response to ischaemic exercise is characteristic of all diseases in which there is impairment of conversion of glycogen to glucose or lactate in muscle (e.g. McArdle's syndrome).

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19.0 PATHOLOGY PROFILES

| CHRONIC LIVER SCREEN (FASTING) |
|---|
| Hepatitis B Antigen, Hepatitis C, Autoantibody Screen, RA Screen, Ferritin, Iron Profile, Coeliac Screen, Fasting Lipids, Thyroid Function, Liver Function Tests, Alpha 1 Antitrypsin |
| Sample Requirements |
| Blood Tube (4.9mL) containing no additive x 2 Blood Tube (4.9mL) containing no additive x 3 |
| Turn Around Time |
| Refer to individual tests for turnaround time in this document |

| CHRONIC LIVER SCREEN (NON-FASTING) |
|---|
| Hepatitis B Antigen, Hepatitis C, Autoantibody Screen, RA Screen, Ferritin, Iron Profile, Coeliac Screen, Non Fasting Lipids, Thyroid Function, Liver Function Tests, Alpha 1 Antitrypsin |
| Sample Requirements |
| Blood Tube (4.9mL) containing no additive x 2 Blood Tube (4.9mL) containing no additive x 3 |
| Turn Around Time |
| Refer to individual tests for turnaround time in this document |

| COVID BLOOD PROFILE |
|---|
| Full Blood Count, Prothrombin Time, International Normalised Ratio, Activated Partial Thromboplastin Time, Fibrinogen, D-Dimer, C-Reactive Protein, Ferritin, U&E, Creatinine |
| Sample Requirements |
| Blood Tube (4.9mL) containing no additive x 1 Blood Tube (2.7mL) containing EDTA x 1 Blood Tube (2.7mL) containing Sodium Citrate x 1 |
| Turn Around Time |
| Refer to individual tests for turnaround time in this document |

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20.0 REPORTING OF TEST RESULTS

20.1 Customer Queries with Respect to Test Results

Refer to section 3.3 of this document titled "Availability of Clinical and Scientific Advice".

20.2 Reporting of Results within the Hospital

All results, once released, are available on the hospital computer system (PIMS). Hardcopy reports, where applicable, are printed for distribution to ward locations once per day for inclusion in the patient file. In addition, designated Doctor's receive an additional copy of their reports on a daily basis.

20.3 Reports for External Locations

Reports are issued to external customers in hard copy format or electronically through result reporting system (Medibridge). All such reports are posted on the day of testing if results are available before 3.30 p.m. Where results are not available, they will be posted the next working day. Note reports traceable to Bon Secours Glasnevin and Bon Secours Tralee are issued directly to printers located in the respective Pathology Departments.

20.4 Telephoned Results

- It is the policy of the Pathology Department to telephone reports only when results for specific clinical parameters have reached critical levels or where the Clinician/ nominee has requested for results to be phoned.
- **At ward level where information is received concerning verbal reports then a record must be maintained of the communication using Form 37a titled "Verbal/ Telephone Order Form".**

20.4.1 External Telephone Results

It is the policy of the Pathology department not to telephone results to external practitioners or patients with the exception of:-

- Warfarin patients following attendance at the outpatient warfarin clinic
- Where specific parameters have reached critical levels

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20.5 Reference Ranges (Biological Reference Intervals)

Reference ranges for test attributes are documented on all reports.

Warning: Many diaries and handbooks provide lists of reference intervals for common analytes. You are asked not to refer to these in the interpretation of results generated by the Pathology Laboratory. We have prepared our own reference intervals which are dependent on the method of analysis used and are also specific to the population which we serve. The use of inappropriate reference intervals can be at best confusing and at worst dangerous. If you are in any doubt about the validity of any reference interval provided to you, please contact the Pathology Laboratory for clarification.

20.6 Interpretation of Numerical Results

Note: The information provided below for flagging/ marking of results is valid for numerical results where the test has an associated reference range defined in the Laboratory Information System. Not all numerical tests have defined reference ranges so will not be flagged/marked as abnormal.

20.6.1 **Hardcopy Reports**

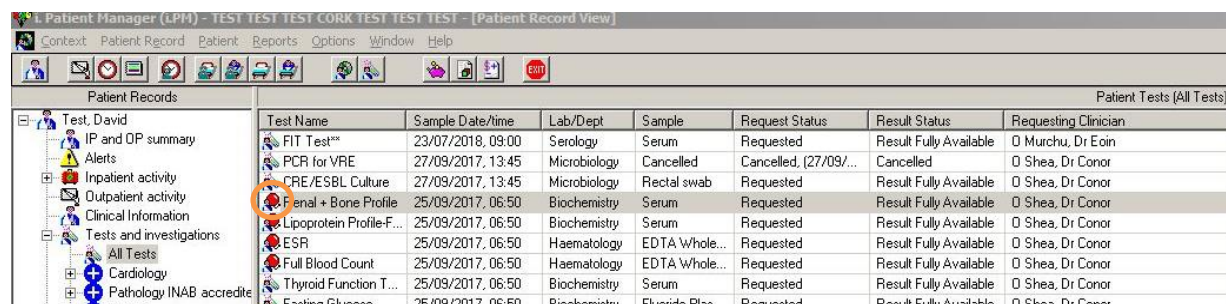
Results are indicatively marked as follows:-

| | | |
|----|---|----------------------|
| H | - | Abnormally High |
| HH | - | Very Abnormally High |
| L | - | Abnormally Low |
| LL | - | Very Abnormally Low |

Within range results have no associated marking or comment.

20.6.2 **Electronic Reports on PIMS (Hospital System)**

- (a) **Using patient record view**, a master list of test requests appears on screen. A red balloon preceding the test name indicates or flags an abnormal result. Refer to the attached printscreen.


















| Test Name | Sample Date/time | Lab/Dept | Sample | Request Status | Result Status | Requesting Clinician |
|--------------------------|-------------------|--------------|------------------|-----------------------|------------------------|----------------------|
| FIT Test™ | 23/07/2018, 09:00 | Serology | Serum | Requested | Result Fully Available | O Murchu, Dr Eoin |
| PCR for VRE | 27/09/2017, 13:45 | Microbiology | Cancelled | Cancelled, (27/09/... | Cancelled | O Shea, Dr Conor |
| CRE/ESBL Culture | 27/09/2017, 13:45 | Microbiology | Rectal swab | Requested | Result Fully Available | O Shea, Dr Conor |
| Fecal + Bone Profile | 25/09/2017, 06:50 | Biochemistry | Serum | Requested | Result Fully Available | O Shea, Dr Conor |
| Lipoprotein Profile-F... | 25/09/2017, 06:50 | Biochemistry | Serum | Requested | Result Fully Available | O Shea, Dr Conor |
| ESR | 25/09/2017, 06:50 | Haematology | EDTA Whole... | Requested | Result Fully Available | O Shea, Dr Conor |
| Full Blood Count | 25/09/2017, 06:50 | Haematology | EDTA Whole... | Requested | Result Fully Available | O Shea, Dr Conor |
| Thyroid Function T... | 25/09/2017, 06:50 | Biochemistry | Serum | Requested | Result Fully Available | O Shea, Dr Conor |
| Fasting Glucose | 25/09/2017, 06:50 | Biochemistry | Fluoride Plas... | Requested | Result Fully Available | O Shea, Dr Conor |

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(b) On review of individual test results the analysis column in PIMS notes the following for each result:-

- Within Range
- Abnormally Low
- Very Abnormally Low
- Abnormally High
- Very Abnormally High

Refer to the attached printscreen.

| Display Test Result Details - Renal + Bone Profile - Mrs David Test 258864 F 03/07/1928 | | | | |
|---|-------|--------------------------|------------|-----------------|
| Result Values | | Sample and Result Source | | |
| Result Name | Value | Units | Ref. Range | Analysis |
|  Sodium | 137 | mmol/L | 136-145 | Within Range |
|  Potassium | 5.1 | mmol/L | 3.5-5.1 | Within Range |
|  Chloride | 101 | mmol/L | 98-107 | Within Range |
|  Urea | 18.2 | mmol/L | 3.5-7.2 | Abnormally High |
|  Creatinine | 164 | umol/L | 49-90 | Abnormally High |
|  CKDEPI | 24 | mL/min/1.73... | N/A | Not Applicable |
|  Urate | 473 | umol/L | 150-350 | Abnormally High |
|  Calcium | 2.33 | mmol/L | 2.10-2.55 | Within Range |
|  Corr. Calcium for Al... | 2.57 | mmol/L | 2.10-2.55 | Abnormally High |
|  Inorganic Phosphate | 1.14 | mmol/L | 0.74-1.52 | Within Range |
|  Total Protein | 56 | g/L | N/A | Not Applicable |
|  Albumin | 28 | g/L | 34-48 | Abnormally Low |
|  Globulin | 28 | g/L | 25-35 | Within Range |
|  Alkaline Phosphatase | 107 | IU/L | 35-110 | Within Range |
|  Magnesium | 0.79 | mmol/L | 0.7-1.07 | Within Range |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| | | | | |

20.6.3 Electronic Reports on Medibridge Results Reporting System

Results are indicatively marked as follows:-

- *H - Abnormally High
- *L - Abnormally Low

Within range results have no associated marking or comment.

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20.6.4 Interpretation of Textual Results

For textual results, the interpretive comment will not be flagged, but rather incorporated into the body of the report (hardcopy/electronic).

20.7 Specimen Type on Blood Reports

Please note as per accreditation requirements our test reports identify the specific specimen type used in the testing process.

20.8 Authorisation of Test Reports

The table below identifies clinical responsibility for the authorization of test reports as it relates to each Laboratory discipline.

| Laboratory Discipline | Position | Deputy |
|---|---|---|
| Biochemistry | Consultant Chemical Pathologist (Dr. Michael Louw) | Consultant Chemical Pathologist (Prof. Carel Le Roux) |
| Blood Transfusion/ Haematology | Consultant Haematologist (Dr. Eileen Kelleher) (Dr. Susan O'Shea) (Dr. Khalil Alnajjar) | Consultant Haematologist (Dr. Eileen Kelleher) (Dr. Susan O'Shea) (Dr. Khalil Alnajjar) |
| Histopathology including Diagnostic Cytology | Consultant Pathologist (Dr. Triona Hayes, Dr. Paul Ryan, Dr. Aoife McCarthy, Dr. Adeline Chelliah, Dr. Juan Pinto, Dr. Adeyemi Idowu) | Consultant Pathologist (Dr. Triona Hayes, Dr. Paul Ryan, Dr. Aoife McCarthy, Dr. Adeline Chelliah, Dr. Juan Pinto, Dr. Adeyemi Idowu) |
| Microbiology including serology | Consultant Microbiologist (Dr. Olive Murphy, Dr. Marianne Fraher, Dr. Deirdre O'Brien) | Consultant Microbiologist (Dr. Olive Murphy, Dr. Marianne Fraher, Dr. Deirdre O'Brien) |
| Immunology | Consultant Immunologist (Prof. Conleth Feighery) | Consultant Immunologist TBC |

20.9 Requirements Regarding Patients

Where appropriate, there is open disclosure to relevant persons i.e. requesting Clinician, of incidents that resulted or could have resulted in patient harm. It is the policy of Pathology Bon Secours Hospital, Cork to issue test results to the requesting Clinician and not to the patient. Test results may be released to additional bodies /agencies including but not limited to GPs, insurers, statutory bodies and in transition of care events.

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21.0 CUSTOMER COMPLAINTS

- 21.1** The Pathology department operates a complaints system. The objectives of our complaints handling system are:-
- That all complaints are rapidly and effectively handled.
 - The customer and/or patient difficulties are alleviated promptly.
 - That the same problem will not occur again because the cause has been identified and corrected.
 - That customer confidence is restored in our service.
 - That relevant information is recorded and reported to Clinical Directorate and the Laboratory Services Manager.

- 21.2** If the service provided is not satisfactory, please contact the Pathology Department/ Laboratory Services Manager/ Laboratory Quality Assurance Officer to process the complaint.

22.0 DATA PROTECTION

The Bon Secours Health System is registered with the Data Commissioner in Ireland to capture and process patient information. It is the policy of the Pathology department to manage data and information with the highest degree of integrity, security and confidentiality.

The Laboratory is responsible for the management of all patient information obtained or created during the performance of Laboratory activities. Management of patient information shall include privacy and confidentiality.

The Laboratory shall inform the user and/or patient in advance of the information it intends to place in the public domain.

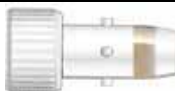





When required by law the Laboratory may share patient data without their consent; examples include provision of data to the National Cancer Registry or to the Health Protection Surveillance Centre (notifiable diseases), National Haemovigilance Office (Serious Adverse Events/Serious Adverse Reactions). This is noted to the patient during the hospital admission process.

A copy of Bon Secours Data Protection and Privacy Statement is available within the Laboratory Outpatients reception area for patients who attend Laboratory Outpatients.

On www.bonsecours.ie on the webpage footer click on “data protection and privacy”. This describes the Health Systems description of how it collects and uses personal data in a way that is consistent with the obligations and patient rights under the General Data Protection Regulation.

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23.0 GUIDE TO THE USE OF BLOOD TUBES FOR ROUTINE LABORATORY TESTS

| TUBE CAP COLOUR/ CONTENTS (EUROPEAN) | COLOUR | APPLICATION |
|---|---|---|
| White (No Anticogulant) |  | Clinical Chemistry/ Virology/ Bacteriology/ Immunology |
| Orange (Lithium Heparin) |  | Troponin |
| Red/ Pink (EDTA) |  | F.B.C./ ESR/ Blood Transfusion |
| Yellow (Fluoride EDTA) |  | Glucose (Adult) |
| Yellow (Heparin Fluoride) |  | Glucose (Paediatric) |
| Green (Trisodium Citrate 1:9) |  | Coagulation |

Where multiple blood tubes are to be drawn, the tubes should be taken in the following sequence:-

| | | |
|--|---|-------|
| Clotted Citrate Lithium Heparin EDTA Fluoride | } | CCLEF |
|--|---|-------|

Reference: Gurr et al "Musterstandardarbeitsanweisung Präanalytik" J Lab Med 2011

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24.0 **PATHOLOGY CRITICAL TEST RESULT VALUES**

Process for Verbal Communication of Critical Results

- ❖ Policy requires that the clinician responsible for patient care is verbally alerted to Pathology critical test results in <60 mins from the time the Laboratory has released the results.
- ❖ The critical values listed have been defined by the Laboratory Consultant in charge of each scientific discipline and agreed with Paediatric Consultants in the case of paediatric values.
- ❖ **Paediatric = Child <16yrs**
- ❖ A critical result value is defined as "A critical test result value is a markedly abnormal test result that may signify a pathophysiological state that may be life threatening or of immediate clinical significance and that requires urgent action".
- ❖ An episode is one hospital stay and it can be composed of a number of care events.
- ❖ To comply with Hospital/ Laboratory accreditation and clinical requirements please note:-
 - Laboratory will request and document the following information from clinical personnel in receipt of a verbal notification for a critical test result:-
 - Clinical staff member's full name and title
 - Confirm clinical location
 - Read back the test results
 - Laboratory staff must phone clinical personnel (Phone to inform ward, Consultant, GP or external agency) within 30 mins of releasing the test result
 - Clinical personnel on receipt of a critical result will:-
 - Document the verbal communication of results on from 37a
 - Ensure clinician/ nominee responsible for patient care is alerted within 30 mins of the Laboratory communication.

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BIOCHEMISTRY CRITICAL TEST RESULT VALUES – CATEGORY A

Results require communication within 30 minutes. This classification indicates potential immediate danger to the patient, or a potentially life-threatening illness when urgent intervention is required.

| Parameter | Unit of Measurement | Lower Limit ≤ | Higher Limit ≥ | Required Action by Laboratory |
|---------------------------------|---------------------|---------------|------------------------|--|
| Sodium | mmol/L | 120 | 150 | Repeat and phone first time in an episode and if value deteriorates by more than 5 mmol/L. Note for MAU: Repeat and phone Sodium values < 126 mmol/L first time in an episode and if value deteriorates by more than 5 mmol/L. |
| Sodium (Paediatric <16 yrs) | mmol/L | 128 | 145 | Phone first time in an episode and if value deteriorates by more than 3.0 mmol/L. |
| Potassium | mmol/L | 2.5 | 6.0 | Repeat and phone first time in an episode and if value deteriorates by more than 0.5 mmol/L. |
| Adjusted Calcium | mmol/L | 1.8 | 3.0 | Phone first time in an episode and if value deteriorates by more than 0.3 mmol/L. Calcium to be phoned if there is no adjusted calcium result |
| Urea | mmol/L | N/A | 30.0 | Phone the first time in an episode and if the value rises by more than 10 mmol/L per day. |
| Urea (Paediatric <16 yrs) | mmol/L | N/A | 10.0 | Phone the first time in an episode and if the value rises by more than 10 mmol/L per day. |
| Creatinine | μmol/L | N/A | 354 | Phone the first time in an episode and if the value rises by more than 100 μmol/L per day |
| Creatinine (Paediatric <16 yrs) | μmol/L | N/A | 200 | Phone the first time in an episode and if the value rises by more than 100 μmol/L per day |
| CKDEPI | ml/Min | 15 | N/A | Phone the first time in an episode. |
| Amylase | IU/L | N/A | 250 | Phone the first time in an episode and if the value rises by more than 100 IU/L per day |
| Phosphate | mmol/L | 0.3 | N/A | Phone the first time in an episode. |
| Magnesium | mmol/L | 0.4 | N/A | Phone the first time in an episode. |
| Glucose | mmol/L | 3 | 20 | Phone the first time in an episode. |
| Glucose (Paediatric <16 yrs) | mmol/L | 3 | 15 | Phone the first time in an episode. |
| Ammonia | μmol/L | N/A | 100 | Phone the first time in an episode and all Ammonia results to the Mercy |
| BNP | ng/L | N/A | 500 | Phone the first time in an episode. |
| Lactate | mmol/L | N/A | 2.0 4.0 | Phone the first time in an episode. Phone all values >4.0 mmol/L. |
| pH | pH units | 7.3 | 7.6 | Phone. |
| Troponin | ng/L | N/A | Female >16 Male >34 | Phone the first time in an episode. |
| CK-total (Creatinine Kinase) | IU/L | N/A | 5000 | Phone the first time in an episode. |
| Beta-hCG (Pregnancy) | mIU/mL | N/A | >5 | Phone*. |
| CRP | mg/L | N/A | 300 | Phone the first time in an episode. |
| CRP (Paediatric <16 yrs) | mg/L | N/A | 100 | Phone the first time in an episode. |

Note: It is not necessary to phone Beta-hCG to Fertility Clinics unless marked urgent or instruction for a result to be phoned is indicated on the request form.

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BIOCHEMISTRY CRITICAL TEST RESULT VALUES – CATEGORY B

Results require communication within 24 hours.

| Parameter | Unit of Measurement | Lower Limit ≤ | Higher Limit ≥ | Required Action by Laboratory |
|---|---------------------|---------------|-------------------------------|---|
| ALT | IU/L | N/A | 510 | Phone the first time in an episode. |
| ALT (Paediatric <16 yrs) | IU/L | N/A | 510 | Phone the first time in an episode. |
| AST | IU/L | N/A | 510 | Phone the first time in an episode. |
| AST (Paediatric <16 yrs) | IU/L | N/A | 510 | Phone the first time in an episode. |
| Triglyceride | mmol/L | N/A | 20 | Phone the first time in an episode. |
| Free T4 | pmol/L | 8 | 50 | Phone the first time in an episode. |
| TSH | IU/L | N/A | 30.0 | Phone the first time in an episode. |
| Prolactin (non-pregnant) | mIU/L | N/A | 1000 | Phone the first time in an episode. |
| Vitamin B12 | pg/L | 148 | N/A | Phone the first time in an episode. |
| Cortisol Unless part of Dexamethasone suppression test | nmol/L | <50 | None | Phone. |
| Cortisol (30 mins Post Syn) | nmol/L | <250 | None | Phone. |
| Gentamicin (Pre/ Random/ Unlabelled) | mg/L | N/A | >1 | Phone. |
| Gentamicin (Post) | mg/L | N/A | >5 | Phone. |
| Tobramycin | mg/L | N/A | >1 | Phone. |
| Vancomycin | mg/L | N/A | >20 | Phone. |
| DPD Gene Screening | N/A | N/A | N/A | Inform Dr Bird if Mutation is identified. |
| Paraprotein | g/L | Any IgE/IgD | IgG >15 IgA >10 IgM >10 | Phone the first time identified |
| Hypogammaglobulinaemia | g/L | IgG <3 | N/A | Phone the first time identified |

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HAEMATOLOGY CRITICAL TEST RESULT VALUES

| Parameter | Units of Measurement | Lower Limit | Oncology Patient Limits # | Higher Limit | Required Action by Laboratory |
|---|----------------------|--|---------------------------|--------------|---------------------------------|
| Haemoglobin § | g/dl | <8.5 | <7.5 | Male: >20 | *Phone |
| | | | | Female: >18 | |
| White cell count * | X 10 ⁹ /L | <3.0 | <1.0 | >30 | *Phone |
| Patients with known CLL | X 10 ⁹ /L | N/A | N/A | >100 | |
| Platelets | X 10 ⁹ /L | <30 | <20 | >1000 | *Phone |
| Platelet (previously within normal range) | X 10 ⁹ /L | <75 | N/A | N/A | *Phone |
| Platelet (Paediatric <16 yrs) | X 10 ⁹ /L | <40 | N/A | >1000 | *Phone |
| Neutrophil Count | X 10 ⁹ /L | <0.5 | <0.5 | N/A | *Phone |
| Blood Film | N/A | Presence of blast cells on first presentation or any morphology suggestive of blood disorder requiring clinical intervention | | | *Phone Consultant Haematologist |
| | | | | | |
| Prothrombin Ratio (INR) | | | | | *Phone |
| Non-anticoagulated | ratio | N/A | N/A | >1.2 | |
| Anticoagulated | ratio | N/A | N/A | >5.0 | |
| Activated Partial Thromboplastin Time (APTT) | | | | | *Phone |
| Non-anticoagulated | sec | N/A | N/A | >32 | |
| Anticoagulated | sec | N/A | N/A | >70 | |
| Fibrinogen | g/L | <1.0 | N/A | N/A | *Phone |
| Heparin Induced Thrombocytopenia Screen (HITs) | N/A | All Positive Results | | | *Phone |
| | | | | | |
| Infectious Mono Screen (Monospot) | N/A | All Positive Results | | | *Phone |
| Pregnancy Test (Urine) | N/A | All Positive Results | | | *Phone |
| Malaria and Sickle Cell Screen Screen | N/A | All Positive Results | | | *Phone |

- # Oncology patients include those from the following locations: BERN, VER, SVPAC, ONC, OLC, Radiotherapy (UPMC) as well as those under the care of an Oncology/ Haematology Consultant.
- § If a patient Haemoglobin has been <8.5 (or <7.5 for oncology patients) and has decreased by 1.5g/l then the result will be phoned.
- * If a patient WBC has previously been >30 and has subsequently increased by >50% then the result will be phoned.
- ⊗ If a patient has FBC results that are in keeping with a known diagnosed blood disorder, then results will not be phoned, however, parameters that may influence ordering blood product support will be communicated.

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BLOOD BANK CRITICAL TEST RESULTS

| Parameter* | Units of Measurement | Lower Limit | Higher Limit | Required Action by Laboratory |
|---|----------------------|-------------|---------------------------|---------------------------------|
| Direct Antiglobulin Test | N/A | N/A | 3+ or 4+ with Hb <8.5g/dl | *Phone Consultant Haematologist |
| A new red cell antibody where transfusion is required urgently when there could be a delay in finding compatible blood | N/A | | N/A | *Phone |
| Wrong blood in tube – an unexpected change in blood group compared to a historical blood sample, query misidentified patient. | N/A | | N/A | Request urgent resampling |

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HISTOPATHOLOGY CRITICAL TEST RESULT VALUES

| Test | Critical Test Value | Required Action by Laboratory |
|-----------------|----------------------------|--------------------------------|
| Frozen section* | All results when available | Phone to Consultant in Theatre |

* For frozen sections, results will be phoned by the Consultant Pathologists within 20 mins of specimen registration.

Note: The referring clinician is responsible for ensuring both that they have indicated any degree of clinical urgency to the Laboratory, and that they have received and acted upon the report. This primary responsibility is not dependent on any communication from the Laboratory.

Pathologists should use their experience and judgement to identify unexpected/ critical results/ diagnoses such as unexpected finding of malignancy, cases where a diagnosis is significantly modified after the initial report.

Pathologists should communicate directly with clinicians, using a satisfactory method of communication based on experience/ judgement, preferably on the same day on which the diagnosis is made, when circumstances permit.

IMMUNOLOGY CRITICAL TEST RESULT VALUES

| Test | Critical Test Value | Required Action by Laboratory |
|--|--|--|
| ANCA | Positive pANCA and MPO antibodies or cANCA and PR3 antibodies (first time) | Phone Clinician and notify Consultant Immunologist by e-mail |
| GBM Antibodies | Positive (first time) | Phone Clinician and notify Consultant Immunologist by e-mail |
| Liver Kidney Microsomal Antibodies (LKM) | Positive (first time) | Phone Clinician and notify Consultant Immunologist by e-mail |
| Anti NMDA Antibodies | Positive (first time) | Phone Clinician and notify Consultant Immunologist by e-mail |

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MICROBIOLOGY CRITICAL TEST RESULT VALUES

| Test | Critical Test Value | Required Action by Laboratory |
|----------------------------|---|--|
| Blood Culture (Gram Stain) | All positive Blood Culture gram stain results | Phone to inform ward, Consultant, GP or external agency (EXCEPT partner bottle of a blood culture set previously reported as positive to the ward with the same gram result) |
| CSF | All positive CSF direct microscopic results | Phone to inform ward, Consultant, GP or external agency |
| Joint Fluid | All positive Joint Fluid gram stain results | Phone to inform ward, Consultant, GP or external agency |

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25.0 LIST OF TESTS PERFORMED ON-CALL

BIOCHEMISTRY

Profiles

| |
|--|
| ADMISSION PROFILE (Biochemistry Section) |
| Sodium, Potassium, Chloride, Urea, Creatinine, Urate, Calcium, Phosphate, Total Bilirubin, Direct Bilirubin, Alkaline Phosphatase (ALP), Aspartate Aminotransferase (AST), Alanine Aminotransferase (ALT), Gamma-Glutamyl Transferase (GGT), Total Protein, Albumin, Globulin, Glucose, Magnesium (Mg) |
| CHILDREN'S WARD BIOCHEMISTRY |
| Sodium, Potassium, Chloride, Urea, Creatinine, Urate, Calcium, Magnesium (Mg), Total Protein, Albumin, Globulin, Alkaline Phosphatase (ALP), Alanine Aminotransferase (ALT), Aspartate Aminotransferase (AST), C – Reactive Protein (CRP), Random Glucose |
| LIPOPROTEIN PROFILE (FASTING) |
| Cholesterol, Triglycerides, High Density Lipoprotein (HDL), Low Density Lipoprotein (LDL) |
| LIPID PROFILE (NON-FASTING) |
| Cholesterol, Triglycerides, High Density Lipoprotein (HDL), Low Density Lipoprotein |
| LIVER/ RENAL / BONE PROFILE |
| Sodium, Potassium, Chloride, Urea, Creatinine, Urate, Calcium, Phosphate, Total Bilirubin, Direct Bilirubin, Alkaline Phosphatase (ALP), Alanine Aminotransferase (ALT), Aspartate Aminotransferase (AST), Gamma-Glutamyl Transferase (GGT), Total Protein, Albumin, Globulin, Magnesium (Mg) |
| LIVER FUNCTION TESTS |
| Total Bilirubin, Direct Bilirubin, Alkaline Phosphatase (ALP), Alanine Aminotransferase (ALT), Aspartate Aminotransferase (AST), Gamma-Glutamyl Transferase (GGT), Total Protein, Albumin, Globulin |
| PATIENT PROFILE (NON FASTING) |
| Sodium, Potassium, Chloride, Urea, Creatinine, Urate, Calcium, Phosphate, Cholesterol, Total Bilirubin, Direct Bilirubin, Alkaline Phosphatase (ALP), Alanine Aminotransferase (ALT), Aspartate Aminotransferase (AST), Gamma-Glutamyl Transferase (GGT), Total Protein, Albumin, Globulin, Magnesium (Mg) |
| PATIENT PROFILE (+LPP - FASTING) |
| Sodium, Potassium, Chloride, Urea, Creatinine, Urate, Calcium, Phosphate, Cholesterol, Triglycerides, High Density Lipoprotein (HDL), Low Density Lipoprotein (LDL), Total Bilirubin, Direct Bilirubin, Alkaline Phosphatase (ALP), Alanine Aminotransferase (ALT), Aspartate Aminotransferase (AST), Gamma-Glutamyl Transferase (GGT), Total Protein, Albumin, Globulin, Magnesium (Mg) |

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| |
|--|
| RENAL / BONE PROFILE |
| Sodium, Potassium, Chloride, Urea, Creatinine, Urate, Calcium, Albumin, Phosphate, Alkaline Phosphatase (ALP), Total Protein, Globulin, Magnesium (Mg) |
| UREA / ELECTROLYTES / CREATININE |
| Sodium, Potassium, Chloride, Urea, Creatinine |
| UREA / ELECTROLYTES / CREATININE & LIVER |
| Sodium, Potassium, Chloride, Urea, Creatinine, Total Bilirubin, Direct Bilirubin, Alkaline Phosphatase (ALP), Alanine Aminotransferase (ALT), Aspartate Aminotransferase (AST), Gamma-Glutamyl Transferase (GGT), Total Protein, Albumin, Globulin |

Individual Tests

- Ammonia
- Amylase
- hCG Serum
- Blood Gases (including Lactate if required if sample < 15 mins)
(Venous, Arterial, Angiography)
- BNP (B-Type Natriuretic Peptide)
- CK-total
- C – Reactive Protein (CRP)
- EGFR (Calculation for Chemotherapy Only – Weight and Height required))
- Gentamicin (Pre, Post, Random)
- Glucose (Fasting)
- Glucose (Non-Fasting)
- Glucose 2 hours post (Prandial)
- Iron (Fe) and Transferrin.
- Lactate (Adult and Paediatric)
- Lactate Dehydrogenase (LDH)
- Osmolality (Serum)
- Troponin-I
- Vancomycin (Pre and Post)
- Tobramycin
- Occupational Blood/ Body Fluid Exposure

CSF

- Protein
- Glucose
- Globulin
- Note: Xanthochromia – special bottle and form required. – Assayed in Biochemistry, CUH and is not available from them during the on-call period

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Urine

- Urinary Na, K, Osmolality

Fluids

Please state source of fluid as tests performed are specific to collection location.
See section 9.7 of this document for details.

- Amylase (Fluid)
- Albumin (Fluid)
- Alkaline Phosphatase (Fluid)
- LDH (Fluid)
- pH in Pleural Fluid
- Creatinine (Fluid)
- Protein Total (Fluid)
- Glucose (Fluid)
- Urate (Fluid)
- Triglyceride (Fluid)

BLOOD BANK

- Group and Crossmatch
- Group and Screen (Antibody Screen if required)
- Direct Antiglobulin Test
- Issue of Blood Products
 1. Albumin 5% / 20%
 2. PCC
 3. FEIBA
 4. Fibrinogen Concentrate
 5. Anti-D Immunoglobulin
- Platelets (Pooled or Single)
- Solvent Detergent Plasma (SDP)
- Red Cells
- Transfusion Reaction Investigation**

HAEMATOLOGY

Profiles

| COAGULATION SCREEN |
|--|
| Prothrombin Time (PT) |
| International Normalised Ratio (INR) |
| Activated Partial Thromboplastin Time (APTT) |

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Individual Tests

- Full Blood Count (FBC) including Blood Film Review for Manual WBC Differential and Red Cell Morphology
- Activated Partial Thromboplastin Time (APTT)
- D-Dimer
- Fibrinogen
- International Normalised Ratio (INR)
- Prothrombin Time (PT)
- Infectious Mononucleosis Screening Test (Monospot)
- Malaria Parasites **
- Urinary hCG (Pregnancy Test)
- Erythrocyte Sedimentation Rate (ESR) (performed on-call at the request of a Consultant Haematologist only)

** Haematology/ Blood Bank staff member must be called in to perform this test.

MICROBIOLOGY

- Blood Cultures for Adult
- Blood Cultures for Babies and Children
- Cerebrospinal Fluid for Microscopy, Culture and Susceptibility
- Fluid from Sterile Site for Culture and Susceptibility (Synovial (Joint) Fluid)
- Urine for Microscopy, Culture and Sensitivities
- Urine Bilirubin
- Influenzae A and B (Test performed between 9am – 11pm 7 days a week)
- Covid 19 PCR (Test performed between 9am – 11pm 7 days a week)
- Respiratory Syncytial Virus (RSV)
- Extended Respiratory Panel

Note: Please refer to this document for test specific information.

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26.0 TEST INDEX

| TEST NAME | DEPARTMENT |
|---|-------------------------------------|
| 5-Hydroxyindoleacetic Acid (5-HIAA) | Biochemistry |
| 11 Deoxycortisol | Biochemistry |
| 17 Hydroxy Progesterone | Biochemistry |
| Acetyl Cholinesterase Receptor Antibodies | Biochemistry |
| Abatacept (Orencia) | Biochemistry |
| Aciclovir Pre and Post Levels | Microbiology |
| Acid Fast Bacilli Culture (AFB) | Microbiology |
| Activated Clotted Time (ACT) | Point of Care/ Near Patient Testing |
| Activated Partial Thromboplastin Time (APTT) | Haematology |
| Acyl Carnitine Profile | Biochemistry |
| Adalimumab (Humira) | Biochemistry |
| ADAMTS 13 Factor Willibrand Cleavage Protease | Haematology |
| Adenovirus Antibodies | Microbiology |
| Adrenal Antibody | Immunology |
| Adrenocorticotrophic Hormone (ACTH) | Biochemistry |
| Admission Profile | Biochemistry |
| Albumin | Biochemistry |
| Albumin (Blood Product) | Blood Bank |
| Albumin (Fluid) | Biochemistry |
| Albumin (Urinary) Microalbumin | Biochemistry |
| Albumin Creatinine Ratio | Biochemistry |
| Alcohol | Biochemistry |
| Aldolase | Biochemistry |
| Aldosterone | Biochemistry |
| Alkaline Phosphatase | Biochemistry |
| Alkaline Phosphatase (Placental) | Biochemistry |
| Alkaline Phosphatase (Isoenzymes) | Biochemistry |
| Allergen Screening | Biochemistry |
| Alpha-Aminoadipic Semialdehyde (AASA) | Biochemistry |
| Alpha 1 Antitrypsin | Biochemistry |
| Alpha 1 Antitrypsin in Faeces | Biochemistry |
| Alpha 1 Antitrypsin Genotype | Biochemistry |
| Alpha 1 Antitrypsin phenotype | Biochemistry |

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| TEST NAME | DEPARTMENT |
|--|----------------|
| Alpha Galactosidase (See Fabry Disease Screen) | Biochemistry |
| Alpha Feto Protein (AFP) | Biochemistry |
| Alanine Amino Transferase (ALT) | Biochemistry |
| Amino Acids | Biochemistry |
| AML Cytogenetic Panel | Haematology |
| Ammonia | Biochemistry |
| Amoebiasis Antibodies | Microbiology |
| Amylase | Biochemistry |
| Amylase (Fluid) | Biochemistry |
| Amylase (Urinary) | Biochemistry |
| Amyloid A Protein | Biochemistry |
| Androstenedione | Biochemistry |
| Angelman Syndrome | Histopathology |
| Angiotensin Converting Enzyme (ACE) | Biochemistry |
| Anti D Immunoglobulin (Blood Product) | Blood Bank |
| Anti Diuretic Hormone (ADH) | Biochemistry |
| Anti -GAD Antibodies | Biochemistry |
| Anti GM1 | Immunology |
| Anti-Mullarian Hormone (AMH) | Biochemistry |
| Anti-Neutrophil Cytoplasmic Autoantibodies (ANCA) | Immunology |
| Antinuclear Antibody | Immunology |
| Anti Streptolysin – O Titre (ASOT) | Microbiology |
| Anti -tyrosine Antibodies | Biochemistry |
| Antibiotic Assay Aminoglycosides including - Gentamycin - Amikacin - Tobramycin Glycopeptides including - Vancomycin - Teicoplanin | Microbiology |
| Apolipoprotein A | Biochemistry |
| Apolipoprotein B | Biochemistry |
| Apolipoprotein E (Apo E) | Biochemistry |
| APO E Lipoprotein Genotyping | Biochemistry |
| Aquaporin 4 Antibody | Immunology |
| Arsenic | Biochemistry |
| Aspergillus Antibodies | Microbiology |

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| TEST NAME | DEPARTMENT |
|---|-------------------------------------|
| Aspergillus Antigen (Galactomannan (GM)) | Microbiology |
| Aspartate Amino Transferase (AST) | Biochemistry |
| Atypical Pneumoniae Screen | Microbiology |
| Autoantibody Screen (Anti – Nuclear, Mitochondrial, LKM, Smooth Muscle, Gastric Parietal Cell Antibody) | Immunology |
| Avian Precipitins | Immunology |
| Bacterial PCR | Microbiology |
| Bartonella henselae Antibodies (Catscratch Fever) | Microbiology |
| BCR/ABL Fusion Gene | Haematology |
| BCR-ABL Transcripts (Quantitation) | Haematology |
| Beta 2 Microglobulin | Biochemistry |
| Beta 2 Transferrin | Biochemistry |
| Beta Glucan Antigen | Microbiology |
| Bicarbonate | Biochemistry |
| Biliary Brush Sample | Histopathology |
| Bilirubin Direct | Biochemistry |
| Bilirubin Total | Biochemistry |
| Biologics Profile | Microbiology |
| Biotinidase | Biochemistry |
| Bisoprolol | Biochemistry |
| Blastomyces Antibodies | Microbiology |
| Blood Cultures | Microbiology |
| Blood Gases (Venous, Arterial, Angiography) | Biochemistry |
| Blood Gas CCU | Point of Care/ Near Patient Testing |
| Blood Gas CCU Venous Including Potassium and Lactate | Point of Care/ Near Patient Testing |
| BNP B-Type Natriuretic Peptide | Biochemistry |
| Bone Marrow Aspirate | Haematology |
| Bone Marrow FISH | Haematology |
| Bordetella pertussis anti-toxin Antibodies | Microbiology |
| Breath Test for Helicobacter pylori | Point of Care/ Near Patient Testing |
| Brucella Titre | Microbiology |
| Bullous Antibodies | Immunology |
| CA15-3 | Biochemistry |
| CA19-9 | Biochemistry |

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| TEST NAME | DEPARTMENT |
|--|----------------|
| CA-50 | Biochemistry |
| CA72-4 | Biochemistry |
| CA125 | Biochemistry |
| Cadmium | Biochemistry |
| Caffeine | Biochemistry |
| Calcitonin | Biochemistry |
| Calcium | Biochemistry |
| Calcium (Urinary) | Biochemistry |
| Calcium (Ionised) | Biochemistry |
| Calcium Creatinine Ratio in Urine | Biochemistry |
| Calculi Analyses (Kidney Stone, Gall Stone) | Biochemistry |
| CALR Mutation (Flow Cytometry) | Haematology |
| Campylobacter Antibodies | Microbiology |
| Carbamazepine | Biochemistry |
| Carbohydrate Deficient Transferrin | Biochemistry |
| Carcinoembryonic Antigen (CEA) | Biochemistry |
| Cardiolipin IgG Antibodies (Anti Phospholipid Antibodies) | Histopathology |
| Carnitine (Total) | Biochemistry |
| Carnitine (Urine) | Biochemistry |
| CASPR2 Antibody | Immunology |
| Catecholamines (Blood) | Biochemistry |
| Catecholamines and Metanephrines (24 hr urine collection) | Biochemistry |
| CD4, CD8 (T4, T8) T Cell Lymphocyte Subset Levels | Haematology |
| CD34 (Marker) | Haematology |
| Cerebrospinal Fluid for Cryptococcus | Microbiology |
| Cerebrospinal Fluid for Microscopy, Culture and Susceptibility | Microbiology |
| Ceruloplasmin | Biochemistry |
| Chikungunya Serology | Microbiology |
| Children's Ward Biochemistry Profile | Biochemistry |
| Chlamydia pneumoniae IgM | Microbiology |
| Chlamydia sp. Specific Antibodies | Microbiology |
| Chlamydia trachomatis/Gonorrhoea/Trichomonas PCR | Microbiology |
| Chloride | Biochemistry |
| Cholesterol | Biochemistry |
| Cholesterol - HDL | Biochemistry |
| Cholesterol - LDL | Biochemistry |

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| TEST NAME | DEPARTMENT |
|--|-------------------------------------|
| Cholinesterase | Biochemistry |
| Chromium | Biochemistry |
| Chromosome Analysis on Blood (Karyotyping) | Histopathology |
| Chromosome Analysis Bone Marrow (Cytogenetics Bone Marrow) | Haematology |
| CK Isoenzymes | Biochemistry |
| CLL Prognostic Factors (TP53, 17pdel, IGVH) | Haematology |
| Clo Test for Helicobacter pylori | Point of Care/ Near Patient Testing |
| CMV IgG | Microbiology |
| CMV IgM | Microbiology |
| Coagulation Factor Assays | Haematology |
| Coagulation Screen Profile | Haematology |
| Cobalt | Biochemistry |
| Coccidioides Antibodies | Microbiology |
| Cold Agglutinins | Haematology |
| Colorectal Panel | Histopathology |
| Complement C1 Inhibitor Quantitation and Function | Biochemistry |
| Complement Function Test | Biochemistry |
| Complement C₂ | Biochemistry |
| Complement C₃ | Biochemistry |
| Complement C₄ | Biochemistry |
| Complement C₅ | Biochemistry |
| Complement Total (CH50) | Biochemistry |
| Copper | Biochemistry |
| Cortisol (as part of procedure for Dexamethasone test) | Biochemistry |
| Cortisol (as part of procedure for Synacthen test) | Biochemistry |
| Cortisol (pre synacthen) | Biochemistry |
| Cortisol (30 mins post synacthen) | Biochemistry |
| Cortisol (60 mins post synacthen) | Biochemistry |
| Cortisol (am) | Biochemistry |
| Cortisol (Midnight) | Biochemistry |
| Cortisol (pm) | Biochemistry |
| Cortisol (Random) | Biochemistry |
| Cortisol (Urinary) | Biochemistry |

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| TEST NAME | DEPARTMENT |
|--|-------------------------------------|
| Counterweight Plus Fasting Profile | Biochemistry |
| COVID 19 PCR | Microbiology |
| Coxiella burnetii (Q Fever Antibodies) | Microbiology |
| C-peptide | Biochemistry |
| C – Reactive Protein (CRP) | Biochemistry |
| CPE/ ESBL Screen | Microbiology |
| Creatine Kinase MB Fraction (CKMB) | Biochemistry |
| Creatine Kinase Total (CK-Total) | Biochemistry |
| Creatinine | Biochemistry |
| Creatine (Capillary) | Point of Care/ Near Patient Testing |
| Creatinine Clearance | Biochemistry |
| Creatinine Clearance Calculated GFR for Chemotherapy | Biochemistry |
| Creatinine (Pleural Fluid) | Biochemistry |
| Creutzfeldt-Jakob Disease | Microbiology |
| Cryoglobulins | Biochemistry |
| Cryptococcal Antigen | Microbiology |
| CSF Amino Acids | Biochemistry |
| CSF Neurotransmitters | Biochemistry |
| CSF PCR Panel | Microbiology |
| CSF Phospho-Tau Protein | Biochemistry |
| CSF Spectrophotometry for Xanthochromia | Biochemistry |
| CSF Tau A-Beta Proteins | Biochemistry |
| CTX (C-Telopeptide of Type 1 Procollagen) | Biochemistry |
| Cyclic Citrullinated Peptide Antibodies (CCP) | Immunology |
| CYFRA 21-1 | Biochemistry |
| Cystic Fibrosis DNA Studies | Biochemistry |
| Cystine | Biochemistry |
| Cytomegalovirus CMV Culture | Microbiology |
| Cytomegalovirus CMV PCR | Microbiology |
| Cytopathology Body Fluid (Various) | Histopathology |
| Cytopathology Bronchial Lavage | Histopathology |
| Cytopathology Cervical Specimen (Thin Prep) and HPV | Histopathology |

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| TEST NAME | DEPARTMENT |
|---|----------------|
| Cytopathology Cerebral Spine Fluid (CSF) | Histopathology |
| Cytopathology Fine Needle Aspirate (FNA) | Histopathology |
| Cytopathology Hepatic Brush Biopsy | Histopathology |
| Cytopathology Sputum | Histopathology |
| D-Dimer | Haematology |
| Dehydro Epiandrosterone DHEA | Biochemistry |
| Dehydro Epiandrosterone DHEA (Urinary) | Biochemistry |
| Dehydro Epiandrosterone Sulphate DHEAS | Biochemistry |
| Dengue Virus Serology | Microbiology |
| Dexamethasone Suppression Test: Cortisol | Biochemistry |
| Di George Syndrome (Karotype/ FISH) | Histopathology |
| Digoxin | Biochemistry |
| Dihydropyrimidine dehydrogenase Gene Screening | Biochemistry |
| Dihydrotestosterone DHT | Biochemistry |
| Diphtheria IgG (Immunity) | Microbiology |
| Direct Antiglobulin Test | Blood Bank |
| DOAC Level | Haematology |
| dsDNA Antibody | Immunology |
| Echinococcus Antibodies | Microbiology |
| EGFR (Calculation for Chemotherapy Only) | Biochemistry |
| Elastase | Biochemistry |
| Electrophoresis – Serum Protein Electrophoresis (SPE) | Biochemistry |
| Electrophoresis – Urine Immunoelectrophoresis | Biochemistry |
| EMA Screen for Hereditary Spherocytosis | Haematology |
| Endomysial IgA Antibodies (Confirmatory Test for tTG) | Immunology |
| Enteroviruses Culture (Faeces, CSF, Pleural Fluid, Viral Throat Swab) | Microbiology |
| Enterovirus PCR | Microbiology |
| Epstein-Barr Virus IgG | Microbiology |
| Epstein-Barr Virus IgM | Microbiology |
| Epstein-Barr Virus PCR | Microbiology |
| Erythrocyte Sedimentation Rate (ESR) | Haematology |
| Erythropoietin | Biochemistry |
| Ethosuzimide | Biochemistry |
| Extended Resp. Viruses | Microbiology |
| Extractable Nuclear Antigen (ENA) | Immunology |
| Fabry Disease Screen | Biochemistry |

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| TEST NAME | DEPARTMENT |
|--|----------------|
| Factor II Prothrombin Gene Mutation | Haematology |
| Factor V Leiden DNA Studies | Haematology |
| Factor Xa activity (Heparin Assay) | Haematology |
| Faecal Calprotectin | Biochemistry |
| Faeces For Clostridium Difficile Toxin | Microbiology |
| Faecoagules for Clostridium Difficile Antigens (GDH) Glutamate dehydrogenase | Microbiology |
| Faeces for Cryptosporidium | Microbiology |
| Faeces For Intestinal Pathogens | Microbiology |
| Faeces For Ova Cysts and Parasites | Microbiology |
| Faeces For Rotavirus and Adenovirus | Microbiology |
| Faeces Occult Blood | Microbiology |
| Familial Hypercholesterolaemia Genetics | Biochemistry |
| Farmers Lung Antibodies | Microbiology |
| Ferritin | Biochemistry |
| Fibrinogen | Haematology |
| Fibroblast Growth Factor 23 | Biochemistry |
| Filaria Serology | Microbiology |
| Flecainide | Biochemistry |
| Fluid from Sterile Site for Culture and Susceptibility | Microbiology |
| Folate (Folic Acid) | Biochemistry |
| Follicle Stimulating Hormone (FSH) | Biochemistry |
| Fragile X DNA Studies | Histopathology |
| Francisella tularensis | Microbiology |
| Free Fatty Acids | Biochemistry |
| Very Long Chain Fatty Acids (VLCFA) | Biochemistry |
| Free T₃ (Triiodothyronine) | Biochemistry |
| Free T₄ (Thyroxine) | Biochemistry |
| Frozen Section (Tissue) | Histopathology |
| Fructosamine | Biochemistry |
| Full Blood Count (FBC) | Haematology |
| Full Blood Count Including Manual Blood Film Examination | Haematology |
| Fungal Culture (Mycology) | Microbiology |
| GABA and AMPA Receptor Antibodies | Immunology |
| Gamma GT | Biochemistry |
| Ganglionic Acetylcholine Receptor Antibody | Immunology |
| Ganglioside Antibodies | Immunology |

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| TEST NAME | DEPARTMENT |
|---|-------------------------------------|
| Gastric Her2 | Histopathology |
| Gastrin | Biochemistry |
| GCH1 | Biochemistry |
| Gentamicin (Once Daily Dosing) | Biochemistry |
| Gentamicin (Multi Daily Dosing) | Biochemistry |
| GIST Panel | Histopathology |
| Gliadin Antibodies (IgA, IgG) | Immunology |
| Glomerular Basement Membranes Antibody (GBM) | Immunology |
| Globulin | Biochemistry |
| Globulin (CSF) | Biochemistry |
| Glucagon | Biochemistry |
| Glucose (Capillary) | Point of Care/ Near Patient Testing |
| Glucose (Fasting) | Biochemistry |
| Glucose (Non-Fasting) | Biochemistry |
| Glucose 2 hrs Post (Prandial) | Biochemistry |
| Glucose (CSF) | Biochemistry |
| Glucose (Fluid) | Biochemistry |
| Glucose 6 Phosphate Dehydrogenase (G6PD) | Haematology |
| Glucose Tolerance Test | Biochemistry |
| Glycine Receptor Antibody | Immuology |
| Glycosaminoglycans (Mucopolysaccharides) | Biochemistry |
| GQ1b | Immunology |
| Group and Crossmatch | Blood Bank |
| Group and Screen | Blood Bank |
| Growth Hormone | Biochemistry |
| Guthrie Card (New Born Screen) | Biochemistry |
| Haematology Profile | Haematology |
| Haemochromatosis DNA | Biochemistry |
| Haemoglobin Screening (Abnormal) | Haematology |
| Haemophilus influenzae Antibodies (Serum) | Microbiology |
| Haemophilus influenzae PCR | Microbiology |
| Haptoglobin | Biochemistry |
| HbA1c Glycosylated Haemoglobin | Biochemistry |
| hCG Total (Serum Quantitative as Tumour Marker) | Biochemistry |
| hCG Serum | Biochemistry |

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| TEST NAME | DEPARTMENT |
|---|----------------|
| Helicobacter pylori IgG Antibodies | Microbiology |
| Helicobacter pylori Culture | Microbiology |
| Heparin Induced Thrombocytopenia screen (HITs) | Haematology |
| Hepatitis A Antibodies Total | Microbiology |
| Hepatitis A IgM | Microbiology |
| Hepatitis B (VRL Markers) | Microbiology |
| Hepatitis B Core Antibodies | Microbiology |
| Hepatitis B DNA | Microbiology |
| Hepatitis B Surface Antigen (HbsAg) | Microbiology |
| Hepatitis B Surface Antibodies (Immunity Check) | Microbiology |
| Hepatitis C Antibodies | Microbiology |
| Hepatitis C PCR (Polymerase Chain Reaction) | Microbiology |
| Hepatitis D Antibodies | Microbiology |
| Hepatitis E Antibodies | Microbiology |
| Her 2 FISH Analysis on Breast Carcinoma | Histopathology |
| Hereditary Transthyretin Mediated Amyloidosis Gene (h ATTR) | Biochemistry |
| Herpes simplex Antibodies | Microbiology |
| Herpes simplex PCR | Microbiology |
| Histamine (Serum) | Biochemistry |
| Histamine (Urine) | Biochemistry |
| Histopathology Specimens | Histopathology |
| Histoplasma Antibodies | Microbiology |
| HIV PCR | Microbiology |
| HIV Antigen/ Antibody Test | Microbiology |
| Homocysteine | Biochemistry |
| Homogentistic Acid | Biochemistry |
| HTLV 1 and 2 | Microbiology |
| Human Herpes Virus 6 | Microbiology |
| Human Leucocyte Antigen (HLA) Typing | Haematology |
| Human Leucocyte Antigen (HLA) Antibodies | Haematology |
| Huntington's Disease | Histopathology |
| Hydroxyproline | Biochemistry |
| Hypocretin (Orexin) | Biochemistry |
| IgA Immunoglobulin A | Biochemistry |

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| TEST NAME | DEPARTMENT |
|--|-------------------------------------|
| IgA Subclasses | Biochemistry |
| IgD Immunoglobulin D | Biochemistry |
| IgE Immunoglobulin E | Biochemistry |
| IgG Immunoglobulin G | Biochemistry |
| IgG Immunoglobulin G Subclasses | Biochemistry |
| IgM Immunoglobulin M | Biochemistry |
| Immune Complexes | Biochemistry |
| Immunoglobulin Gene Mutation (IgVH Mutation) | Haematology |
| Immunoreactive Trypsin | Biochemistry |
| Infectious Mononucleosis Screening Test (Mono Spot) | Haematology |
| Inflammatory Bowel Disease (IBD Profile) | Microbiology |
| Infliximab Antibodies | Biochemistry |
| Influenzae A and B | Microbiology |
| Inhibin B | Biochemistry |
| Inhibitor Screen | Haematology |
| Insulin | Biochemistry |
| Insulin C-peptide | Biochemistry |
| Insulin Like Growth Factor Binding Protein 3 (IGF BP3) | Biochemistry |
| Insulin Like Growth Factor 1 (IGF1) | Biochemistry |
| Insulin Like Growth Factor 2 (IGF2) | Biochemistry |
| Interleukin-6 | Biochemistry |
| International Normalised Ratio (INR) | Haematology |
| Intrinsic Factor Antibody | Immunology |
| Iron (Fe) | Biochemistry |
| Ischaemic Forearm Exercise Test (Dynamic): Lactate | Biochemistry |
| Islet Cells Antibody | Immunology |
| JAK 2 | Haematology |
| Kappa Lambda Ratio | Biochemistry |
| Ketone (Capillary) | Point of Care/ Near Patient Testing |
| Lactate | Biochemistry |
| Lactate Dehydrogenase (LDH) | Biochemistry |
| Lactate Dehydrogenase Fluid | Biochemistry |
| Lactate Dehydrogenase Isoenzymes | Biochemistry |
| Lamotrigine | Biochemistry |
| Laxative Screen | Biochemistry |
| Lead | Biochemistry |

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| TEST NAME | DEPARTMENT |
|---|----------------|
| Legionella pneumophila Urinary Antigen Test | Microbiology |
| Legionella Culture | Microbiology |
| Leishmania Antibodies | Microbiology |
| Leishmania Detection | Microbiology |
| Leptospira Antibodies | Microbiology |
| Leucocyte Immunopheno-Typing | Haematology |
| Leucocyte ImmunoPheno-Typing on CSF | Haematology |
| Levetiracetam (Keppra) | Biochemmistry |
| LGI1 Antibody | Immunology |
| LH/ RH Test (Dynamic): LH and FSH | Biochemistry |
| Lipase | Biochemistry |
| Lipoprotein (a) | Biochemistry |
| Lipoprotein Profile (Fasting) | Biochemistry |
| Lipid Profile (Non Fasting) | Biochemistry |
| Lithium | Biochemistry |
| Liver/ Renal/ Bone Profile | Biochemistry |
| Liver Function Test Profile | Biochemistry |
| Lung Panel | Histopathology |
| Lupus Anticoagulant | Haematology |
| Lupus Anticoagulant Profile | Haematology |
| Lutinising Hormone (LH) | Biochemistry |
| Lymes Disease (Borrelia Burgdorferi Antibodies) | Microbiology |
| Lymphoma Viral Screen | Biochemistry |
| Magnesium | Biochemistry |
| Magnesium (Urine) | Biochemistry |
| Malaria Parasites | Haematology |
| Manganese | Biochemistry |
| Mantoux Testing | Pathology |
| Maternally Inherited Diabetes and Deafness Genetic Screen | Biochemistry |
| MDSNGS Panel | Haematology |
| Measles IgG | Microbiology |
| Measles IgM | Microbiology |

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| TEST NAME | DEPARTMENT |
|--|----------------|
| Measles RNA PCR | Microbiology |
| Melanoma Panel | Histopathology |
| MEN Type 2 Genetics | Biochemistry |
| Mercury | Biochemistry |
| Metabolic Screen - Childrens Ward Amino Acids (Blood) Amino Acids (Urine) Organic Acids | Biochemistry |
| Metanephrines (Plasma) | Biochemistry |
| Methaemoglobin | Biochemistry |
| Methotrexate | Biochemistry |
| Methylene Tetrahydrofolate Reductase Deficiency (MTHFR) | Haematology |
| Methylhistamine | Biochemistry |
| Methyl Malonic Acid | Biochemistry |
| Microalbumin Creatinine Ratio | Biochemistry |
| Microsatellite Instability | Histopathology |
| Mitotane | Biochemistry |
| MODY Genetics | Biochemistry |
| Molybdenum | Biochemistry |
| Monomeric Prolactin | Biochemistry |
| MPL Mutation Studies (MPLS) | Haematology |
| MRSA Screen | Microbiology |
| Mumps IgG, IgM | Microbiology |
| MUSK Antibody | Immunology |
| Myelin Associated Glycoprotein Antibody (MAG) | Immunology |
| Myelin Oligodendrocyte Glycoprotein Antibody (MOG) | Immunology |
| Myostis Marker Antibodies (Anti Synthetase Antibodies) | Immunology |
| Mycobacterium TB PCR | Microbiology |
| Mycology, Fungal Culture | Microbiology |
| Mycoplasma pneumoniae IgM | Microbiology |
| Myoglobin | Biochemistry |
| Myotonic Dystrophy | Histopathology |
| Neisseria meningitidis Antibodies | Microbiology |
| Neisseria meningitidis PCR | Microbiology |
| Neuroblastoma screen | Biochemistry |

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| TEST NAME | DEPARTMENT |
|---|----------------|
| Neuronal Antibodies | Immunology |
| NMDA Receptor Antibody | Immunology |
| Norovirus (Previously SRSV, Winter Vomiting Bug) | Microbiology |
| N-telopeptide (NTx) | Biochemistry |
| Occupational Blood/ Body Fluid Exposure | Microbiology |
| Occupational Health Drugs of Abuse Screen | Biochemistry |
| Oestradiol | Biochemistry |
| Oligoclonal banding | Biochemistry |
| Oncomine | Histopathology |
| Organic Acids | Biochemistry |
| Osmolality | Biochemistry |
| Osteocalcin | Biochemistry |
| Ovarian Antibody | Immunology |
| Oxalate | Biochemistry |
| Oxalate/ Creatinine Ratio | Biochemistry |
| Pancreatic Polypeptide (PPP) | Biochemistry |
| Paracetamol | Biochemistry |
| Parainfluenzae virus Immunofluorescence | Microbiology |
| Parathyroid Hormone (PTH) | Biochemistry |
| Parathyroid Hormone Related Protein | Biochemistry |
| Parvovirus B19 IgG (Non Acute) Serology | Microbiology |
| Parvovirus B19 IgM (Acute) Serology | Microbiology |
| Pathology Profiles | Pathology |
| Patient Profile (Non Fasting) | Biochemistry |
| Patient Profile (Fasting) | Biochemistry |
| Perampanel (Fycompa) | Biochemistry |
| pH in Pleural Fluid | Biochemistry |
| Pharmacy Culture | Microbiology |
| Phenobarbital (Phenobarbitone) | Biochemistry |
| Phenytoin (Epanutin) | Biochemistry |
| Phosphate (Inorganic) | Biochemistry |
| Phosphate (Urinary) | Biochemistry |
| Phospholipid Antibodies (IgG and IgM Cardiolipin and β2 Glycoprotein Antibodies) | Immunology |
| Phytanic Acid | Biochemistry |
| Platelet Antibodies | Haematology |

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| TEST NAME | DEPARTMENT |
|---|----------------|
| Platelet Function Assessment (PFA100) | Haematology |
| Platelet Genotyping by DNA | Haematology |
| Platelets (Blood Component) | Blood Bank |
| Pneumococcal Antibodies (IgG) | Microbiology |
| Pneumococcal Urinary Antigen | Microbiology |
| Pneumocystis carinii PCR (PCP) | Microbiology |
| PNH (Flow Cytometry) | Haematology |
| PNPO Pyrimidine 5' Phosphate Oxidase | Biochemistry |
| Polyoma BK Virus | Microbiology |
| Polyoma Virus (JC) | Microbiology |
| Porphyrins (Blood) | Biochemistry |
| Porphyrins (Faeces) | Biochemistry |
| Porphyrins (Plasma) | Biochemistry |
| Porphyrins (Urine) | Biochemistry |
| Potassium (K) | Biochemistry |
| Prader Willi Syndrome (Karotype/ FISH) | Histopathology |
| Prealbumin | Biochemistry |
| Pregnenetriol | Biochemistry |
| Procalcitonin | Microbiology |
| Procollagen 3 | Biochemistry |
| Progesterone | Biochemistry |
| Proinsulin | Biochemistry |
| Prolactin | Biochemistry |
| Prostate Specific Antigen (PSA) | Biochemistry |
| Prostate Specific Antigen Free (PSA) | Biochemistry |
| Protein (Urinary) | Biochemistry |
| Protein Creatinine Ratio | Biochemistry |
| Protein Total | Biochemistry |
| Protein Total (CSF) | Biochemistry |
| Protein Total (Fluid) | Biochemistry |
| Prothrombin Complex Concentrate (PCC Blood Component) | Blood Bank |
| Prothrombin Time (PT) | Haematology |
| PTH-Related Protein | Biochemistry |
| Pyruvate | Biochemistry |
| Pyruvate Kinase | Haematology |
| Quantiferon Test for MTB Complex | Microbiology |

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| TEST NAME | DEPARTMENT |
|--|----------------|
| Quinidine | Biochemistry |
| Red Cells | Blood Bank |
| Red Cell Folate | Biochemistry |
| Reagent Strip Urinalysis | Microbiology |
| Renal/ Bone Profile | Biochemistry |
| Renin | Biochemistry |
| Respiratory Syncytial Virus Antigen (RSV) | Microbiology |
| Reticulocyte Count | Haematology |
| Reverse T₃ | Biochemistry |
| Rheumatoid Screen | Immunology |
| Rickettsia Antibodies | Microbiology |
| Rubella IgG (Immunity) | Microbiology |
| Rubella IgM | Microbiology |
| Saccharomyces cerevisiae Antibodies | Microbiology |
| Salicylate | Biochemistry |
| Salivary Cortisol | Biochemistry |
| Schistosomal Antibodies | Microbiology |
| Selenium | Biochemistry |
| Serotonin | Biochemistry |
| Sex Hormone Binding Globulin (SHBG) | Biochemistry |
| Sickle Cell Screen (HBS) | Haematology |
| Sodium (Na) | Biochemistry |
| Solvent Detergent Plasma (SDP) (Frozen Plasma) (Blood Component) | Blood Bank |
| Somatostatin | Biochemistry |
| Sperm Antibodies | Biochemistry |
| Sputum | Histopathology |
| Sputum for Culture and Sensitivities | Microbiology |
| Bronchial Lavage for culture and sensitivity. | |
| Strep. pneumoniae PCR | Microbiology |
| Strongyloides Antibodies | Microbiology |
| Sulphonylureas | Biochemistry |
| Swabs for Culture and Sensitivity including ear, eye, mouth, throat, nasal, High vaginal swab (HVS), penile, cervical, urethral, wound and ulcer | Microbiology |
| Swabs for Whooping Cough (Bordetella pertussis) | Microbiology |

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| TEST NAME | DEPARTMENT |
|---|--------------|
| Synacthen Test (Dynamic): Cortisol | Biochemistry |
| Tacrolimus (Prograf) | Biochemistry |
| T & B Lymphocytes | Haematology |
| T Cell Lymphocyte (T4, T8) Subset Levels (CD4, CD8) | Haematology |
| Testosterone with SHBG | Biochemistry |
| Testosterone (Free) | Biochemistry |
| Testosterone (Urinary) | Biochemistry |
| Tetanus IgG (Immunity) | Microbiology |
| Theophylline (Aminophylline) | Biochemistry |
| Thiopurine-S-Methyl Transferase (Red Cell) (TPMT) | Biochemistry |
| Thrombophilia Screen | Haematology |
| Thrombophilia Screen Profile | Haematology |
| Thyroglobulin | Biochemistry |
| Thyroglobulin Antibody | Immunology |
| Thyroid Function Test Profile | Biochemistry |
| Thyroid Peroxidase Antibody (TPO) | Immunology |
| Thyroid Stimulating Hormone (TSH) | Biochemistry |
| Thyroid Stimulating Hormone Receptor Antibody | Immunology |
| Tips for Culture and Sensitivity | Microbiology |
| Tissue for Culture and Sensitivity | Microbiology |
| Tissue Transglutaminase IgA Antibody, Coeliac Disease (TTG) | Immunology |
| Tobramycin | Biochemistry |
| Total Syphilis Antibodies | Microbiology |
| Toxicology Screen | Biochemistry |
| Toxocara Antibodies | Microbiology |
| Toxoplasma IgM | Microbiology |
| Toxoplasma IgG | Microbiology |
| Transferrin | Biochemistry |
| Transferrin Glycoform Analysis | Biochemistry |
| Transferrin Saturation (Calculation) | Biochemistry |
| Transfusion Reaction Investigation | Blood Bank |
| TRH Test (Dynamic) Tests Measured: Free T₄, TSH, Prl | Biochemistry |
| Trichinosis Antibodies | Microbiology |
| Triglycerides | Biochemistry |
| Troponin-I | Biochemistry |
| Trypsin | Biochemistry |

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| TEST NAME | DEPARTMENT |
|---|-------------------------------------|
| UGT1A1 (Gilberts Syndrome) | Biochemistry |
| Urate (Uric acid) | Biochemistry |
| Urate (Urinary) | Biochemistry |
| Urea | Biochemistry |
| Urea/ Electrolytes/ Creatinine Profile | Biochemistry |
| Urea/ Electrolytes/ Creatinine/ Liver Profile | Biochemistry |
| Urinalysis (4 Panel) | Point of Care/ Near Patient Testing |
| Urinalysis (10 Panel) | Point of Care/ Near Patient Testing |
| Urinary Cotinine | Biochemistry |
| Urinary Electrolytes (24 Hour Collection) (Na, K, Cl, Urea) | Biochemistry |
| Urinary Electrolytes (Spot) (Na, K, Cl, Urea) | Biochemistry |
| Urinary hCG (Pregnancy Test) | Haematology |
| Urinary hCG (Pregnancy Test) | Point of Care/ Near Patient Testing |
| Urinary Lipase | Biochemistry |
| Urinary Na, K, Osmolality | Biochemistry |
| Urine Bilirubin | Microbiology |
| Urine for Microscopy, Culture and Sensitivities | Microbiology |
| Urine for Pneumococcal Antigen | Microbiology |
| Urine Purine and Pyrimidine Screen | Biochemistry |
| Valproic Acid | Biochemistry |
| Vancomycin | Biochemistry |
| Varicella zoster DNA | Microbiology |
| Varicella zoster IgG Antibodies | Microbiology |
| Vasoactive Intestinal Peptide VIP | Biochemistry |
| Vedolizumab Antibodies | Biochemistry |
| Vigabatrin | Biochemistry |
| Viral Culture | Microbiology |
| Viscosity | Biochemistry |
| Vitamin A (Retinol) | Biochemistry |
| Vitamin B₁ (Thiamine) | Biochemistry |
| Vitamin B₆ (Pyridoxal Phosphate) | Biochemistry |
| Vitamin B₁₂ | Biochemistry |
| Vitamin C (Ascorbic Acid) | Biochemistry |
| Vitamin D | Biochemistry |
| Vitamin D₃ | Biochemistry |

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| TEST NAME | DEPARTMENT |
|--|--------------|
| Vitamin E (Tocopherol) | Biochemistry |
| Vitamin K (Phylloquinone) | Biochemistry |
| Voltage Gated Calcium Channel Antibody | Immunology |
| Von Willebrands Screen | Haematology |
| VRE Screen | Microbiology |
| Water Deprivation Test | Biochemistry |
| West Nile Virus IgM | Microbiology |
| Xylose | Biochemistry |
| Zinc | Biochemistry |